

1 UNIVERSIDADE ESTADUAL DO NORTE FLUMINENSE DARCY RIBEIRO - UENF

2

3

4

5

6

7

8

9

JOAQUIM BARBOSA LEITE JUNIOR

10

11

12

13

14

15 EFEITO DA ADMINISTRAÇÃO DE MORFINA E MK-801 NA EXPRESSÃO DE UMA  
16 RESPOSTA LOCOMOTORA CONDICIONADA E SENSIBILIZADA E NA ATIVAÇÃO DA  
17 PROTEÍNA ERK 1/2 EM ESTRUTURAS ENCEFÁLICAS RELACIONADAS À  
18 DEPENDÊNCIA QUÍMICA

19

20

21

22

23

24

25

26

27

28

29

CAMPOS DOS GOYTACAZES - RJ

30

2023

31

32

1 JOAQUIM BARBOSA LEITE JUNIOR

2

3

4

5

6

7

8 EFEITO DA ADMINISTRAÇÃO DE MORFINA E MK-801 NA EXPRESSÃO DE UMA  
9 RESPOSTA LOCOMOTORA CONDICIONADA E SENSIBILIZADA E NA ATIVAÇÃO DA  
10 PROTEÍNA ERK 1/2 EM ESTRUTURAS ENCEFÁLICAS RELACIONADAS À  
11 DEPENDÊNCIA QUÍMICA

12

13

14

15

16

17

Tese apresentada ao Centro de Ciências e Tecnologias  
Agropecuárias da Universidade Estadual do Norte  
Fluminense Darcy Ribeiro, como parte das exigências do  
Doutorado no programa de Pós-Graduação em Ciência  
Animal.

18

19

20

21

22

23

24

25

26

Orientadora: Prof<sup>ª</sup>. Dr<sup>ª</sup>. Marinete Pinheiro Carrera

27

28

29

30

31

CAMPOS DOS GOYTACAZES - RJ

32

2023

33

1 JOAQUIM BARBOSA LEITE JUNIOR

2  
3  
4  
5 EFEITO DA ADMINISTRAÇÃO DE MORFINA E MK-801 NA EXPRESSÃO DE UMA  
6 RESPOSTA LOCOMOTORA CONDICIONADA E SENSIBILIZADA E NA ATIVAÇÃO DA  
7 PROTEÍNA ERK 1/2 EM ESTRUTURAS ENCEFÁLICAS RELACIONADAS À  
8 DEPENDÊNCIA QUÍMICA

9  
10  
11  
12 Tese apresentada ao Curso de Doutorado do Programa de  
13 Pós-graduação em Ciência Animal, Centro de Ciências e  
14 Tecnologias Agropecuárias da Universidade Estadual do  
15 Norte Fluminense Darcy Ribeiro, Área de Concentração da  
16 Sanidade Animal e Psicofarmacologia, como requisito para  
17 obtenção do grau de Doutor em Ciência Animal.

18  
19  
20 Aprovada em 29 de março de 2023

21  
22  
23 BANCA EXAMINADORA

24  
25  
26  
27  
28 \_\_\_\_\_  
29 Prof<sup>ª</sup>. Dr<sup>ª</sup>. Marinete Pinheiro Carrera  
30 Universidade Estadual do Norte Fluminense Darcy Ribeiro - UENF  
31 (Orientadora)

32  
33  
34 \_\_\_\_\_  
35 Prof. Dr. Enrrico Bloise  
36 Universidade Federal de Minas Gerais - UFMG

37  
38  
39 \_\_\_\_\_  
40 Prof. Dr. João Marcos de Mello Bastos  
41 Universidade Salgado de Oliveira – UNIVERSO - Campos Dos Goytacazes

42  
43  
44 \_\_\_\_\_  
45 Prof<sup>ª</sup>. Dr<sup>ª</sup>. Rosemary Bastos  
46 Universidade Estadual do Norte Fluminense Darcy Ribeiro – UENF

## AGRADECIMENTOS

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37

Agradeço primeiramente a Deus, manter-me firme e sereno na maior parte dos momentos;  
Agradeço à minha família, ao meu pai, Joaquim, à minha mãe Carla, ao meu irmão Jhean, o amor e apoio incondicionais;  
Ao meu avô, Sílvio e vó Morena, saudosos, que sempre adubaram as esperanças para o meu futuro;  
À minha Tia Dedé, todo o amor e apoio na minha caminhada desde sempre;  
Agradeço à Gizele, toda compreensão, carinho e paciência;  
Agradeço à Professora, Marinete Carrera, suas formação, confiança, atenção e oportunidade, bem como seus conhecimento e apoio;  
Agradeço à UENF, a oportunidade e toda a estrutura;  
Agradeço à Pós-Graduação em Ciência Animal, o apoio e oportunidade;  
Agradeço a todos os membros do nosso grupo do Laboratório de Farmacologia, pela amizade, os conhecimentos e a atenção;  
Agradeço à Professora Fernanda, o apoio e ajuda com os fármacos usados no experimento;  
Agradeço a todos os meus amigos da Graduação e da Pós-graduação a amizade;  
Agradeço aos amigos da música, em especial Meristema, Evokes, Tubarão Martelo, BluesCana, Undreffect, Ed Gomes e Diego Black;  
Agradeço à CAPES, todo o apoio financeiro;  
Agradeço ao João Marcos, a parceria na execução dos experimentos;  
Agradeço ao Gustavo Crespo, a parceria e confiança em todos os trabalhos;  
Agradeço à Victoria Benazio, a parceria, o apoio e a confiança;  
Agradeço ao Breno Garrone, a capacidade de me fazer entender os conceitos;  
Agradeço ao Professor Richard, a ajuda com materiais usados nos experimentos;  
Agradeço aos Professores, membros da Banca Examinadora, João Marcos de Mello Bastos, Enrrico Bloise e Rosemary Bastos a contribuição e conhecimentos.

## RESUMO

Joaquim Barbosa Leite Junior, Médico Veterinário, filho de Joaquim Barbosa Leite e Carla Aparecida Torres Leite e irmão de Jhean Torres Leite, nasceu na cidade de Bom Jesus do Itabapoana – RJ, em 15 de junho de 1990. Morou em Carabuçu (4º distrito de Bom Jesus do Itabapoana) até 2009. Ingressou na Universidade Estadual do Norte Fluminense - RJ em 2010. Desde 2010 mora em Campos dos Goytacazes.

## RESUMO

A dependência química é considerada uma doença grave, que evolui de forma complexa, crônica e progressiva. Sendo um problema de saúde pública, que se caracteriza pela tendência constante à recaída, mesmo após um longo período de abstinência. A morfina é um analgésico opioide para dores intensas e possui expressiva taxa de abuso nos últimos anos. Em altas doses a morfina provoca ativação das vias dopaminérgica e glutamatérgica e atua causando hiperlocomoção. O maleato de dizocilpina (MK-801), é um antagonista glutamatérgico dos receptores do tipo NMDA (N-metil-D-aspartato), em baixas, doses produz diminuição da locomoção e, em altas doses, causa hiperlocomoção. A dopamina é o neurotransmissor associado como o prazer e a recompensa, enquanto o glutamato se relaciona à recaída, abstinência e à memória de longo prazo. A sensibilização e o condicionamento são processos importantes para manter da dependência. A sensibilização é o processo de aumento progressivo da resposta, com a mesma dose do fármaco. O condicionamento é o processo onde um estímulo inicialmente neutro se torna um estímulo incondicionado, após constantes pareamentos. A sensibilização é mediada pela ERK (proteína quinase ativada por mitógenos), na via mesocorticolímbica (área tegmental ventral, córtex pré-frontal, Amígdala, Hipotálamo e núcleo accumbens), e está relacionada à memória de longo prazo na dependência química. O objetivo do presente trabalho foi verificar o efeito das manipulações dopaminérgicas, por meio de menor atividade dopaminérgica com apomorfina (0,05 mg/kg) ou maior ativação dopaminérgica com o uso de morfina (10 mg/kg) e MK-801 (0,025, 0,1 e 1,0 mg/kg) em um protocolo de condicionamento pavloviano de atraso, avaliando a resposta locomotora condicionada e sensibilizada e ativação da ERK. Para tanto, foram desenvolvidos 2 conjuntos experimentais. O primeiro conjunto experimental testou o efeito de doses baixas de apomorfina (0,05 mg/kg) na atenuação/bloqueio da aquisição de sensibilização locomotora e condicionamento por morfina (10 mg/kg). O segundo conjunto experimental examinou os efeitos de diferentes doses de MK-801 (0,025, 0,1 e 1,0 mg/kg) na aquisição de sensibilização locomotora e condicionamento. Os resultados mostram aumento na locomoção após tratamentos repetidos, porém sem alteração locomotora nos tratamentos agudos tanto nos experimentos com MK-801, quanto Morfina. Os grupos tratados com morfina, apresentaram hiper locomoção, sinalizando sensibilização comportamental. No primeiro conjunto experimental, os resultados mostraram maior ativação de ERK, nas regiões da área tegmental ventral e núcleo accumbens, nos grupos tratados com Morfina. Entretanto os grupos Morfina tratados previamente com Apomorfina não tiveram maior ativação de ERK, se igualando aos grupos veículo. Apenas uma dose de apomorfina é capaz de interferir na aquisição de sensibilização locomotora, mostrada por meio de diminuição da locomoção e menor ativação de ERK. No segundo conjunto experimental, os resultados dos tratamentos com dose alta de MK-801 (1.0 mg/kg) mostraram sensibilização locomotora. Por outro lado, nos animais do grupo MK-801 (0,1 mg/kg), só foi possível observar sensibilização locomotora a partir do quinto dia de experimentação.

1 Nos tratamentos com MK-801 (0,025 mg/kg), não houve diferença na locomoção se  
2 comparados ao grupo veículo. Portanto os resultados obtidos no presente trabalho indicam  
3 que manipulações dopaminérgicas, com morfina ou MK-801, assim como a avaliação da  
4 ERK, se traduzem em um modelo mais completo para examinar o processo de  
5 condicionamento e sensibilização induzido por drogas, fornecendo informações  
6 importantes sobre os processos neurobiológicos para a compreensão das complexidades  
7 da dependência de opioides.

8  
9 **Palavras-chave:** Morfina; MK-801; Dopamina; Glutamato; ERK.

## 11 ABSTRACT

12  
13 Addiction is considered a serious disease, which evolves in a complex, chronic and  
14 progressive way. Being a public health problem, which is characterized by the constant  
15 tendency to relapse, even after a long period of abstinence. Morphine is an opioid analgesic  
16 for severe pain and has had a significant rate of abuse in recent years. In high doses,  
17 morphine activates the dopaminergic and glutamatergic pathways and acts by causing  
18 hyper locomotion. Dizocilpine maleate (MK-801) is a glutamatergic antagonist of NMDA (N-  
19 methyl-D-aspartate) receptors. In low doses, it reduces locomotion and in high doses it  
20 causes hyperlocomotion. Dopamine is the neurotransmitter associated with pleasure and  
21 reward, while glutamate is related to relapse, withdrawal and long-term memory.  
22 Sensitization and conditioning are important processes to maintain addiction. Sensitization  
23 is the process of progressively increasing response with the same drug dose. Conditioning  
24 is the process where an initially neutral stimulus becomes an unconditioned stimulus after  
25 constant pairing. Sensitization is mediated by ERK (mitogen-activated protein kinase), in  
26 the mesocorticolimbic pathway (ventral tegmental area, prefrontal cortex, amygdala,  
27 hypothalamus and nucleus accumbens), and is related to long-term memory in addiction.  
28 The objective of the present study was to verify the effect of dopaminergic manipulations,  
29 through lower dopaminergic activity with apomorphine (0.05 mg/kg) or greater  
30 dopaminergic activation with the use of morphine (10 mg/kg) and MK-801 (0.025 , 0.1 and  
31 1.0 mg/kg) in a delayed Pavlovian conditioning protocol, evaluating the conditioned and  
32 sensitized locomotor response and ERK activation. For this purpose, 2 experimental sets  
33 were developed. The first experimental set tested the effect of low doses of apomorphine  
34 (0.05 mg/kg) on attenuating/blocking the acquisition of locomotor sensitization and  
35 conditioning by morphine (10 mg/kg). The second experimental set examined the effects of  
36 different doses of MK-801 (0.025, 0.1 and 1.0 mg/kg) on the acquisition of locomotor  
37 sensitization and conditioning. The results show an increase in locomotion after repeated  
38 treatments, but without locomotor alteration in the acute treatments both in the experiments  
39 with MK-801 and Morphine. The groups treated with morphine showed hyper locomotion,  
40 signaling behavioral sensitization. In the first experimental set, the results showed greater  
41 activation of ERK, in the regions of the ventral tegmental area and nucleus accumbens, in  
42 the groups treated with Morphine. However, the morphine groups previously treated with  
43 apomorphine did not have greater ERK activation, matching the vehicle groups. Only one  
44 dose of apomorphine is capable of interfering with the acquisition of locomotor sensitization,  
45 shown by decreased locomotion and lower ERK activation. In the second experimental set,  
46 the results of treatments with a high dose of MK-801 (1.0 mg/kg) showed locomotor  
47 sensitization. On the other hand, in the animals of the MK-801 group (0.1 mg/kg), it was

1 only possible to observe locomotor sensitization from the fifth day of experimentation. In  
2 treatments with MK-801 (0.025 mg/kg) there was no difference in locomotion compared to  
3 the vehicle group. Therefore, the results obtained in the present work indicate that  
4 dopaminergic manipulations, with morphine or MK-801, as well as the ERK evaluation,  
5 translate into a more complete model to examine the drug-induced conditioning and  
6 sensitization process, providing important information about the neurobiological processes  
7 for understanding the complexities of opioid Addiction.

8

9 **Keywords:** Morphine; MK-801; Dopamine; Glutamate; ERK.

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

**SUMÁRIO**

<b>1</b>	<b>INTRODUÇÃO .....</b>	<b>10</b>
1.1	Hipótese .....	14
<b>2</b>	<b>OBJETIVOS .....</b>	<b>15</b>
2.1	Objetivos gerais .....	15
2.2	Objetivos Específicos .....	15
<b>3</b>	<b>REVISÃO DE LITERATURA .....</b>	<b>16</b>
3.1	Dependência Química .....	16
3.2	Morfina .....	18
3.3	Condicionamento Pavloviano e Morfina .....	22
3.4	Sensibilização e Morfina .....	24
3.5	MK-801, Sistema Dopaminérgico e o Sistema Glutamatérgico ...	25
3.6	Proteína Quinase Ativada Por Mitógenos – ERK .....	32
<b>4</b>	<b>MATERIAIS E MÉTODOS .....</b>	<b>34</b>
4.1	Sujeitos .....	34
4.2	Ambiente Experimental .....	34
4.3	Fármacos .....	35



4.4	Procedimento Experimental .....	35
<b>5</b>	<b>MANUSCRITOS .....</b>	<b>36</b>
5.1	Capítulo I .....	37
5.2	Capítulo II .....	81
<b>6</b>	<b>DISCUSSÃO GERAL .....</b>	<b>111</b>
<b>7</b>	<b>CONCLUSÃO .....</b>	<b>114</b>
<b>8</b>	<b>REFERÊNCIAS BIBLIOGRÁFICAS .....</b>	<b>115</b>

1

2

3

4

5

6

7

8

9

10

11

12

13

14

## 1 - INTRODUÇÃO

A dependência química ou vício, como é mais comumente determinada, é uma doença grave na qual se desenvolvem processos como a sensibilização, o condicionamento, a abstinência, a recaída e a tolerância. Trata-se de uma doença incurável induzida pelo uso de substâncias psicoativas, ademais se manifesta de forma crônica e progressiva (CUNHA e NOVAIS, 2004). Essa doença se caracteriza pela tendência constante à recaída, mesmo após um longo período de abstinência o que envolve alterações biológicas e comportamentais. A dependência e o alto risco à recaída acompanham o indivíduo por toda sua vida (WISE, 1996; O'BRIEN *et al.*, 1998; WISE, 2000). O dependente tem prejuízos cognitivos na percepção, atenção, associação, memória, raciocínio, juízo, imaginação, pensamento e linguagem que podem variar em intensidade de indivíduo para indivíduo (MANN *et al.*, 1999).

A ONU qualifica a dependência química como uma epidemia mortal, sendo o uso de opioides uma das maiores crises de saúde atuais, em todo o mundo cerca de 1068 pessoas morrem diariamente vítimas de overdose por opioides (KOLODNY *et al.*, 2015; CHANG *et al.*, 2018; UNODC, 2019). No cenário atual, os opioides já contabilizam 76% das mortes envolvendo distúrbios relacionados ao uso de drogas no mundo (UNODC, 2022). Só em 2021, ocorreram mais de 107 mil mortes por overdose de drogas nos EUA, 17% maior quando comparado ao ano de 2020 (UNODC, 2022). Nos últimos 5 anos, cerca de 80% dos usuários de heroína dos Estados Unidos buscaram a substância ilícita após fazerem uso de opioides comerciais (HUECKER E LEAMING, 2020; KIM *et al.*, 2019). Segundo dados do UNODC 2022, a produção mundial de ópio aumentou 7% entre 2020 – 2021.

Dados do Relatório Mundial sobre Drogas em relação ao uso de opioides, tanto para uso médico quanto para uso recreativo como droga de abuso, indica o Brasil como o maior mercado consumidor da América do Sul, com cerca de 600 mil usuários (UNODC, 2016). No Brasil, houve 1,6 milhões de prescrições de opioides no ano de 2009, já em 2015 foram 9 milhões de prescrições, um aumento de 465% em um período de 6 anos (KRAWCZYK *et al.*, 2018). Cabe ressaltar que os fármacos com maiores incrementos foram a codeína (alcaloide natural do ópio) e a oxicodona (alcaloide semi-sintética), ambas substâncias tem ação semelhante à da morfina (KRAWCZYK *et al.*, 2018; ALEXANDER *et al.*, 2020).

1           Nesse contexto, o desenvolvimento da dependência química ocorre devido a uma  
2 complexa interação entre fatores sociais, biológicos e genéticos (ROBBINS e EVERITT,  
3 1999). O dependente, na ausência da droga, quando interrompe o uso, sente forte  
4 motivação para recobrar o consumo, mesmo depois de decorridos meses ou até anos do  
5 último uso. A recaída é uma situação em que o dependente volta ao consumo depois de  
6 algum tempo de abstinência (ZALESKI *et al.*, 2017).

7           Atualmente se sabe que a recaída é mantida não apenas pela sensação de prazer  
8 ou ainda pela tentativa de evitar os possíveis efeitos da abstinência da droga, mas sim  
9 pelos mecanismos que envolvem aprendizagem de longa duração, ou seja, a memória  
10 associada às drogas (NESTLER, 2002). Para a dependência química, o desejo resultante  
11 da associação entre os estímulos ambientais e os efeitos subjetivos da droga (particular a  
12 cada indivíduo) estão amplamente ligados com os fatores de risco de longa duração para  
13 a recaída (HYMAN e MALENKA (2001).

14           As alterações comportamentais geradas pelo uso de drogas de forma crônica  
15 produzem adaptações neurobiológicas e moleculares permanentes, instituindo um  
16 importante modelo de neuroplasticidade (ROBINSON e KOLB, 1999; EISCH *et al.*, 2000).  
17 Dentre os processos relacionados com a gênese e manutenção da dependência, dois  
18 processos são fundamentais: o primeiro consiste em uma forma de aprendizagem  
19 associativa estabelecida entre os efeitos da droga (estímulo farmacológico) e a interação  
20 com o ambiente (objetos, lugares e som), conhecido como processo de condicionamento  
21 (SHALEV *et al.*, 2002); o segundo é uma forma de aprendizagem não associativa,  
22 denominada sensibilização comportamental, e se caracteriza pelo aumento progressivo de  
23 uma determinada resposta comportamental, quando se administra a mesma dose da droga  
24 repetidas vezes (ROBINSON e BERRIDGE, 1993). Dessa forma, o estudo do  
25 condicionamento e sensibilização comportamental é de grande importância para o  
26 entendimento dessa doença. Os dois processos explicam os comportamentos compulsivos  
27 de desejo, ingestão e recaída presentes na dependência química (WISE e ROMPRE, 1989;  
28 PIERCE e BARI, 2001; SEE, 2002).

29           Já se encontra bem aceito na literatura científica que a característica comum a todas  
30 as substâncias de abuso é a ativação do sistema dopaminérgico e glutamatérgico  
31 (SCHULTZ *et al.*, 1993; TRUDEAU *et al.*, 2014; SAKAE *et al.*, 2015). Entretanto o fator de  
32 maior importância compartilhado por essas substâncias é a transformação dos estímulos  
33 contextuais contíguos (próximos) em estímulos condicionados e estímulos de incentivo, os

1 quais podem motivar e manter os comportamentos relacionados à dependência (ROBBINS  
2 e EVERITT, 2002).

3 As catecolaminas, como a dopamina, atuam no circuito de recompensa no sistema  
4 nervoso central (SNC), e são importantes para a aquisição de memórias de longo prazo,  
5 pois variações nos seus níveis circulantes estão envolvidas na modificação do  
6 aprendizado, tornando o aprendizado prazeroso ou ainda angustiante (ORDOÑEZ, 2012).  
7 Níveis altos de dopamina e de glutamato facilitam o aprendizado, contribuindo para a  
8 consolidação de memórias mal adaptadas como é o caso da dependência química. Por  
9 outro lado, níveis baixos de dopamina podem dificultar a consolidação de memórias e  
10 assim serem usados como estratégia para o tratamento do dependente químico (NODA *et*  
11 *al.*, 1998; NODA e NABESHIMA, 2004).

12 Os sistemas, dopaminérgico bem como o glutamatérgico estão implicados na  
13 dependência por opioides e outras drogas psicoativas como o álcool e a cocaína. Nesse  
14 contexto, os estudos terapêuticos atuais se voltam para drogas, que se sejam capazes de  
15 equilibrar a transmissão do glutamato e da dopamina no contexto de dependência química  
16 (COLLINS *et al.*, 1998; OLIVE *et al.*, 2012; D'SOUZA, 2015).

17 Trabalhos de DOBI e colaboradores (2010) e WYLLIE e colaboradores (2013)  
18 mostraram diminuição da locomoção quando antagonistas glutamatérgicos NMDA como  
19 por exemplo o MK-801 (maleato de dizocilpina) foram administrados em baixas doses  
20 (0,01-0,025 mg/kg), os receptores do subtipo GLUN-2A são os mais impactados e deixam  
21 de inibir a glicina (FRANTZ e HARTESVELDT, 1999; TANG *et al.*, 2006). Essa diminuição  
22 da locomoção aponta para a importância do glutamato no processo da dependência  
23 química (TANG *et al.*, 2006).

24 A morfina é uma droga psicoativa capaz de gerar dependência química, pois é uma  
25 substância que apresenta uma característica comum com as drogas de abuso, a  
26 capacidade de aumentar os níveis circulantes de glutamato pós-sináptico (NARITA *et al.*,  
27 2008) e também de dopamina (KREEK, 2007). De acordo com os trabalhos do nosso  
28 grupo, que mostraram através de experimentos com administração de apomorfina como  
29 pré-tratamento antes da administração de morfina, houve uma menor ativação de ERK 1/2  
30 em regiões encefálicas como a área tegmental ventral (VTA) e o NAc (DE MELLO BASTOS  
31 *et al.*, 2019; LEITE JUNIOR *et al.*, 2019; DIAS *et al.*, 2021; CRESPO *et al.*, 2022), essas  
32 estruturas que são responsáveis por reforçar o aprendizado e aumentar a probabilidade de

1 se usar a droga novamente, como o NAc e VTA, são capazes de gerar e manter a  
2 dependência química (SHOBLOCK *et al.*, 2005).

3 A hipótese do presente é que o aumento da atividade da dopamina por meio dos  
4 tratamentos com MK-801 em alta dose (1.0 mg/kg) e com morfina (10 mg/kg),  
5 administrados imediatamente antes do teste experimental, provocam aumento da  
6 locomoção, como também um aumento de ERK, sugerindo aumento das respostas  
7 condicionada e sensibilizada. Bem como a diminuição da atividade dopaminérgica pelo uso  
8 de apomorfina (0.05 mg/kg), dificulta a consolidação de memórias associadas às drogas  
9 psicoativas, resultando em menor atividade locomotora e menor ativação de ERK.

10 Assim sendo, o objetivo do presente trabalho foi verificar o efeito das manipulações  
11 dopaminérgicas, por meio de menor atividade dopaminérgica com apomorfina (0,05 mg/kg)  
12 ou maior ativação dopaminérgica com o uso de morfina (10 mg/kg) e MK-801 (0,025, 0,1 e  
13 1,0 mg/kg) em um protocolo de condicionamento pavloviano de atraso, avaliando a  
14 resposta locomotora condicionada e sensibilizada e a ativação da proteína ERK. Para  
15 tanto, foram desenvolvidos 2 conjuntos experimentais. O primeiro conjunto experimental  
16 testou o efeito de doses baixas de apomorfina (0,05 mg/kg) na atenuação/bloqueio da  
17 aquisição de sensibilização locomotora e condicionamento por morfina (10 mg/kg). O  
18 segundo conjunto experimental examinou os efeitos de diferentes doses de MK-801 (0,025,  
19 0,1 e 1,0 mg/kg) na aquisição de sensibilização locomotora e condicionamento. O presente  
20 trabalho busca uma maior compreensão da interação do glutamato e da dopamina na  
21 dependência química, principalmente de drogas como as opioides.

22

23

24

25

26

27

28

29

30

## 1 1.1 HIPÓTESE

2

3 Trabalhos de colaboradores (1992) e Carey (1995), utilizando MK-801 nas doses (0,1 – 0,3  
4 mg/kg), em ratos, descreveram alguns efeitos excitatórios como a hiperatividade  
5 locomotora. Nos trabalhos do nosso grupo CRESPO e colaboradores (2022), utilizando  
6 morfina na dose (10 mg/kg), observou-se aumento da atividade locomotora e maior  
7 ativação de ERK em regiões como o VTA, Nac. Outros resultados foram encontrados por  
8 De Mello e colaboradores (2020), com o uso de injeções prévias de apomorfina (0,05  
9 mg/kg) antes de injeções de morfina (10 mg/kg), em que houve bloqueio da aquisição de  
10 sensibilização e condicionamento.

11 Assim sendo, a hipótese do presente estudo é que manipulações dopaminérgicas  
12 por meio de uma menor atividade dopaminérgica com o uso de apomorfina (0,05 mg/kg)  
13 dificultam ou bloqueiam as respostas locomotoras que indicam condicionamento e  
14 sensibilização, como a hiperlocomoção. Em outro ponto, uma manipulação onde ocorra  
15 maior ativação dopaminérgica com o uso de morfina (10 mg/kg) e MK-801 (1,0 mg/kg) em  
16 um protocolo de condicionamento pavloviano de atraso, provocam aumento na resposta  
17 locomotora condicionada e sensibilizada e também na ativação da ERK.

18

19

20

21

22

23

24

25

26

27

28

29

30

31

## 1 II - OBJETIVOS

2

### 3 **Objetivo Geral:**

4 O objetivo do presente trabalho foi verificar o efeito das manipulações  
5 dopaminérgicas, por meio de menor atividade dopaminérgica com apomorfina (0,05 mg/kg)  
6 ou maior ativação dopaminérgica com o uso de morfina (10 mg/kg) e MK-801 (0,025, 0,1 e  
7 1,0 mg/kg) em um protocolo de condicionamento pavloviano de atraso, avaliando a  
8 resposta locomotora condicionada e sensibilizada e a ativação da proteína ERK

9

### 10 **Objetivos Específicos:**

11 a) Avaliar a administração de MK-801 1.0 mg/kg no desenvolvimento de uma resposta  
12 condicionada e sensibilizada, empregando-se um protocolo de condicionamento  
13 pavloviano de atraso;

14 b) Avaliar a administração de morfina 10 mg/kg no desenvolvimento de uma resposta  
15 condicionada e sensibilizada, juntamente com a ativação da ERK 1/2, empregando-se um  
16 protocolo de condicionamento pavloviano de atraso;

17 c) Avaliar a administração de Apomorfina na dose de 0,05 mg/kg, pré-arena  
18 experimental, na atenuação e/ou bloqueio de uma resposta locomotora previamente  
19 condicionada e sensibilizada por morfina 10 mg/kg;

20 d) Avaliar a administração de Apomorfina na dose de 0,05 mg/kg antes de uma  
21 administração de morfina 10 mg/kg pré-arena experimental na atenuação e/ou bloqueio da  
22 ativação da proteína ERK nas regiões do VTA, Nac, Hipo, AM e CPF.

23

24

25

26

27

28

## 1 III - REVISÃO DE LITERATURA

2

### 3 3.1 Dependência Química

4

5 A dependência química é uma doença crônica que tem como característica a  
6 tendência recorrente à recaída e pode ser entendida dessa forma como um problema  
7 complexo e contínuo, sendo classificada entre os transtornos psiquiátricos (CID10, F19).

8 De acordo com as últimas estimativas do Relatório Mundial sobre Drogas de 2022,  
9 cerca de 284 milhões de pessoas (3,55% da população mundial) entre 15 e 64 anos  
10 usaram drogas no ano de 2020, enquanto cerca de 40 milhões de pessoas (13% da  
11 população de usuários) sofrem de transtornos associados ao uso de drogas. As projeções  
12 estatísticas sugerem um aumento de 11% no número de usuários globalmente até 2030 e  
13 um aumento acentuado de 40% na África devido ao rápido crescimento da população  
14 jovem, juntamente com um acréscimo do número de usuárias mulheres (UNODC, 2022).

15 A dependência é uma forma de aprendizado, que ocorre devido ao consumo de  
16 substâncias psicoestimulantes que provocam alterações do SNC como o aumento de  
17 níveis circulantes de dopamina, o que ocasiona uma ação estimulatória na vigília e na  
18 atenção. Dessa forma, a dependência pode ser separada em dependência física, na qual  
19 o organismo do dependente apresenta distúrbios físicos quando se faz a interrupção do  
20 uso da droga (DIAZ, 1996). A dependência ainda pode ser entendida por dependência  
21 psicológica, em que o uso tem por objetivo o bem-estar e os efeitos iniciais da droga  
22 (RIBEIRO e MINAYO, 2015).

23 A dependência pode ser tratada e controlada objetivando a diminuição dos  
24 sintomas, sendo, por vezes, de difícil controle (PÉREZ-CAJARAVILLE *et al.*, 2005). Os  
25 indivíduos toxicodependentes tendem a ter uma rejeição por atividades que não estejam  
26 ligadas ao consumo da droga, pois ocorre diminuição da sensação de prazer (NERY FILHO  
27 *et al.* 2009).

28 Por muito tempo, a dependência foi entendida como sendo desvio de caráter, falta  
29 de personalidade ou diminuição da força de vontade, pela qual o dependente era tratado  
30 por nomes pejorativos, o que levava a uma interpretação equivocada sobre o uso de drogas  
31 (DE MORAIS *et al.*, 2012). Somente em 1964 foi que a Organização Mundial da Saúde



1 (OMS) introduziu o termo “dependência” em modificação ao termo “vício” e “habituação”  
2 que eram usados até então, assim o problema passou a ser entendido como uma doença  
3 (DUPONT, 2005).

4 O tratamento da dependência química é um desafio árduo, pois suas causas são  
5 diversas, ou seja, sendo determinada por vários fatores. Para o seu entendimento se faz  
6 necessária uma abordagem cuidadosa, feita por equipes multidisciplinares, pois se refere  
7 a um problema em que se relacionam fatores sociais, familiares, emocionais e da psiquê  
8 (FONTES *et al.*, 2006). Na maior parte dos pacientes, cerca de 90%, ocorre recaída em  
9 até 1 ano depois de iniciado o tratamento, tornando o processo bastante penoso (MILTON  
10 e EVERITT, 2012).

11 O consumo de substâncias psicoestimulantes é bem antigo e de certa maneira  
12 contemporâneo ao próprio nascimento das civilizações (NUNES e JÓLLUSKIN, 2007). No  
13 princípio os homens consumiam drogas em cerimônias religiosas como parte da  
14 celebração, depois o consumo de drogas mudou de contexto e passou a ser feito com a  
15 finalidade de diminuir o sofrimento, diminuir a dor, o cansaço e também para se alcançar  
16 sensações prazerosas (MARTINS e CORRÊA, 2004).

17 Dois fatos históricos ocorridos no século XIX contribuíram bastante para que a  
18 dependência química se tornasse pela primeira vez um grave problema social, o primeiro  
19 foi a guerra travada entre China e Inglaterra pelo abastecimento mundial de ópio e o  
20 segundo foi durante a Guerra Civil Americana nos EUA (ARAÚJO e MOREIRA, 2006).  
21 Nesses dois conflitos os soldados feridos em combate fizeram o uso de morfina nas formas  
22 oral e subcutânea, e isso provocou um enorme número de casos de dependência química  
23 entre os sobreviventes que retornaram à Europa e aos EUA (BARAKA, 2000; DUARTE,  
24 2005).

25 O abuso ou uso nocivo seria um momento intermediário entre o uso recreativo (de  
26 baixo risco, não sendo caracterizado como um problema médico) e a dependência. Já há  
27 prejuízo decorrente do consumo da substância, mas ainda há algum controle do indivíduo  
28 quanto à quantidade consumida e à duração dos efeitos (TAMELINI e MARTINS, 2007).

29 Alguns pontos são importantes para entender os limites entre o uso recreativo, o  
30 abuso e a dependência de substâncias psicoativas. Postula-se que sejam fenômenos que  
31 ocorrem em conjunto, com parâmetros orientando a transição de um estágio para o outro,  
32 como: o impacto funcional no trabalho e na família, restrições judiciais e morais, as

1 consequências decorrentes do consumo da substância, assim como o desenvolvimento de  
2 mecanismos fisiológicos de adaptação à presença da substância, como tolerância e  
3 abstinência. Tolerância é a diminuição dos efeitos esperados de uma droga por exposição  
4 excessiva do dependente ao seu princípio ativo, enquanto a abstinência é a privação do  
5 uso da droga e de suas sensações prazerosas, podendo causar perturbações fisiológicas  
6 no organismo como: irritabilidade, depressão, ansiedade, suores, enjoos, dores de cabeça  
7 (RAITH e HOCHHAUS, 2004).

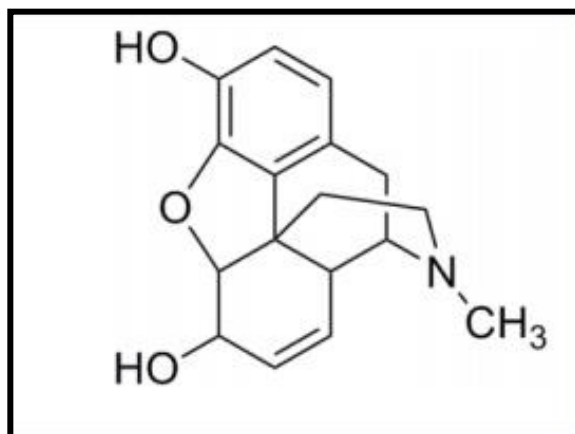
8 As drogas ativam o circuito da recompensa na via mesocorticolímbica (MORGANE  
9 *et al.*, 2005) liberando neurotransmissores responsáveis por uma sensação de bem-estar  
10 e prazer como a dopamina e o glutamato, como consequência, o desejo por repetir a  
11 experiência se mantém. Na dependência química, o comportamento de busca ativa da  
12 droga é no sentido de reequilibrar o sistema de recompensa (FERREIRA *et al.* 2001).

13

### 14 **3.2 Morfina**

15

16 A morfina é um opióide analgésico. A droga foi isolada pelo alemão Friedrich  
17 Serturmer em 1806, mas teve sua difusão somente em 1853 após a invenção da agulha  
18 hipodérmica (PATHAN e WILLIAMS, 2012). Sendo o primeiro fármaco narcótico derivado  
19 do ópio (*Papaver somniferum*) e usado frequentemente na intervenção cirúrgica e clínica  
20 de doenças com presença de dor crônica. A morfina pode ser administrada por via oral,  
21 subcutânea, intramuscular, intravenosa, epidural e ainda pelas vias transdérmica e  
22 intranasal (NESTLER, 2004). Quando usada pelas vias intramuscular e subcutânea, o pico  
23 de ação se dá aos 15-20 minutos, por via intravenosa, o pico do efeito analgésico é obtido  
24 aos 20 minutos. Caso seja administrada em bolus epidural ou intratecal, o pico ocorre de  
25 5 a 10 minutos após a administração e tem a duração da ação analgésica de 4 a 5 horas.  
26 Quando administrada por via oral, a morfina tem seu pico de ação em 30-90 minutos, com  
27 6 horas de duração da ação analgésica (GLARE e WALSH, 1991; LUGO e KERN, 2002;  
28 STEIN, 2015). A droga consegue ultrapassar a placenta e a barreira hematoencefálica e  
29 sua excreção ocorre na maior parte pela via renal, apenas uma pequena parte é excretada  
30 por via biliar. A figura 1 mostra a fórmula estrutural da morfina (SJOGREN *et al.*, 1994).



1

2 Figura 1: Fórmula estrutural da morfina (7,8-didehydro-4,5-epoxy-17-methylmorphinan-3,6-  
3 diol). Adaptado de Trescot e colaboradores (2008).

4

5 Os receptores dos opioides são classificados em 3 tipos principais:  $\mu$  (Mi),  $\kappa$  (Kappa),  
6  $\delta$  (Delta) e ainda mais 2 tipos que se chamam Nor (nociceptina), e, mais recentemente, o  
7  $\zeta$  (Zeta). Esses receptores fazem parte da família de receptores ligados à proteína G no  
8 mecanismo de analgesia. Os receptores do tipo  $\mu$  (OP3) regulam funções como a  
9 nocicepção, o controle do ciclo respiratório, miose, bradicardia, indiferença aos estímulos  
10 do meio circulante e o trânsito intestinal. São receptores onde a maior parte dos fármacos  
11 opioides atua, eles estão localizados nas lâminas III e V do córtex cerebral, no tálamo, área  
12 tegmental ventral (VTA), substância cinzenta periaquedutal, substância gelatinosa e trato  
13 gastrintestinal (PERT e SNYDER, 1973). Já os receptores do tipo  $\kappa$  (OP2) tem relação com  
14 as funções de nocicepção, termorregulação, controle de diurese e secreção  
15 neuroendócrina, eles estão localizados no hipotálamo, substância cinzenta periaquedutal,  
16 substância gelatinosa na medula espinhal, além de neurônios sensitivos periféricos  
17 (DHAWAN *et al.*, 1996). Quanto aos receptores do tipo  $\delta$  (OP1), eles são mais fortemente  
18 expressos nos gânglios da base e nas regiões do neocortex. Esse tipo de receptor tem  
19 relação com o controle do ciclo respiratório, nocicepção e modulação de funções cognitivas  
20 (BROWNSTEIN, 1993). O quarto tipo de receptor o Nor (OP4) é encontrado no córtex,  
21 amígdala, hipocampo e hipotálamo, sendo relacionado com regulação do humor e do  
22 apetite (STEIN *et al.*, 2003). Por fim, há o receptor tipo  $\zeta$  (ZOR) que tem importância para  
23 o crescimento do tecido celular e embrionário, e, ainda na regulação da proliferação de  
24 células cancerosas, esse tipo de receptor se encontra em regiões como o coração, os rins,  
25 os músculos esqueléticos, o cérebro e o pâncreas (VARGA *et al.*, 2004; MARTINS *et al.*,  
26 2012).

1 Os receptores opioides estão envolvidos em uma série de sinais intracelulares,  
2 incluindo a inibição da adenilato ciclase, a diminuição da abertura dos canais de cálcio, o  
3 aumento das correntes de potássio e a ativação da proteína quinase C (PKC). O principal  
4 efeito dos opioides é a redução da excitabilidade celular e da neurotransmissão. Em nível  
5 celular, a morfina age como agonista total nos receptores  $\mu$  acoplados a proteína G do tipo  
6 inibitória (Gi), impedindo a ação da enzima adenilato ciclase, essa enzima é responsável  
7 pela transformação de ATP (adenosina trifosfato) em AMPc (Monofosfato cíclico de  
8 adenosina), dessa forma, não serão feitas novas sínteses de AMPc (importante  
9 mensageiro intracelular); como resultado, há a diminuição das respostas celulares  
10 (PETROFF, 2002).

11 Outro ponto de destaque é o aumento do efluxo de potássio, ou seja, ocorre a  
12 passagem dos íons potássio para o exterior da célula, devido à abertura dos canais de  
13 potássio e fechamento dos canais que levam sódio para a porção interior da célula  
14 (COHEN, 1979). A diminuição da concentração de potássio no meio intracelular faz com  
15 que a membrana da célula fique hiperpolarizada, dessa forma não é gerado o potencial de  
16 ação ocorrendo o impedimento do impulso nervoso (SCHROLL e HAMKER, 2013).

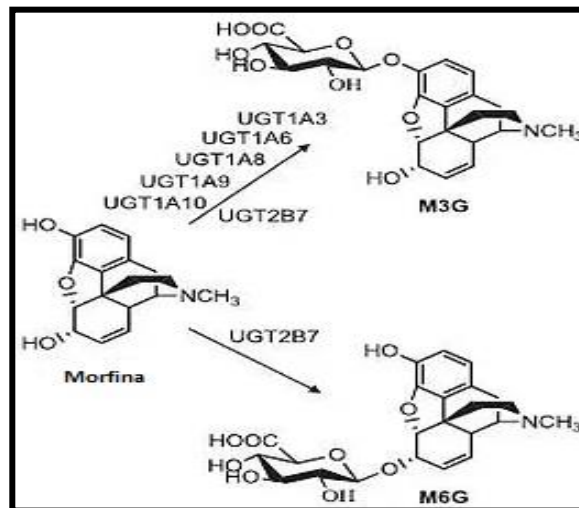
17 Em nível de SNC, a morfina tem sua ação nos receptores do tipo  $\mu$  (Mi) dos  
18 neurônios gabaérgicos, onde ocorre uma diminuição da síntese do ácido gama-  
19 aminobutírico (GABA) e posterior aumento da dopamina e glutamato circulantes devido à  
20 excitação dos neurônios dopaminérgicos e desinibição dos neurônios glutamatérgicos,  
21 posteriormente ocorre a ativação do sistema de recompensa das vias mesolímbica e  
22 mesocortical, duas vias simpáticas e também ativação das vias glutamatérgicas  
23 descendentes (via córtico-troncar, córtico-estriatal, córtico-accumbens, córtico-talâmica).  
24 Essa excitação do sistema nervoso central pelo aumento de dopamina e glutamato é o  
25 mecanismo responsável por causar a dependência química (YAMAKAGE e NAMIKI, 2002).

26 Com as diminuições da fração de mensageiros celulares e da liberação de  
27 neurotransmissores inibitórios pelas células que compõem o SNC respectivamente,  
28 somado a um aumento da liberação de dois importantes neurotransmissores excitatórios  
29 glutamato e dopamina, ocorre uma excitação neuronal com ativação do sistema de  
30 recompensa cerebral (GOLDBERG *et al.*, 2013). Com o aumento de glutamato e dopamina  
31 há uma redução da síntese de importantes neurotransmissores inibitórios, responsáveis  
32 pela regulação do humor, como a glicina e o GABA (WATANABE *et al.*, 2002).

1 Os receptores opioides estão localizados em neurônios de diversas áreas do  
2 encéfalo (córtex pré-frontal, tálamo, hipocampo, bulbo olfatório, núcleos pontinhos,  
3 substância gelatinosa na medula espinhal e na substância cinzenta periaquedutal no  
4 mesencéfalo). Além disso, podem ser encontrados no sistema nervoso periférico. A  
5 ativação desses receptores é feita pelos opióides endógenos, como as endorfinas e as  
6 encefalinas que são neurotransmissores semelhantes à morfina (HEMMINGS e  
7 JEVTOVIC-TODOROVIC, 2013).

8 A morfina atuando nos receptores  $\mu$  (Mi) tem as seguintes ações: analgesia, miose,  
9 bradicardia, depressão respiratória e indiferença aos estímulos do meio circulante. Quando  
10 sua atuação é nos receptores  $\delta$  (Delta), ocorre primariamente a analgesia, mas também a  
11 modulação de funções cognitivas e de dependência física, enquanto que sua ação nos  
12 receptores  $\kappa$  (Kappa), vai alterar a nocicepção, a termorregulação, o controle de diurese, a  
13 secreção neuroendócrina e a reduzir a atividade motora e reflexos (MARTINS *et al.*, 2012).  
14 Os receptores do tipo OP1 ou  $\sigma$  (sigma) estimulam as funções simpáticas provocando  
15 taquicardia, hipertensão, midríase, náuseas e vômitos, também podem causar  
16 alucinações. Os receptores  $\sigma$  (sigma) são parcialmente opioides, depois de reanalisados  
17 por alguns pesquisadores, foi visto que eles interagem com uma variedade de drogas  
18 psicoativas e seu ligante endógeno não é conhecido (embora eles possam reagir com  
19 certos esteroides endógenos), chegou-se à conclusão de que seriam, na verdade,  
20 decorrentes do bloqueio de receptores glutamatérgicos do tipo NMDA (PERT e SNYDER,  
21 1973; BROWNSTEIN, 1993; MARTINS *et al.*, 2012).

22 A morfina é metabolizada no fígado, essencialmente pela UDP-  
23 glucuronosiltansferase 2B7 (UGT2B7). A partir daí, são obtidos dois metabólitos principais:  
24 a morfina-6-glucuronideo (M6G) que é um metabólito ativo e responsável por efeitos  
25 clínicos da morfina como a analgesia (LÖTSCH e GEISSLINGER, 2001) e a morfina-3-  
26 glucuronideo (M3G), que em excesso pode levar a hiperalgesia (sensibilidade aumentada  
27 a dor), através da reação de fase II (glucoronidação) do grupo alcoólico 6-OH e do grupo  
28 fenólico 3-OH (figura 2); (DAHAN *et al.*, 2008; SMITH, 2000).



1

2 Figura 2: Metabolismo da morfina e os seus metabólitos M6G e M3G. Adaptado de  
 3 Wahlström *et al.* (1988).

4

### 5 3.3 Condicionamento Pavloviano e Morfina

6

7 Rehman e Rehman (2018) descrevem que o condicionamento clássico é um método  
 8 de aprendizado inconsciente, uma forma direta através da qual os animais podem  
 9 aprender. O condicionamento clássico é o processo em que uma resposta automática e  
 10 condicionada é emparelhada com estímulos específicos.

11 O condicionamento pode ser segmentado em dois tipos, o primeiro tipo é o  
 12 condicionamento operante ou instrumental, em que se manifesta um aumento na  
 13 probabilidade de ocorrência de um determinado comportamento, por exemplo, se a  
 14 consequência for reforçadora aumenta a probabilidade dela se repetir, mas se a  
 15 consequência for punitiva, pode diminuir ou extinguir a probabilidade de sua ocorrência  
 16 futura (STADDON e CERUTTI, 2003). O segundo tipo é o condicionamento pavloviano  
 17 clássico, que é o processo pelo qual um estímulo ambiental neutro adquire funções  
 18 similares às de um estímulo incondicionado através de emparelhamentos, com drogas de  
 19 abuso ou outro estímulo, de forma prévia e repetida (SCHULTZ e SCHULTZ, 1992; BARDO  
 20 e BEVINS, 2000).

21 O condicionamento clássico foi postulado por Ivan Pavlov, através de trabalhos com  
 22 cães. Em seu experimento mais conhecido, ele tocava um sino, Estímulo Neutro (EN), que  
 23 ocasionava uma Resposta Inespecífica e Neutra (RN), antes de apresentar comida aos  
 24 cães, a comida sendo um Estímulo Incondicionado (EI) e que gerava uma Resposta

1 Incondicionada (RI), que era a sialorreia. Após consecutivos pareamentos entre Estímulo  
2 Incondicionado e Estímulo Neutro, os cães passaram a salivar, Resposta Incondicionada  
3 (RI), ao som do sino sozinho, nesse caso, tendo uma Resposta Condicionada (RC). O  
4 Estímulo que originalmente era um Estímulo Neutro, se torna um Estímulo Condicionado  
5 (EC) (PAVLOV, 1927).

6 Segundo Catania (1999), o condicionamento clássico pode ser diferido em relação  
7 a sua programação temporal, em dois tipos: atraso e traço. No condicionamento de atraso,  
8 o tratamento farmacológico é administrado antes da colocação do animal no ambiente onde  
9 ele será testado. Desse modo, a resposta ao fármaco vai ocorrer concomitantemente com  
10 o estímulo do ambiente de testes. O outro tipo, o condicionamento de traço, tem-se o  
11 Estímulo Condicionado terminando antes do início da administração do fármaco, ou seja,  
12 o animal recebe a administração do fármaco após a saída da arena experimental e, para  
13 tanto, o intervalo entre os dois estímulos precisa ser breve. Em se tratando do  
14 condicionamento de atraso, que é um modelo menos utilizado, o indivíduo experimenta os  
15 estímulos da arena e do fármaco em consonância, ou seja, não existe intervalo temporal  
16 entre cada um dos estímulos (DE MELLO BASTOS *et al.*, 2014; SANTOS *et al.*, 2015). No  
17 condicionamento de traço o animal vivencia os estímulos de forma separada, ou seja,  
18 primeiro ele experimenta os efeitos causados pela permanência na arena experimental,  
19 imediatamente depois, ele vai experimentar os efeitos provocados pelo fármaco (CAREY  
20 *et al.*, 2014).

21 Sabe-se que a morfina, assim como outras drogas psicoativas, por exemplo,  
22 anfetamina e cocaína, causa mudanças permanentes no SNC, e que essa  
23 neuroplasticidade, após uso repetido, está envolvida com a dependência química  
24 (JACOBS *et al.*, 2003). Para tanto, os seus estímulos incondicionados podem ser  
25 facilmente condicionados a estímulos presentes em ambientes experimentais (SIEGEL,  
26 2001).

27 Trabalhos de Walter e Kuschinsky (1989), usando três grupos de ratos: o primeiro  
28 grupo (associado) foi condicionado 8 vezes com morfina 15 mg/kg, na presença de vários  
29 estímulos condicionados definidos (auditivo, olfativo e tátil). O segundo grupo (não-  
30 associado), foi exposto ao mesmo esquema de tratamento de morfina e estímulos, mas  
31 sem associação positiva entre droga e os estímulos. Um terceiro grupo (veículo) foi tratado  
32 com solução salina NaCl 0,9%, e exposto aos mesmos estímulos que os outros dois  
33 grupos. Todos os grupos foram testados para respostas condicionadas na presença dos

1 estímulos condicionados. Uma série de experimentos foi realizada com solução salina,  
2 depois de um intervalo de dois dias dos testes de condicionamento, uma segunda série foi  
3 testada com soro fisiológico após um intervalo de 7 dias, uma terceira série com morfina  
4 15 mg/kg após um intervalo de 2 dias, uma quarta série com a mesma dose de morfina  
5 após um intervalo de 7 dias. Os resultados mostraram que, quando a morfina foi usada  
6 após um intervalo de 2 dias, a tolerância farmacodinâmica "clássica" – mas não os  
7 fenômenos condicionantes – poderia explicar a mudança de comportamento, enquanto  
8 que, nos outros três protocolos descritos, alguns efeitos comportamentais condicionados  
9 puderam ser observados na presença ou na ausência de morfina, para os parâmetros  
10 monitorados (ativação locomotora, melhora das funções motoras, cheirar, roer).  
11 Consequentemente, o condicionamento contribui para a mudança de comportamento após  
12 a administração repetida de morfina.

13

#### 14 **3.4 Sensibilização e Morfina**

15

16 A sensibilização é um processo onde se tem o aumento progressivo da atividade  
17 locomotora, compulsão, comportamento de fuga, sendo que esse aumento ocorre após a  
18 administração repetitiva de psicoestimulantes, usando-se a mesma dose da substância  
19 química (WELLS e WELLS, 1971).

20 Também conhecida como tolerância reversa, a sensibilização comportamental é  
21 essencial para o processo de dependência química. A administração de drogas causa  
22 mudanças em nível cerebral, na via dopaminérgica mesolímbica associada ao núcleo  
23 accumbens (Nac), provocando sensibilização neural. Como consequência, esses sistemas  
24 se tornam hipersensíveis aos efeitos da droga e também a estímulos associados a eles,  
25 mesmo após cessada a administração da substância química. Mesmos depois de algum  
26 tempo sem o uso, a atividade locomotora ainda estará aumentada, devido à  
27 hiperpersensibilidade comportamental que é duradoura (CADOR *et al.*, 1995). Dessa  
28 forma, a sensibilização neural aumenta a busca pela droga (GARCIA *et al.*, 2011). Um  
29 único tratamento com anfetamina ou morfina induz a hiperlocomoção associada à hiper-  
30 responsividade da transmissão de dopamina mesocorticolímbica (VANDERSCHUREN *et*  
31 *al.*, 1999).



1 A sensibilização depende do padrão temporal da exposição ao medicamento,  
2 regimes de tratamento intermitente, com intervalos de 24 horas (dose diária) entre as  
3 administrações, são geralmente mais eficazes na indução da sensibilização do que os  
4 esquemas que empregam exposição a altas doses crescentes de medicamentos  
5 (ROBINSON e BECKER, 1986; STEWART e BADIANI, 1993; VANDERSCHUREN *et al.*,  
6 1997). É possível notar ainda, que até mesmo uma única exposição a drogas  
7 psicoestimulantes, como morfina, pode ser suficiente para provocar uma sensibilização  
8 duradoura (NASCIMENTO, 2011).

9 No trabalho de Vanderschuren e colaboradores (1997), foi observado efeito de  
10 hiperlocomoção em um protocolo de sensibilização comportamental por morfina, mesmo  
11 após transcorrer três semanas da última aplicação, o experimento consistia em uma  
12 aplicação diária de morfina 10 mg/kg, por via subcutânea (SC), em ratos Wistar, por um  
13 período de 14 dias consecutivos.

14 Viganò e colaboradores (2003), após administrarem doses crescentes de morfina  
15 (10 mg/kg - 20 mg/kg - 40 mg/kg), em ratos, duas vezes ao dia, durante três dias  
16 consecutivos, puderam constatar indução de sensibilização comportamental. O teste de  
17 sensibilização foi feito usando-se de uma dose baixa de morfina (5 mg/kg) por via SC,  
18 depois de passados 15 dias da última aplicação. Os resultados mostraram que houve  
19 aumento da locomoção, quando comparado ao grupo controle.

20 Em outro trabalho de Vanderschuren e colaboradores (2001), animais pré-expostos  
21 à dose única de morfina 10 mg/kg, exibiram um significativo aumento psicomotor. O mesmo  
22 tipo de exposição à morfina com dose diferente, 2 mg/kg não afetou o efeito psicomotor.  
23 Os testes de locomoção foram feitos três semanas após o tratamento o último  
24 farmacológico. Portanto, uma única exposição à morfina, em dose alta, induz a  
25 sensibilização comportamental duradoura e também a neuroadaptação associada.

26

### 27 **3.5 MK-801, Sistema Dopaminérgico e o Sistema Glutamatérgico**

28

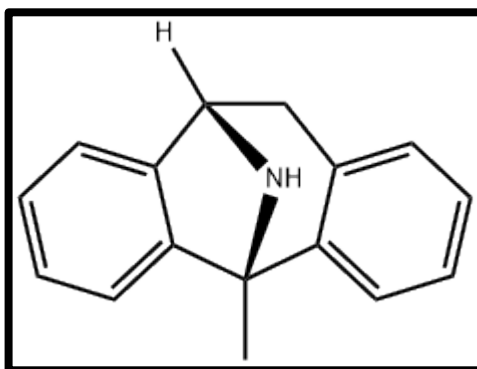
29 A presença da monoamina dopamina no SNC sugere um papel de neuromodulação  
30 celular, que está ligada ao glutamato no processo da dependência química. A liberação  
31 glutamatérgica reflete em excitação neuronal e, por outro lado, a presença do

1 neurotransmissor GABA causa inibição celular (JONAS *et al.*, 1998; NABEKURA *et al.*,  
2 2004)

3 O maleato de dizocilpina-MK-801 (figura 3) é um fármaco antagonista  
4 glutamatérgico dos receptores ionotrópicos do tipo NMDA (N-metil-D-aspartato), tendo os  
5 seus efeitos dependentes da dose, pois se trata de um antagonista não competitivo (FORD  
6 *et al.*, 1989; ÖGREN e GOLDSTEIN, 1994).

7 O uso de doses baixas (0,025 mg/kg) de MK-801 provoca diminuição da locomoção  
8 em ratos em campo aberto e diminuição da ansiedade/medo ligada as reações defensivas  
9 antipredadoras (BLANCHARD *et al.*, 1992). No caso do uso em doses elevadas (0,1 – 0,3  
10 mg/kg), observaram-se os seus efeitos excitatórios como hiperatividade locomotora em  
11 ratos, esses são os resultados encontrados por Carey (1995) e Hargreaves e  
12 colaboradores (1992). As alterações locomotoras como a hipoatividade e hiperatividade,  
13 nos mostram que há uma estreita relação dos níveis de glutamato com algumas mudanças  
14 comportamentais, pois o glutamato é um neurotransmissor que desempenha um papel  
15 fundamental no comportamento locomotor e na memória a longo prazo no processo  
16 dependência química (SPENCER *et al.*, 2016).

17



18

19 Figura 3: Fórmula estrutural do maleato de dizocilpina-MK-801 (5S,10R) -(+) -5 Methyl-  
20 10,11-dihydro-5H-dibenzo[a,d]cyclohepten-5,10-iminemaleate). Adaptado de Yi e  
21 colaboradores (2020).

22

23 Há algum tempo, a importância dos gânglios da base no comportamento locomotor  
24 já é bem esclarecida. Estruturas como o núcleo accumbens, córtex pré-frontal, amígdala,  
25 hipotálamo e área tegmental ventral, medeiam a interação entre as vias glutamatérgicas e

1 dopaminérgicas, e dessa forma a atividade inibitória glutamatérgica é equilibrada pela  
2 atividade excitatória dopaminérgica (DAI e CAREY 1995).

3 Diversos estudos mostraram que fármacos agonistas dopaminérgicos diretos ou  
4 indiretos dopaminérgicos podem induzir hiperatividade e promover condicionamento e  
5 sensibilização (MATTINGLY *et al.*, 1997, 1988; ANAGNOSTARAS e ROBINSON, 1996;  
6 ROWLETT *et al.*, 1997; KELLER e DELIUS, 2001; BLOISE *et al.*, 2007; BRAGA *et al.*,  
7 2009; DE MATOS *et al.*, 2010). Da mesma forma, fármacos antagonistas glutamatérgicos  
8 do tipo NMDA, como o MK-801, podem induzir hiperatividade em ratos, e ainda, que essa  
9 hiperatividade se torna progressiva com o uso de tratamentos repetitivos, indicando  
10 sensibilização (CAREY *et al.* 1995).

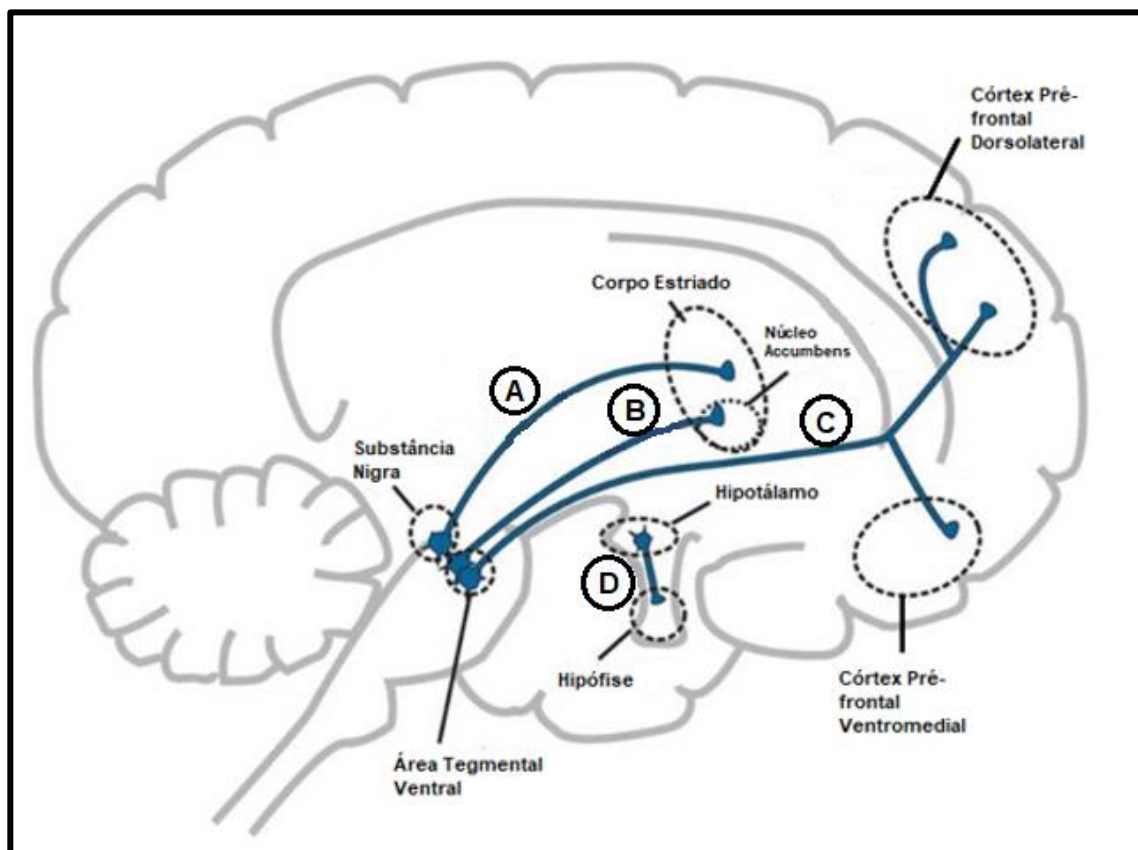
11 Os sistemas dopaminérgicos bem como o glutamatérgico estão implicados na  
12 adição por opioides e outras drogas psicoativas como os opioides, o álcool e a cocaína.  
13 Nesse contexto os estudos terapêuticos atuais se voltam para drogas, que sejam capazes  
14 de equilibrar a transmissão do glutamato e da dopamina no contexto de dependência  
15 química (OLIVE *et al.*, 2012; COLLINS *et al.*, 1998; D'SOUZA, 2015).

16 A dopamina é um neurotransmissor endógeno da família das catecolaminas, sua  
17 produção ocorre na substância nigra e na área tegmental ventral (VTA) no cérebro.  
18 Existem, pelo menos, 5 receptores dopaminérgicos, D1, D2, D3, D4 e D5. Sendo D2, D3 e  
19 D4 com grande afinidade entre si, enquanto D1 e D5 compartilham de características  
20 semelhantes. A dopamina está envolvida no controle da cognição, humor, aprendizado,  
21 memória, emoções e na manutenção das funções renal, gastrointestinal e cardiovascular  
22 (MATSUMOTO *et al.*, 2005).

23 Kenna e colaboradores (2007) mostraram que propriedades reforçadoras de drogas  
24 psicoativas com potencial de abuso têm relação direta e indireta com o sistema  
25 dopaminérgico. Os receptores dopaminérgicos se distribuem de forma ampla no  
26 hipotálamo, na substância nigra e na área tegmental ventral (VTA), dando origem às quatro  
27 principais vias dopaminérgicas (figura 4) no sistema nervoso central, a via mesocortical, a  
28 via mesolímbica, a via nigroestriatal e a via túbero-infundibular (BERKE, 2018).

29 A compreensão e o estudo das quatro vias dopaminérgicas, do sistema nervoso  
30 central e dos aspectos envolvidos na neurotransmissão são fundamentais para entender a  
31 vasta gama de funções desempenhadas por estas vias. Falhas nas estruturas de  
32 neurotransmissão das vias dopaminérgicas mesolímbica e mesocortical, localizadas no

1 sistema nervoso central estão relacionadas com doenças graves, como a dependência  
2 química (SALAMONE e CORREA, 2012).



3  
4 Figura 4: Vias Dopaminérgicas. (A): Via Nigroestriatal; (B): Via Mesolímbica; (C): Via  
5 Mesocortical; (D): Via Tubero-infundibular.

6  
7 A via mesolímbica é uma das vias dopaminérgicas mais importantes, por suas  
8 funções destacadas com relação à dopamina. Está ligada diretamente ao sistema de  
9 recompensa cerebral. A via mesolímbica se origina na área tegmental ventral, uma região  
10 rica em neurônios dopaminérgicos e que cobre parte do mesencéfalo, e projeta os seus  
11 feixes nervosos para outras áreas importantes do sistema nervoso central, como o córtex  
12 pré-frontal, o hipocampo, o complexo amigdalóide e, também, para o *núcleo accumbens*.  
13 É no *núcleo accumbens* que a dopamina exerce, em um primeiro instante, sua ação como  
14 mediadora dos sentimentos de prazer e recompensa (ADINOFF, 2004).

15 Portanto, no instante em que um determinado indivíduo tem contato com um  
16 estímulo de caráter recompensador ou prazeroso, como comida, sexo ou drogas, ocorre  
17 liberação de dopamina cujos sinais percorrem da área tegmental ventral ao núcleo  
18 accumbens. O caminho feito pela dopamina possibilita a criação e o reforço de sensações

1 positivas que acabam por acentuar e modular o comportamento do indivíduo (SOMALWAR  
2 *et al.*, 2018).

3 Outra via dopaminérgica importante com projeções originadas na área tegmental  
4 ventral é a via mesocortical, suas projeções se encaminham para o córtex pré-frontal. A  
5 via mesocortical é uma das maiores vias dopaminérgicas junto com a via mesolímbica e  
6 tem grande importância para o desenvolvimento normal das funções cognitivas,  
7 recompensa, aprendizagem, atenção, tomadas de decisão, memória de trabalho e  
8 resposta emocional. Está presente na fisiopatogênese da esquizofrenia, patologia em que  
9 ocorre a diminuição de dopamina nessa via cerebral (DIAZ, 1996; MORGANE *et al.*, 2005).

10 A via nigroestriatal tem projeções dopaminérgicas, com origem na substância nigra,  
11 indo até o corpo estriado e integra os gânglios basais. A ativação da via nigroestriatal,  
12 produz um aumento na liberação de dopamina no corpo estriado dorsal, e também produz  
13 estereotípias. Essa via está relacionada com a locomoção e promove aprendizado dos  
14 comportamentos do vício (RIVERA *et al.*, 2013). Os neurônios dopaminérgicos presentes  
15 nessa via são responsáveis pela estimulação motora voluntária e contém em torno de 80%  
16 da reserva de toda a dopamina do sistema nervoso central (YADAV *et al.*, 2014).

17 O Glutamato é um aminoácido proteinogênico presente em diversos tipos de  
18 alimentos, tem grande importância para processos metabólicos como a gliconeogênese, a  
19 glicólise e o ciclo dos ácidos tricarbóxicos (REEDS *et al.*, 2000). O glutamato é um  
20 neurotransmissor com grande importância para o SNC, principalmente por seus efeitos  
21 excitatórios (CONTI, 1998). Ele está presente em cerca de 80% de todas as sinapses  
22 nervosas, regula muitas emoções e está envolvido na formação de lembranças,  
23 aprendizado e na atenção, assim como nas situações patológicas responsáveis por  
24 diversas desordens neuropsiquiátricas agudas e crônicas como na dependência química,  
25 ansiedade generalizada, dor neuropática, esquizofrenia e depressão, (LI *et al.*, 2019;  
26 MELDRUM, 2000; FEATHERSTONE, 2010).

27 Existem dois grandes grupos de receptores glutamatérgicos, os ionotrópicos  
28 (iGluRs) e os metabotrópicos (mGluRs). Os receptores metabotrópicos se dividem em 3  
29 grupos: grupo 1 (mGluR1 e mGluR5), grupo 2 (mGluR2 e mGluR3) e grupo 3 (mGluR4,  
30 mGluR6, mGluR7 e mGluR8), (REINER e LEVITZ, 2018; BONSI *et al.*, 2005). Os  
31 receptores ionotrópicos podem ser divididos em três tipos: N-metil-D-aspartato (NMDA),  
32 ácido-amino-3-hidroxi-5-metil-isoxazol-4-propiónico (AMPA) e cainato (KA) (RUGGIERO *et al.*,  
33 *et al.*, 2011). Os receptores ionotrópicos NMDA, AMPA e KA possuem alguns subtipos e

1 esses são classificados conforme as diferentes combinações das subunidades proteicas  
2 que os formam.

3 A neurotransmissão quando mediada pelos receptores ionotrópicos é rápida, pois o  
4 fluxo de íons é afetado de maneira imediata (principalmente Na<sup>+</sup> e Ca<sup>2+</sup>) e da mesma  
5 forma o estado eletroquímico da membrana pós-sináptica (BOWIE, 2008). O receptor do  
6 tipo AMPA é responsável pela maioria das transmissões sinápticas excitadoras rápidas, o  
7 tipo KA participa nas respostas pós-sinápticas em sinapses excitadoras, podendo ainda  
8 modular a liberação pré-sináptica do transmissor glutamato em determinadas sinapses,  
9 enquanto o receptor do tipo NMDA é fundamental na indução de formas específicas de  
10 plasticidade sináptica (ENGELHARDT, 2003).

11 Existem pelo menos sete vias glutamatérgicas (figura 5). A primeira via se chama  
12 córtico-troncar, essa via tem projeções indo do córtex até o tronco cerebral. A via córtico-  
13 troncar descende dos neurônios piramidais corticais no córtex pré-frontal e seguem para o  
14 tronco cerebral. É uma via glutamatérgica muito importante pois é a chave na regulação  
15 da liberação de neurotransmissores, envolve também a substância nigra e VTA para a  
16 dopamina, os núcleos da rafe para a serotonina e o locus coeruleus para a norepinefrina.

17 A segunda, via também descendente se projeta do córtex pré-frontal ao corpo  
18 estriado dorsal (via glutamatérgica córtico-estriatal) ou pode ainda se projetar do córtex  
19 pré-frontal ao núcleo accumbens no corpo estriado ventral (via glutamatérgica cortico-  
20 accumbens), e constituem juntas a porção das alças cortico-estriatal-talâmica. Essas vias  
21 descendentes de glutamato terminam nos neurônios GABA que têm como destino uma  
22 estação de retransmissão chamada globo pálido, essa estação tem grande importância na  
23 coordenação motora, na manutenção e aquisição de informações e processamento de  
24 emoções (FOX *et al.*, 1974; CAMARGO *et al.*, 1981).

25 A terceira via glutamatérgica tem grande importância, pois algumas teorias vinculam  
26 essa via específica à esquizofrenia. Esta via se projeta do hipocampo para o núcleo  
27 accumbens e é conhecida como via glutamatérgica hipocampal-accumbens. Da mesma  
28 forma que a segunda via glutamatérgica (vias córtico-estriatal e córtico-accumbens), a  
29 terceira via também tem neurônios que saem do hipocampo e fazem sinapses com  
30 neurônios gabaérgicos presentes no núcleo accumbens, que por sua vez se projetam para  
31 uma estação de retransmissão no globo pálido.

1 A quarta via é uma via ascendente, os neurônios saem do tálamo e fazem sinapses  
2 com neurônios piramidais no córtex (via glutamatérgica tálamo-cortical). Normalmente,  
3 essa via está envolvida no processamento de informações sensoriais.

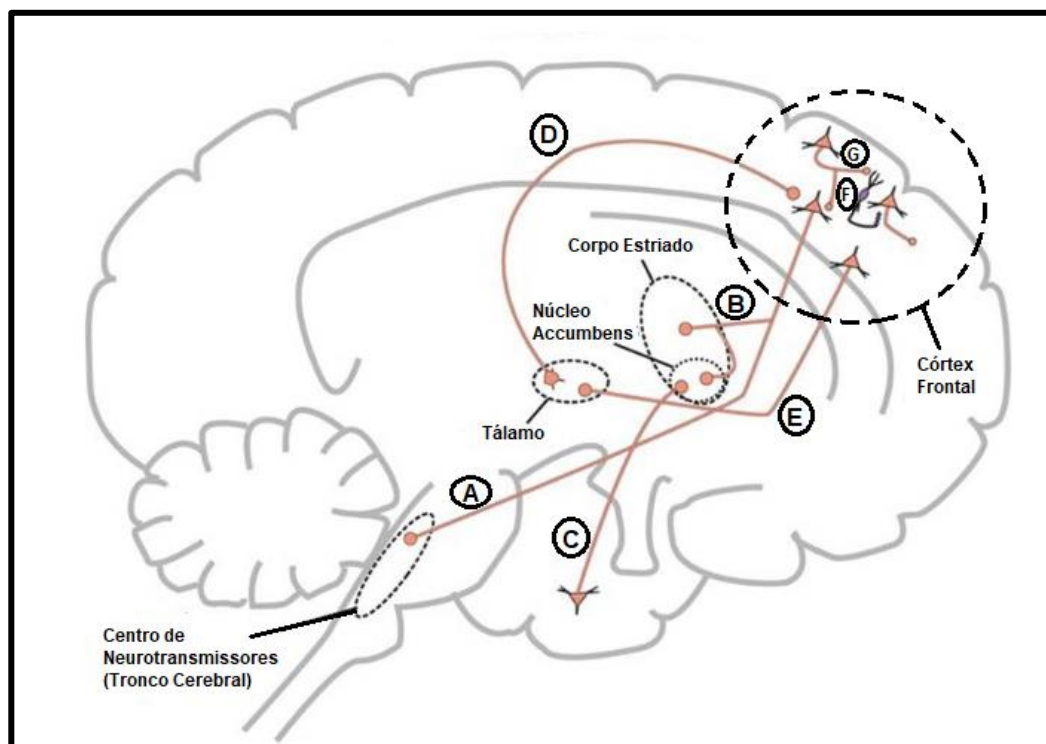
4 A quinta via é descendente, e segue do córtex pré-frontal indo até o tálamo (via  
5 glutamatérgica córtico-talâmica). Essa via pode direcionar a forma pela qual os neurônios  
6 reagem às informações sensoriais.

7 A sexta via é a córtico-cortical direta e é formada pelos neurônios piramidais  
8 intracorticais que liberam o neurotransmissor glutamato, que é excitatório.

9 A sétima via é a via córtico-cortical indireta, nessa via os neurônios piramidais fazem  
10 sinapses com interneurônios (que liberam o GABA) e estes fazem sinapses com neurônios  
11 presentes no córtex.

12 Quatro das sete vias glutamatérgicas se projetam a partir do córtex frontal e  
13 penetram em áreas mais profundas do cérebro, onde exercem controle sobre as estruturas  
14 neuroanatômicas como o corpo estriado ventral e dorsal, o núcleo accumbens, o tálamo e  
15 o tronco cerebral (STAHL, 2008; SCHWARTZ *et al.*, 2012).

16



17

1 Figura 5: Vias glutamatérgicas. (A): via córtico-troncar; (B): via córtico-estriatal e córtico-  
2 accumbens; (C): Via hipocampal-accumbens; (D): via tálamo-cortical; (E): via córtico-  
3 talâmica; (F): via córtico-cortical direta; (G): via córtico-cortical indireta.

4

5 Spencer e colaboradores (2016) esclarecem que o envolvimento da dopamina na  
6 dependência química não é completo, ou seja, ela contribui como o neurotransmissor  
7 responsável pelas associações relacionadas à droga como, por exemplo, o prazer, e,  
8 dessa forma, induzem ao desejo por voltar ao consumo da substância, enquanto que o  
9 glutamato é o maior causador da recaída após um tempo em abstinência, portanto a  
10 memória a longo prazo está mais ligada aos efeitos excitatórios do glutamato pelas vias  
11 descendentes do córtex (córtico-troncar, córtico-estriatal, córtico-accumbens, córtico-  
12 talâmica) no SNC.

13 Os opioides, como a morfina, provocam um aumento da atividade glutamatérgica e  
14 dopaminérgica na via mesolímbica, que acontece por meio da inibição dos neurônios  
15 gabaérgicos; dessa forma, ocorre diminuição do GABA e aumento da dopamina liberada  
16 pelos neurônios dopaminérgicos, por outro lado ocorre liberação pré-sináptica de  
17 glutamato por meio da desinibição por neurônios gabaérgicos no VTA (CHEN *et al.*, 2015;  
18 CHARTOFF e CONNERY, 2014). A morfina, quando administrada repetidamente em um  
19 ambiente determinado, produz uma série de neuroadaptações que levam ao  
20 desenvolvimento de tolerância, dependência e sensibilização. Assim como outras drogas  
21 de abuso, ela afeta as vias dopaminérgicas mesolímbica e mesocortical do SNC. A  
22 ativação do sistema dopaminérgico mesolímbico (circuito de recompensas) provoca um  
23 aumento na liberação de dopamina no *núcleo accumbens* e é responsável pelas  
24 propriedades de reforço dos fármacos (WISE, 1996; BARDO, 1998).

25 Existe uma diferença no curso temporal em que se tem um pico de atividade motora  
26 e níveis de concentração extracelulares no corpo estriado cerebral, para o glutamato, esse  
27 tempo é de 30 – 90 minutos, diferindo da dopamina, que ocorre entre 15 e 30 minutos após  
28 estímulo com psicoestimulantes como exemplo a morfina (GRAY *et al.*, 2001; RAWLS e  
29 MCGINTY, 2000).

30 A ativação das vias dopaminérgicas normalmente está ligada a respostas  
31 ocasionadas por riscos e recompensas imprevisíveis, enquanto que a ativação das vias  
32 glutamatérgicas é informativa sobre o contexto da situação (SCHULTZ, 2010).

33



### 1 3.6 Proteína Quinase Ativada por Mitógenos – ERK

2

3 A ERK é uma proteína quinase regulada por sinais extracelulares (mitógenos);  
4 (JOHNSON e LAPADAT, 2002). A proteína é ativada de forma a coordenar a proliferação,  
5 diferenciação e sobrevivência de muitos tipos de células, principalmente os neurônios no  
6 SNC (LEWIS *et al.*, 1998; DAVIE e SPENCER, 2001; MORRISON, 2012).

7 Algumas doenças como o câncer, as síndromes imunológicas, inflamatórias e  
8 degenerativas podem decorrer de defeitos na via relacionados à ERK (HINDLEY e KOLCH,  
9 2002). Na dependência química, a sinalização celular está desregulada, o que ocasiona  
10 um aumento dos níveis de produção de ERK, quando comparado ao aumento por  
11 estímulos naturais, dessa forma contribuindo para a neuroplasticidade sináptica após  
12 doses repetidas de drogas (GIRAULT *et al.*, 2007; SHIFLETT e BALLEINE, 2011).

13 A dopamina modula a transmissão de glutamato e controla a plasticidade estriatal  
14 induzida por drogas de abuso por meio do receptor do tipo D1. Um dos principais alvos dos  
15 receptores D1 presentes no estriado cerebral é a proteína quinase ativada por mitógenos  
16 (ERK). A ativação da ERK por drogas de abuso se comporta como um fator integrador  
17 chave de sinalização dos receptores dopaminérgicos tipo D1 e glutamatérgicos tipo NMDA,  
18 formando um núcleo central de neuroplasticidade (WICKENS, 2009; CAHILL *et al.*, 2014).

19 Sanguedo e colaboradores (2015 e 2017), mostraram aumento da locomoção e  
20 ativação da ERK1/2 no córtex pré-frontal, amígdala, núcleo accumbens, hipotálamo lateral  
21 em grupos de ratos que receberam apomorfina (2,00 mg/kg), em dose alta, a substância  
22 provoca aumento dos níveis circulantes de dopamina. Essas estruturas cerebrais são as  
23 principais relacionadas ao circuito de recompensa e motivação e são também responsáveis  
24 pela aquisição e manutenção da dependência química.

25 Mitra e Sinatra (2004) mostraram um aumento na densidade e alteração dos  
26 receptores opioides acoplados à proteína G, após o uso crônico de morfina, juntamente  
27 com a alteração dos receptores foi visto ainda uma maior ativação de ERK 1/2 em  
28 indivíduos que se tornaram dependentes de opioides após passarem por procedimentos  
29 cirúrgicos.

30 A ativação de ERK1/2 *in vivo* após uso de morfina sinaliza a expressão gênica em  
31 regiões como o núcleo accumbens e amígdala, e dessa forma contribui para a sua

1 permanência a longo prazo (ASENSIO *et al.*, 2006). Ainda pode ocorrer sinalização no  
2 início da abstinência com aumento da ERK 1/2 em regiões como amígdala, locus  
3 coeruleus, hipotálamo, córtex cerebral, septo lateral (CICCARELLI *et al.*, 2013; HOFFORD  
4 *et al.*, 2009).

5

## 6 **MATERIAL E MÉTODOS**

7

### 8 **4.1 Sujeitos**

9

10 Foram utilizados ratos machos, albinos, Wistar, pesando entre 200 - 250g, oriundos do  
11 Biotério Central da UENF, Campos Dos Goytacazes, RJ. Os animais foram mantidos em  
12 gaiolas individuais de plástico (BEIRA MAR, São Paulo) medindo 25x18x17 cm, com  
13 acesso livre à ração padronizada de laboratório e à água. As gaiolas foram mantidas em  
14 uma sala no setor de Farmacologia do Laboratório de Morfologia e Patologia Animal  
15 (LMPA), com umidade e temperatura controladas ( $22 \pm 2.0^{\circ}\text{C}$ ), e com ciclo de luz claro e  
16 escuro de 12 em 12 horas (luz das 07:00 às 19:00 horas). O experimento foi conduzido na  
17 fase clara, horário entre 09:00 e 14:00 horas. Os animais foram manipulados  
18 individualmente, por um único indivíduo, pelo tempo de 5 minutos durante 7 dias antes do  
19 início do procedimento experimental e foram habituados ao procedimento de injeção com  
20 solução de veículo durante 3 dias antes do início do experimento. O presente projeto foi  
21 aprovado pela comissão de ética de uso de animais (CEUA – UENF), sob o nº 029/2022,  
22 Protocolo 473.

23

### 24 **4.2 Ambiente Experimental**

25

26 O presente experimento foi desenvolvido em quatro salas experimentais  
27 padronizadas medindo 1,40 x 1,40 metros. Cada sala constituída de iluminação vermelha,  
28 temperatura controlada ( $22 \pm 2,0^{\circ}\text{C}$ ) e o som de um ventilador (30 cm de hélice) ligado em  
29 cada uma das salas como ruído de fundo. Cada sala contendo uma arena quadrada  
30 medindo 60x60x45cm, com assoalho e paredes pintados na cor preta. Para o registro do

1 comportamento locomotor, foram utilizadas câmeras (IIKEGAMI, modelo ICD – 49 e  
2 Panasonic, modelo WV – BP334), posicionadas 60 cm acima das arenas experimentais.  
3 As câmeras ficaram acopladas a um computador PC compatível, contendo um programa  
4 de análise de imagens, EthoVision (NOLDUS, Holanda), o qual estava localizado fora das  
5 salas de experimento, o sistema quantificou a atividade locomotora em distância percorrida  
6 (metros).

7

### 8 **4.3 Fármacos**

9

10 O sulfato de morfina (Dimorf 10mg/ml, Cristália®, São Paulo, SP, Brasil) foi utilizado  
11 na dose de 10 mg/kg (VANDERSCHUREN *et al.*, 1997, 2001) e administrado por via  
12 subcutânea (SC). O volume de administração foi de 1ml/kg.

13 A apomorfina-Hcl (Sigma-Aldrich®, Saint Louis, Mo, USA) foi utilizada na dose de  
14 0,05 mg/kg, dissolvida em uma solução salina (NaCl 0,9%) e administrada por via  
15 subcutânea (volume de administração: 1ml/kg).

16 O maleato de dizocilpina (MK-801, Sigma Aldrich®, Saint Louis, Mo, USA) foi  
17 utilizado nas doses de 0,025 mg/kg, 0,1 mg/kg e 1.0 mg/kg por via subcutânea (SC). O  
18 volume de administração foi de 1ml/kg.

19 A solução salina (NaCl 0,9%) foi utilizada como veículo na concentração de 1 ml/kg  
20 e administrada por via subcutânea.

21 A solução Carbamato de etila (Urethane, Sigma Aldrich®, Saint Louis, Mo, USA) foi  
22 utilizado na dose de 3000 mg/kg (CFMV, 2013; CEUA-UNIFESP, 2017; HENKE *et al.*,  
23 2016) e administrado por via intraperitoneal. O volume de administração foi de 15 ml/kg.

24 As soluções foram preparadas antes de cada dia experimental.

### 25 **4.4 Procedimento Experimental**

26

27 Os experimentos foram conduzidos segundo o protocolo experimental de Carrera e  
28 colaboradores (2013), Carrera e colaboradores (1995), Miczek e colaboradores (2011),  
29 Crespo e colaboradores (2022) com modificações.

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26

## **5. MANUSCRITOS**

Os dois manuscritos com os dados desta tese serão apresentados em dois capítulos:

Primeiro capítulo, o manuscrito “Morphine and dopamine: Low dose apomorphine can prevent both the induction and expression of morphine locomotor sensitization and conditioning”, submetido à revista “Behavioural Brain Research” foi apresentado.

Segundo capítulo, o manuscrito MK-801 “Induces dose dependent stimulant sensitization effects but dose independent conditioned stimulant effects: MK-801 effects on sensory information processing versus learning and memory, submetido à revista “Psychopharmacology” foi apresentado.

### **5.1 - Capítulo I**

**Morphine and dopamine: Low dose apomorphine can prevent both the induction**

1 **and expression of morphine locomotor sensitization and conditioning.**

2 Joaquim Barbosa Leite Júnior<sup>1</sup>, Luiz Gustavo Soares Carvalho Crespo<sup>1</sup>, Richard Ian  
3 Samuels<sup>2</sup>, Norberto Cysne Coimbra<sup>3</sup>, Robert J Carey<sup>4</sup> and Marinete Pinheiro Carrera<sup>1</sup>

4

5 <sup>1</sup>Behavioral Pharmacology Group, Laboratory of Animal Morphology and Pathology,  
6 Universidade Estadual do Norte Fluminense Darcy Ribeiro, Avenida Alberto Lamego,  
7 2000, Campos dos Goytacazes, 28013-602, RJ, Brazil.

8 <sup>2</sup>Department of Entomology and Plant Pathology, Universidade Estadual do Norte  
9 Fluminense Darcy Ribeiro, Campos dos Goytacazes, RJ, Brazil.

10 <sup>3</sup>Department of Pharmacology, Ribeirão Preto Medical School of the University of São  
11 Paulo, SP, Brazil.

12 <sup>4</sup>Department of Psychiatry, SUNY Upstate Medical University, 800 Irving Avenue,  
13 Syracuse, NY13210, USA

14

15

16 Number of text pages of the whole

17 manuscript: 30Number of figures and tables:

18 12

19

20

21 Corresponding author: Dr. Marinete Pinheiro Carrera, Behavioral Pharmacology Group,  
22 Laboratory of Animal Morphology and Pathology, Universidade Estadual do Norte  
23 Fluminense Darcy Ribeiro, Avenida Alberto Lamego, 2000, Campos dos Goytacazes,  
24 28013-602, RJ, Brazil. Fax: +55-22-27397197; e-mail: marinete@uenf.br

**1 Abstract**

2  
3 The disinhibition of dopamine neurons in the VTA by morphine is considered an important  
4 contributor to the reward potency of morphine. In this report, three separate experiments  
5 were conducted in which a low dose of apomorphine (0.05 mg/kg) was used as a  
6 pretreatment to reducedopamine activity in the brain. Locomotor hyperactivity was used  
7 as the behavioral response to morphine (10.0 mg/kg). In the first experiment, five  
8 treatments with morphine induced the development of locomotor and conditioned  
9 hyperactivity. Apomorphine, given 10 min prior to morphine, prevented the development  
10 of morphine-induced hyperactivity and conditioning. Apomorphine before either vehicle or  
11 morphine injections induced equivalent reductions inspontaneous activity in the vehicle-  
12 and morphine-treated groups. In the second experiment, a pretreatment with apomorphine  
13 was performed after the morphine-induced conditionedhyperactivity and apomorphine  
14 eliminated the expression of the morphine-conditioned response. To assess the effects of  
15 the apomorphine on VTA and the nucleus accumbens, ERK measurementswere carried  
16 out. Morphine increased ERK activation in these encephalic areas and the increase inERK  
17 activity was associated with enhanced locomotion. A third experiment was conducted to  
18 assess the effects of acute morphine on ERK before locomotor stimulation was induced  
19 by morphine. Acute morphine did not increase locomotion but a robust ERK response was  
20 produced.The increases in ERK activation were impaired by the apomorphine. The results  
21 provide substantial support for the importance of ongoing activity in dopamine neurons for  
22 the efficacy ofmorphine disinhibition of dopamine reward systems in the mediation of  
23 morphine behavioral sensitization and conditioning.

24 **Key words:** Morphine; Dopamine; VTA; ERK; Behavioral sensitization; Drug conditioning.

## 1 **1. Introduction**

2

3 It has been well established that dopamine neurons in the ventral tegmental area (VTA) are  
4 of substantial importance in the mediation of behavioral reward effects [12, 18, 41]. Not  
5 surprisingly, dopaminergic drugs that can mediate reward effects also activate VTA  
6 dopaminergic neurons [17, 36, 40]. Morphine can also activate VTA dopaminergic neurons  
7 but, unlike psychostimulant dopaminergic drugs, morphine produces this pharmacological  
8 action indirectly by a disinhibitory and inhibitory effects. Morphine binds to  $\mu$ -opioid  
9 receptors located on  $\gamma$ -aminobutyric acid (GABA)ergic neurons that inhibit VTA  
10 dopaminergic neurons. This inhibition of GABAergic neurons by morphine recruiting  $\mu$ -  
11 opioid receptor-signaling can reduce the GABAergic inhibition on dopaminergic neurons  
12 and thereby enhance the ongoing activity in the dopaminergic neurons [6, 16, 21, 23, 34,  
13 38]. Somewhat analogous to the indirect effects of morphine on VTA dopaminergic  
14 neurons, it has been suggested [8, 11] that morphine-induced behavioral increases in  
15 spontaneous activity [20, 26, 28, 35, 39] are not a result of a direct morphine activation of  
16 the motor system but rather a consequence of morphine activation of VTA-induced reward  
17 effects that adventitiously occur in association with ongoing spontaneous activity and  
18 reinforce and potentiate the spontaneous behavior.

19 The magnitude of the impact of the disinhibitory effect of morphine on dopaminergic  
20 neurons to increase dopamine activity is presumably related to the level of the activity of  
21 the dopaminergic neurons. Seemingly, the lower the ongoing activation state of VTA  
22 dopaminergic neurons, the less the magnitude of the increase in VTA dopamine activity  
23 induced by a reduction of GABAergic inhibition. Clearly, if the dopaminergic system is  
24 inactive then the morphine disinhibitory effects would not increase VTA dopaminergic  
25 system reward activation and, consequently, there would be no reinforcement and  
26 potentiation of spontaneous behavioral

1 activity. One pharmacologically effective treatment for reducing activity in dopaminergic  
2 neurons is the use of a low dose of apomorphine. At low dose range in rats (<0.1 mg/kg),  
3 apomorphine can induce inhibition of dopaminergic neurons by preferential stimulation of  
4 dopamine autoreceptors, thereby inhibiting dopaminergic activity in the brain [2, 7, 10, 24].  
5 This work was undertaken to use the pretreatment with apomorphine (0.05 mg/kg)  
6 followed by the treatment with morphine to reduce ongoing activity in dopaminergic  
7 neurons. We measured the effects of apomorphine pretreatment on morphine-induced  
8 effects on spontaneous activity and on extracellular signal regulated kinase activity (ERK)  
9 in subcortical brain areas implicated in dopamine-related reward system nuclei namely the  
10 VTA and nucleus accumbens (NAc). The rationale for measuring ERK in these encephalic  
11 structures was based on findings that have reported increases in ERK activity in  
12 dopaminergic projection areas [1, 14, 22, 29] including the NAc [5, 13, 19, 37] following  
13 treatment with psychostimulant drugs such as cocaine. Additionally, in previous studies  
14 [30, 31, 32, 33] focused on apomorphine behavioral sensitization, we showed that this  
15 sensitization was associated with a strong ERK signal in brain dopaminergic systems. In  
16 the present study, we used ERK as an indicator of activation in VTA and NAc subcortical  
17 areas associated with reward systems. This report details the effects of a low dose  
18 apomorphine pretreatment on morphine-induced behavioral sensitization and  
19 conditioning, and ERK activity in the VTA and NAc of laboratory animals submitted to three  
20 independent experiments.



## 1    **2. Method**

2

### 3    2.1. Subjects

4

5    Male Wistar rats provided by the Universidade Estadual do Norte Fluminense Darcy  
6    Ribeiro, initially weighing 200-300 g, were housed in individual plastic cages (25 X 18 X  
7    17 cm) until the end of the experiment. Food and water were freely available at all times.

8    The vivarium was maintained at a constant temperature ( $22 \pm 2$  °C), and a 12h/12h  
9    light/dark cycle (lights on at 7 a.m. and off at 7 p.m.). All experiments were performed

10    between 8 a.m. and 12 p.m. For 7 days prior to all experimental procedures, each animal  
11    was weighed and handled daily for 5 min. This process included being placed in a transport

12    cage and taken to the injection administration bench located in an anteroom adjacent to

13    the experimental testing room. The animals were given vehicle (VEH) injections. In this

14    way, the animals were familiarized with the handling and injection aspects of the testing

15    protocol. All the experiments were performed in compliance with Brazilian Council for

16    Animal Experimentation (CONCEA), which are based on the US National Institutes of

17    Health guide for the care and use of laboratory animals (NIH Publications No. 8023,  
18    revised 1978).

19

20

### 21    2.2. Drugs

22

23    Morphine sulfate (Cristália, São Paulo, Brazil) was used from 10 mg ampoules (1 ml) and  
24    was injected subcutaneously in the nape of the neck at a dose of 10 mg/kg. Apomorphine-

25    HCl (Sigma, St. Louis, MO, USA) was dissolved in 0.9% NaCl solution and was injected

26    subcutaneously in the nape of the neck in a dose of 0.05 mg/kg. Physiological saline was

27    used as

1 vehicle (VEH). All doses were administered in a volume of 1.0 ml/kg body weight. Drug  
2 solutions were freshly prepared before each experiment.

3  
4

### 5 2.3. Apparatus and behavioral measurements

6  
7 The behavioral measurements were conducted in a black open field test enclosure (60 x  
8 60x 45 cm). A closed-circuit camera (IKEGAMI, model ICD-49; Ikegami Electronics Inc.,  
9 Torrance, California, USA) mounted 60 cm above the arena was used to record behavioral  
10 data. Locomotion, measured as distance traveled (m), was automatically analyzed using  
11 EthoVision (Noldus, the Netherlands). The complete test procedure was conducted  
12 automatically without the presence of the experimenter in the test room. All behavioral  
13 testing was conducted under dim red light that favors exploratory behavior and avoids the  
14 possible aversive quality of white light [25] as well as to enhance the contrast between the  
15 white subject and the dark background of the test chamber and reduce the animal's shadow.

16  
17

### 18 2.4. General experimental procedure

19  
20 Initially, all rats were acclimated to the test arena by being injected with VEH and placed  
21 in the arena for 30 min on 3 successive days (habituation phase). The habituation protocol  
22 was conducted so that a stable baseline of the locomotor behavior could be established  
23 prior to the start of the drug treatments. On the basis of the locomotion distance scores  
24 from the third habituation session, groups were formed that equated for the distance  
25 scores. Next, the groups received their pharmacological treatment immediately prior to the  
26 5 min placement in the open-field arena and the locomotor activity was measured. These  
27 treatments were administered for 5 consecutive days

1 (experiments 1 and 2), one trial per day, and were designed as the induction phase. On  
2 the following day, the final test was performed, in which the animals received their  
3 treatment immediately before being placed into the open-field arena for a 5-min session.  
4 Immediately upon completion of the final test, the animals were euthanized, intracardially  
5 perfused with physiological saline and fixative and immunohistochemistry was performed.  
6 Figure 1A shows the experimental timeline.

7  
8

9 2.5. Experiment 1: Effect of a low dose of apomorphine pretreatment on the acquisition  
10 of morphine-induced locomotor sensitization, conditioning, and ERK activity in VTA  
11 and NAc.

12 The protocol described in the general experimental procedure was followed. In the  
13 induction phase, the rats received the first injection and were then put back into home-  
14 cage and after 10 min they received the second injection and were immediately placed  
15 into the open-field test arena (Fig. 1C). There were 2 VEH+VEH groups, when rats  
16 received 2 injections of VEH; 2 VEH+MOR, in which the first injection was VEH and the  
17 second was morphine at a dose of 10 mg/kg (MOR); 2 APO+VEH groups in which the first  
18 injection was apomorphine at 0.05 mg/kg (APO) and the second was VEH and; 2  
19 APO+MOR groups in which the first injection was apomorphine at 0.05 mg/kg (APO) and  
20 the second was MOR at 10 mg/kg. For the final test, all groups received VEH as the first  
21 injection, and for the second injection, received either VEH or MOR. The experimental  
22 groups (n=7 for each group) were labeled in terms of (a) their induction treatments / (b)  
23 their final test treatments. The treatment groups are summarized and presented in Table  
24 1.

1 2.6. Experiment 2: The effect of APO on the expression of morphine-induced conditioned  
2 locomotor stimulation.

3 The protocol described in the general experimental procedure was followed. In the  
4 induction phase, the rats received the first injection and were put back into home-cage.  
5 After 10 min they received the second injection and were immediately placed into the open-  
6 field test arena. There were 2 VEH+VEH groups, in which rats received 2 injections of VEH,  
7 and 2 VEH+MOR groups, in which the first injection was VEH and the second was  
8 morphine at a dose of 10 mg/kg(MOR). After completion of the 5 sessions induction phase,  
9 the VEH/MOR groups developed a strong locomotor stimulant response and the next day  
10 a conditioning test was conducted to assess the effects of APO on the expression of the  
11 conditioned MOR response. For the conditioning test, the two VEH/MOR groups were  
12 divided into 2 groups (n=7) with comparable locomotor stimulant responses, and one group  
13 received two injections of VEH, while the other group received APO as the first injection  
14 and VEH as the second injection. The experimental groups (n=7 for each group) were  
15 labeled in terms of (a) their induction treatments / (b) their final test treatments. The  
16 treatment groups are summarized and presented in Table 2A.

17  
18

19 2.7. Experiment 3: Acute 1-day experiment.

20

21 After the 3 days of the habituation phase, that was used in the previous two experiments,  
22 the rats received the first treatment injection and were put back into home-cage and after  
23 10 min. they received the second treatment injection and immediately were placed into the  
24 open-field test arena. All groups received a one day 5 min test and immediately upon  
25 completion of the test the animals were euthanized, perfused and immunohistochemistry was  
26 performed. The objective of this experiment was to assess the effects of morphine on  
27 ERK activity in the VTA and NAc before

1 locomotor stimulant effects were induced. There were 4 groups (n=6 for each group):  
2 VEH+VEH group in which rats received 2 injections of VEH; VEH+MOR group in which the  
3 first injection was VEH and the second was morphine at 10 mg/kg (MOR); APO+VEH in  
4 which the first injection was apomorphine at 0.05 mg/kg (APO) and the second injection  
5 was VEH; and APO+MOR group where the first injection was APO and the second injection  
6 was MOR. The treatment groups are summarized and presented in Table 2B.

7  
8

## 9 2.8. Immunohistochemistry

10  
11 The immunohistochemistry protocol was conducted as previously described by Crespo and  
12 co-workers [8] and Dias and co-workers [11]. In brief, rats were rapidly anaesthetized by  
13 intraperitoneal injection of urethane (15 ml/kg; Sigma Aldrich, Saint Louis, MO, USA) prior  
14 to intracardiac perfusion with 4% paraformaldehyde (500 ml) in 0.1 M sodium phosphate  
15 buffer (pH 7.4) for 30 min. The encephalon of each rat was removed and post-fixed for 2 h  
16 in 4% paraformaldehyde solution before transferring to 20% sucrose in 0.1 M sodium  
17 phosphate buffer (pH 7.4) for 48 h at 4 °C. Each brain was placed on an aluminum paper  
18 base and cryoprotected using a solution of water-soluble glycols and resins (Tissue Tek®  
19 O. C. T. Sakura Finetek®, USA). The brain tissue was then frozen and maintained in liquid  
20 nitrogen until being processed for immunohistochemistry.

21 The encephalic areas sampled for the immunohistochemical analysis were ventral  
22 tegmental area (approximately -4.80 mm to -5.04 mm from bregma) and nucleus  
23 accumbens (core and shell approximately +2.52 mm to +2.76 mm from bregma). The  
24 coordinates adopted as the reference were obtained from Paxinos and Watson [27]. Four  
25 slices were collected sequentially from each brain structure of each animal and sectioned  
26 in a cryostat (Zeiss, Germany) at a thickness

1 of 30  $\mu\text{m}$ . The sections were placed onto previously gelatinized glass microscope slides to  
2 allow fixation. For immunohistochemistry, sections were rinsed three times for 10 min in  
3 phosphate- buffered saline (PBS) and placed in blocking buffer (3% normal goat serum  
4 and 0.25% Triton X-100 in PBS) for 1 h at 22 °C. The sections were then treated with a  
5 freshly prepared solution of hydrogen peroxide (3% vol/vol) in chilled absolute methanol  
6 for 10 min at 22 °C. Sections were then incubated for 24 h at 4 °C in 1:500 dilution of anti-  
7 phosphorylated-ERK protein antibody diluted in blocking buffer (Cat # 9101, Cell Signaling  
8 Technology®, Boston, MA, USA) as previously described. After the end of the incubation  
9 time, sections were washed 3 times for 10 min each in PBS and incubated at 22 °C with a  
10 1:100 dilution of biotinylated goat anti-rabbit IgGsecondary antibody (BA-1000, Vector  
11 Laboratories®, CA, USA) in 1% normal goat serum and 0.25% Triton X-100 in PBS.  
12 Sections were then washed three times for 10 min in PBS and processed using an ABC  
13 Elite kit (Vector Laboratories ®, Burlingame, CA, USA). In the next step, sections were  
14 washed again in PBS and processed with a DAB substrate kit for peroxidase (SK-4100,  
15 Vector Laboratories®, Burlingame, CA, USA) and incubated in DAB substrate  
16 simultaneously and timed precisely at 22 °C for 3 min for color development of signal  
17 intensity. Sections were lightly counterstained with hematoxylin. After drying, the slides  
18 were mounted withDPX (Sigma ®, USA).

19 Photomicrographs of the brain sections were obtained using a CCD camera (Nikon  
20 Photometrics Cool Snap) attached to a light microscope (Nikon 80i, USA). Digital images  
21 were obtained at low magnification (10X) from regions of the brain identified from the  
22 Paxinos and Watson [27] rat brain atlas and the labeled nuclei from each brain structure  
23 of each animal were quantified using the Image J® software "multi-point" tool. The number  
24 of labeled neurons was counted bilaterally from four sections and the average count of  
25 labeled neurons for these sections

1 was used as the score for each animal. The number of p-ERK-labeled cells was normalized  
2 to the area of the quadrant counted. In order to demonstrate details of the ERK-labeled  
3 cells, specific sections were photographed at higher magnification (40X).

4 Negative control slices were incubated with normal serum instead of primary antibody  
5 (data not shown). In order to minimize any potential bias in the scoring, two experimenters  
6 unaware of the treatment groups independently performed the labeled nuclei counts. The  
7 counts obtained by each experimenter for each encephalic area and for each group were  
8 compared using Student's t-tests and no statistically significant differences ( $p > 0.05$ ) were  
9 found between groups.

10  
11

## 12 2.9. Statistics

13

14 After the determination of normal distributions of data using the Shapiro-Wilk normality  
15 test, considering the induction treatment phase, a repeated two-way analysis of variance  
16 (ANOVA) was used in order to determine the group effect, day effect, as well as the  
17 interactions between both factors. When a significant effect of interaction was recorded,  
18 data were further statistically evaluated by one-way ANOVA followed by Tukey's test with  
19  $p < 0.05$  as the criterion for statistical significance. For the final test, a one-way ANOVA was  
20 performed followed by Tukey's post-hoc tests using  $p < 0.05$  as the criterion for statistical  
21 significance. For the morphine induced ERK activation analysis, a one-way ANOVA was  
22 performed followed by Tukey's post-hoc tests using  $p < 0.05$  as the criterion for statistical  
23 significance.

### 1 3. Results

2

3 3.1 Experiment 1: Effect of a low dose of apomorphine pretreatment on the acquisition of  
4 morphine-induced locomotor sensitization, conditioning, and ERK activity in the VTA and  
5 NAc.

6 Figure 2 shows the locomotor activity over the course of days 1 to 5 of the induction phase.

7 According to a repeated two-way ANOVA, there were significant effects of interaction of

8 groupsX days [F (28, 192) = 12.75;  $p < 0.01$ ], of groups [F (7, 48) = 45.30;  $p < 0.01$ ], and of

9 days of treatment [F (4, 192) = 10.10;  $p < 0.01$ ]. According to a one-way ANOVA followed

10 by Tukey's post hoc test, to further analyze the interaction of group X days, on day 1 there

11 was a significant difference among the groups [F (7, 48) = 11.11;  $p < 0.01$ ], the groups that

12 received apomorphine at a dose of 0.05 mg/kg before being placed in the arena had lower

13 locomotion than all other groups (Tukey's post hoc test;  $p < 0.05$ ). There was no significant

14 difference between the groups that received vehicle only and the groups that received

15 morphine at a dose of 10 mg/kg (Tukey's posthoc test;  $p > 0.05$ ). On day 2 [F (7, 48) = 9.0;

16  $p < 0.01$ ], the results showed that the VEH+MOR groups had higher locomotion than the

17 APO groups ( $p < 0.05$ ). On day 3 [F (7, 48) = 32.50;  $p < 0.01$ ], day 4 [F (7, 48) = 51.70;

18  $p < 0.01$ ], and day 5 [F (7, 48) = 84.0;  $p < 0.01$ ], the VEH+MOR groups

19 had higher locomotion than all other groups ( $p < 0.05$ ). The APO, APO+MOR and

20 APO+VEH groups had significantly lower locomotion than all other groups ( $p < 0.01$ ). For

21 both VEH+MOR groups, there was a significant increase in locomotion across the days of

22 the induction phase ( $p < 0.01$ ).

23 Figure 3 shows the locomotor activity during the final test. According to a one-way ANOVA,

24 there was a significant difference among the groups [F (7, 48) = 106.43;  $p < 0.01$ ], and the

25 VEH+MOR/VEH+MOR and VEH+MOR/VEH+VEH groups had higher locomotion than the

26 other groups (Tukey's post hoc test;  $p < 0.01$ ). These findings suggest that the conditioned



1 morphine locomotor stimulant response was equivalent to the morphine induced locomotor  
2 stimulant response. The APO+MOR/VEH+MOR group had the highest locomotion of all  
3 other groups ( $p < 0.05$ ), except the APO+MOR/VEH+VEH group, which was not  
4 significantly different ( $p > 0.05$ ). There was no significant difference between the  
5 APO+MOR/VEH+MOR group and the APO+MOR/VEH+VEH group ( $p > 0.05$ ). In addition,  
6 rats from the APO+MOR/VEH+VEH group had higher locomotion than the  
7 VEH+VEH/VEH+VEH group ( $p < 0.05$ ). There was neither a significant difference among  
8 the APO+MOR/VEH+VEH-, APO+VEH/VEH+VEH-, APO+VEH/VEH+MOR- and  
9 VEH+VEH/VEH+MOR-treated groups ( $p > 0.05$ ), nor among the VEH+/VEH+VEH-,  
10 APO+VEH/VEH+VEH-, APO+VEH/VEH+MOR- and  
11 VEH+VEH/VEH+MOR-treated groups ( $p > 0.05$ ).

12  
13 Figure 4 shows the values for ERK immunoreactive nuclei for the ventral tegmental area.  
14 Regarding the number of Phosphor ERK protein immunoreactive nuclei (Fig. 4A),  
15 according to a one-way ANOVA, there was a significant effect of the treatment [ $F(7, 48) =$   
16  $89.01$ ;  $p < 0.01$ ]. VEH+MOR/VEH+MOR and VEH+MOR/VEH+VEH groups had a  
17 significantly higher number of phosphors ERK immunoreactive nuclei than all other groups  
18 (Tukey's post hoc test;  $p < 0.01$ ). Figures 4B-D show examples of the sections used for  
19 counting phosphorylated-ERK- immunoreactive cells in the VTA following VEH, MOR, and  
20 APO administration.

21 Figure 5 shows the values for ERK immunoreactive nuclei in the nucleus accumbens.  
22 Regarding the number of Phosphor ERK immunoreactive nuclei (Fig. 5A), according to a  
23 one-way ANOVA, there were significant differences among the treatments [ $F(7, 48) =$   
24  $94.91$ ;  $p < 0.01$ ]. VEH+MOR/VEH+MOR and VEH+MOR/VEH+VEH groups had a  
25 significantly higher number of phosphor ERK immunoreactive nuclei than all the other  
26 groups (Tukey's post hoc test;  $p <$

1 0.01). Figures 4B-D show examples of the sections used for counting phosphorylated-  
2 ERK- immunoreactive cells in the NAc following VEH, MOR, and APO administration.

3  
4  
5 3.2. Experiment 2: Effect of a low dose of apomorphine on the expression of conditioned  
6 morphinelocomotor stimulant effects, and ERK activity in VTA and NAc.

7 Figure 6 shows the locomotor activity over the course of days 1-5 induction phase.  
8 According to a repeated two-way ANOVA, there was a significant effect of interaction  
9 between groups X days [ $F(12, 96) = 15.34; p < 0.01$ ], of groups [ $F(3, 24) = 33.32; p < 0.01$ ],  
10 and of days of treatment [ $F(4, 96) = 9.52; p < 0.01$ ]. According to one-way ANOVAs followed  
11 by Tukey's post hoc test on day 1, there was no significant difference among the groups [ $F$   
12  $(3, 24) = 0.30; p > 0.05$ ]. On day 2 [ $F(3, 24) = 30.0; p > 0.05$ ], day 3 [ $F(3, 24) = 33.0;$   
13  $p < 0.01$ ], day 4 [ $F(3, 24) = 23.45;$   
14  $p < 0.01$ ], and day 5 [ $F(3, 24) = 37.61; p < 0.01$ ], the VEH+MOR-treated groups had  
15 significantly higher locomotion than all of the VEH-treated groups (Tukey's post hoc test;  
16  $p < 0.01$ ).

17 Figure 7 shows the locomotor activity during the final test. According to a one-way ANOVA,  
18 showed that there was a significant effect of treatment [ $F(3, 24) = 120.0; p < 0.01$ ]. The  
19 VEH+MOR/VEH+VEH-treated group had the highest locomotion score (Tukey's post hoc  
20 test;  $p < 0.01$ ), indicative of a strong conditioned morphine response. There was no  
21 significant difference between the VEH+VEH/VEH+VEH- and VEH+MOR/APO+VEH-  
22 treated groups ( $p > 0.05$ ), indicating a complete blockade of the expression of the morphine-  
23 induced conditioned response.

24 Figure 8 shows the ERK values for the ventral tegmental area and for the nucleus  
25 accumbens. Regarding the number of Phosphor ERK protein immunoreactive nuclei for  
26 the VTA (Fig. 8A), according to a one-way ANOVA, there was a significant effect of

1 treatment [F (3, 24) = 8.0;  $p < 0.01$ ]. The VEH+MOR/VEH+VEH-treated group had a higher  
2 number of phosphors ERK immunoreactive nuclei than all other groups (Tukey's post hoc  
3 test;  $p < 0.01$ ) and the APO pretreatment blocked the ERK response. Considering the  
4 number of Phosphor ERK protein immunoreactive nuclei for the NAc (Fig. 8C), according  
5 to a one-way ANOVA, there was a significant effect of treatment [F (3, 24) = 11.50;  $p < 0.01$ ].  
6 The VEH+MOR/VEH+VEH-treated group had a higher number of phosphors ERK  
7 immunoreactive nuclei than all other groups (Tukey's post hoc test;  $p < 0.01$ ), and the APO  
8 pretreatment blocked the ERK response. Figures 8B and 8D show examples of the sections  
9 used for counting phosphorylated-ERK-immunoreactive cells in the VTA and NAc following  
10 VEH, MOR, and APO administration.

11  
12

### 13 3.3. Experiment 3: Acute 1-day experiment.

14  
15 Figure 9 shows the locomotor activity during the one-day test session. According to a one-  
16 way ANOVA, there was a significant effect of treatment [F (3, 20) = 69.30;  $p < 0.01$ ]. The  
17 VEH+VEH- and VEH+MOR-treated groups had higher locomotion than the APO+VEH-  
18 and APO+MOR-treated groups (Tukey's post hoc test;  $p < 0.01$ ). Importantly, the  
19 VEH+VEH and VEH+MOR groups did not significantly differ in activity levels ( $p > 0.05$ ).

20 Figure 10 shows the ERK values for the ventral tegmental area and for the nucleus  
21 accumbens. Regarding the number of Phosphor ERK protein immunoreactive nuclei for  
22 the VTA (Fig. 10A), according to a one-way ANOVA, there was a significant effect of  
23 treatment [F (3, 20)

24 = 48.0;  $p < 0.01$ ]. The VEH+MOR-treated group had a higher number of phosphors ERK  
25 immunoreactive nuclei than all the other groups ( $p < 0.01$ ). Regarding the number of  
26 Phosphor ERK protein immunoreactive nuclei for the NAc (Fig. 10C), according to a one-  
27 way ANOVA, there was a significant effect of treatment [F (3, 20) = 55.31;  $p < 0.01$ ]. The

1 VEH+MOR-treated group had a higher number of phosphors ERK immunoreactive nuclei  
2 than all the other groups (Tukey'spost hoc test;  $p<0.01$ ). Figures 10B and 10D show  
3 examples of the sections used for counting phosphorylated-ERK-immunoreactive cells in  
4 the VTA and NAc following VEH, MOR, and APOadministration.

5  
6

#### 7 **4. Discussion**

8  
9 In two separate experiments, morphine (10.0 mg/kg) administered immediately prior to 5  
10 min placements in an open field test arena generated a marked hyperactivity. In both  
11 experiments, the first morphine treatment had no significant effect on activity level as  
12 compared to vehicle controls. Hyperactivity began to emerge by the second treatment with  
13 morphine and steadily increased with repeated treatments. In the first experiment,  
14 morphine groups that were given a pretreatment with apomorphine (0.05 mg/kg) 10 min  
15 before the morphine injection had a substantial reduction in spontaneous activity during  
16 the first test session and that reduction persisted unchanged over the course of the five  
17 test sessions. Vehicle-treated groups that received the apomorphine pretreatment 10 min  
18 before placement in the test arena also had a reduction in spontaneous activity that was  
19 indistinguishable from the combined apomorphine/morphine treatment. After completion  
20 of these five test sessions, the rats from different groups were given an additional test to  
21 assess if the drug induced changes in spontaneous activity were conditioned to the test  
22 environment contextual cues. In the conditioning test the groups that received morphine  
23 without the apomorphine pretreatment remained hyperactive regardless of whether they  
24 received either morphine or vehicle immediately pretest or not, indicating that the  
25 morphine-induced hyperactivity response was conditioned to the test arena cues. In  
26 contrast, the groups that had received the apomorphine pretreatment with or without  
27 morphine were hypoactive throughout the

1 five-day acquisition phase and when tested for a conditioned drug response exhibited either  
2 vehiclelevel activity or were slightly more active than the vehicle-treated groups. Whereas  
3 morphine stimulant effects were conditioned to test arena cues the apomorphine response  
4 inhibitory effectswere not conditioned.

5 While the initial acquisition experiment established that apomorphine prevented the  
6 development of sensitization and conditioning of morphine-induced hyperactivity, the  
7 second experiment was conducted to determine if the apomorphine would also block the  
8 expression of the morphine conditioned stimulant effect. As in the first acquisition  
9 experiment, morphine administered immediately before testing did not elicit a hyperactivity  
10 response on the first test session, but hyperactivity developed progressively over the five  
11 test sessions. These morphine- treated groups were then tested for a conditioned stimulant  
12 response. One morphine-treated groupreceived the apomorphine pretreatment 10 min  
13 pretest and the other morphine group received vehicle 10 min pretest. The morphine-  
14 vehicle pretreatment group exhibited a strong conditioned stimulant response whereas the  
15 morphine-treated group given the apomorphine pretreatment did not have a conditioned  
16 stimulant response, and its behavioral response was equivalent to the vehicle-treated  
17 control group. Thus, apomorphine prevented both the acquisition and expression of the  
18 morphine conditioned stimulant response.

19 The rationale for using a test protocol, in which the behavioral testing is done immediately  
20 post-injection and is brief (5 min), was developed in our recent study [8]. The behavioral  
21 and ERKeffects of the morphine treatments in the vehicle morphine groups in the present  
22 report essentiallyreplicate this previous report. While no unpaired morphine groups were  
23 used in our experiments, we have shown previously [11] that the same morphine  
24 treatments (10.0 mg/kg) given unpaired 15 min after a 5 min test session are not  
25 significantly different from vehicle injections given

1 immediately pretest and do not generate hyperactivity effects to test either cues or  
2 morphine. Thus, the immediate morphine pretest injection stimulant effects are not context  
3 independent sensitization effects.

4 Ostensibly, the present conditioning findings do not readily fit into a Pavlovian drug  
5 conditioning framework. That is, morphine did induce a strong behavioral stimulant  
6 response but initially did not elicit an unconditioned behavioral stimulant drug response so  
7 that there was no initial unconditioned behavioral response to be associated with the test  
8 arena cues. The ERK findings, however, appear to have relevance to the morphine  
9 conditioning effects. That is, while there was no behavioral response elicited by morphine  
10 in the first arena test session in the first two experiments, the ERK findings obtained in  
11 experiment three, the acute treatment-related experiment, showed that there was a strong  
12 ERK response evoked during the first test session, in which no behavioral response was  
13 elicited. From a conditioning perspective, the ERK response could be considered an  
14 unconditioned response. This unconditioned response could be viewed in terms of either  
15 instrumental or Pavlovian conditioning. When considered in terms of instrumental  
16 conditioning, the ERK activation in the VTA could be seen as the elicitation of a reward  
17 response that served to reinforce and enhance the preceding behavioral activity and  
18 possibly even exerting this effect on the behavioral trace during the post-trial consolidation  
19 process. In line with this latter consideration, we have demonstrated in several previous  
20 reports [8, 9, 11] that morphine (10 mg/kg) administered immediately after a brief 5 min  
21 exposure to a test arena induced a marked behavioral stimulant response, whereas the  
22 same morphine treatment given 15 min after removal from the test arena was without effect  
23 [11]. This analysis seems to align with an operant conditioning process wherein morphine  
24 administered after a lever press response increases the frequency of the response as the  
25 morphine reward effect is associated with the lever press response

1 trace. The response trace to a lever press response is brief [15]. In that the lever press  
2 response trace is occurring in the context of ongoing behavior responses, the lever press  
3 trace would quickly be replaced by subsequent occurring response traces, so the  
4 reinforcement necessarily needs to be precisely timed to immediately follow the emission of  
5 the lever press response. In the present open-field testing arrangement, the initial 5 min  
6 sensory/motor exploration of the open-field test arena occurs concurrently with the  
7 activation of VTA and NAc as manifested in the ERK activation. Thus, it can be argued that  
8 this apparent activation of reward systems during this initial locomotor activation serves to  
9 reinforce and strengthen this behavior analogous to a conventional instrumental  
10 reinforcement [3, 4].

11 In contrast, to conventional instrumental conditioning in which the experimenter delivers  
12 the reward following a specified response in this open-field test drug treatment  
13 arrangement, the drug treatment is administered so the drug induced activation of reward  
14 systems occurs during a prepotent locomotor exploration response. This arrangement  
15 seemingly creates the opportunity for a Pavlovian/Instrumental conditioning fusion. The  
16 drug induced Pavlovian unconditioned response of activation of reward brain areas  
17 becomes associated with the test arena cues and consequently can reinforce and  
18 strengthen the associated occurring behavior. Consequently, the conditioning effects  
19 occur as both locomotor stimulation and ERK activation of selected brain reward areas.  
20 This creates an interesting situation in which the non-drug conditioning tests would appear  
21 to be able to function as additional drug treatment sessions. Indeed, in the conditioning tests  
22 we performed either with or without morphine, both the locomotor and ERK responses  
23 evoked were equivalent. When considered in this way, it would seem possible that the initial  
24 drug induced behavior could, following conditioning, be maintained independent from  
25 further drug use. This is

1 an issue of both theoretical importance as well as pragmatic importance from the  
2 perspective of drug addiction.

3 In the groups treated with apomorphine, a strong inhibitory response was elicited with the  
4 first drug treatment so this inhibitory response seemingly could become associated with  
5 the arena test cues. This inhibitory response, however, was not conditioned. In that low  
6 dose apomorphine decreases dopamine activity broadly [7], including sensory/motor and  
7 reward systems, the absence of inhibitory conditioning is not surprising as attention to arena  
8 cues would be severely suppressed and essentially would be equivalent to not being placed  
9 in the test arena. Indeed, the performance of the apomorphine-treated groups in the  
10 conditioning tests did not differ from the performance of the vehicle groups during their first  
11 test session.

12 The ERK results were consistent with the behavioral results in that the morphine stimulant  
13 effects were associated with a marked ERK response in the VTA and NAc, and these ERK  
14 responses were impaired by the apomorphine pretreatments that eliminated the morphine  
15 stimulant responses. Thus, the apomorphine pretreatments eliminated both the behavioral  
16 effects of the morphine treatment, as well as the morphine activation of the ERK response  
17 in the VTA and NAc. In all three experiments, pretreatments with apomorphine at low dose  
18 prevented morphine-induced increases in ERK activity in the VTA and NAc. Together,  
19 these findings are consistent with the dependence of morphine GABAergic disinhibitory  
20 effects on the level of ongoing activity in the dopaminergic system.



## 1 **Acknowledgements**

2  
3 This research was supported by UENF and FAPERJ (grant E-26/211.903/2021). J.B.L.J.,  
4  
5 L.G.S.C.C. and J.M.M.B. are recipients of fellowships from CAPES-Brazil. M.P.C., R.I.S.  
6 and  
7  
8 N.C.C. are CNPq research fellows.  
9

10

11

12

## 12 **References**

13

14 [1] J.P. Adams, J.D. Sweatt, Molecular psychology: roles for the ERK MAP kinase  
15 cascade in memory, *Annual Review of Pharmacology and Toxicology*. 42 (2002) 135-  
16 163. [https://doi: 10.1146/annurev.pharmtox.42.082701.145401](https://doi:10.1146/annurev.pharmtox.42.082701.145401).

17 [2] G.K. Aghajanian, B.S. Bunney, Central dopaminergic neurons: neurophysiological  
18 identification and responses to drugs. In: Snyder S, Usdin E, editors. *Frontiers in*  
19 *Catecholamine Research*. New York: Pergamon Press; 1973. p. 643-648.

20 [3] R.J. Beninger, The role of dopamine in locomotor activity and learning, *Brain*  
21 *Research*. 287 (1983)173-196. [https://doi: 10.1016/0165-0173\(83\)90038-3](https://doi:10.1016/0165-0173(83)90038-3).

22 [4] R.J. Beninger, R. Miller, Dopamine D1-like receptors and reward-related incentive  
23 learning, *Neuroscience and Biobehavioral Reviews*. 22 (1998) 335-345. [https://doi:](https://doi:10.1016/s0149-7634(97)00019-5)  
24 [10.1016/s0149-7634\(97\)00019-5](https://doi:10.1016/s0149-7634(97)00019-5).

25 [5] A. Borgkvist, E. Valjent, E. Santini, D. Hervé, J.A. Girault, G. Fisone, Delayed, context-  
26 and dopamine D1 receptor-dependent activation of ERK in morphine-sensitized mice,  
27 *Neuropharmacology*. 55 (2008) 230-237. [https://doi: 10.1016/j.neuropharm.2008.05.028](https://doi:10.1016/j.neuropharm.2008.05.028).

- 1 [6] M.T. Brown, K.R. Tan, E.C. O'Connor, I. Nikonenko, D. Muller, C. Lüscher, Ventral  
2 tegmentalarea GABA projections pause accumbal cholinergic interneurons to enhance  
3 associative learning, *Nature*. 492 (2012) 452-456. [https://doi: 10.1038/nature11657](https://doi.org/10.1038/nature11657).
- 4 [7] R.J. Carey, G. DePalma, E. Damianopoulos, A. Hopkins, A. Shanahan, C.P. Müller,  
5 J.P. Huston, Dopaminergic and serotonergic autoreceptor stimulation effects are  
6 equivalent and additive in the suppression of spontaneous and cocaine induced  
7 locomotor activity, *Brain Research*. 1019 (2004) 134-143. [https://doi:  
8 10.1016/j.brainres.2004.05.091](https://doi.org/10.1016/j.brainres.2004.05.091).
- 9 [8] L.G.S.C. Crespo, J.B. Leite Júnior, J.M. de Mello Bastos, R.I. Samuels, N.C. Coimbra,  
10 R.J. Carey, M.P. Carrera, Context evoked morphine conditioned effects can be equivalent  
11 to morphineinduced drug effects in terms of behavioral response and ERK activation in  
12 reward associated subcortical brain structures, *Pharmacology, Biochemistry and  
13 Behavior*. 214 (2022) 173356. [https://doi: 10.1016/j.pbb.2022.173356](https://doi.org/10.1016/j.pbb.2022.173356).
- 14 [9] J.M. de Mello Bastos, J.B. Leite Junior, R.I. Samuels, R.J. Carey, M.P. Carrera, Post-  
15 trial low dose apomorphine prevents the development of morphine sensitization,  
16 *Behavioural Brain Research*. 380 (2020) 112398. [https://doi: 10.1016/j.bbr.2019.112398](https://doi.org/10.1016/j.bbr.2019.112398).
- 17 [10] G. Di Chiara, M.L. Porceddu, W. Fratta, G.L. Gessa, Postsynaptic receptors are not  
18 essentialfor dopaminergic feedback regulation, *Nature*. 267 (1977) 270-272. [https://doi:  
19 10.1038/267270a0](https://doi.org/10.1038/267270a0).
- 20 [11] F.P. Dias, L.G.S.C. Crespo, J.B. Leite Junior, R.I. Samuels, N.C. Coimbra, R.J.  
21 Carey, M.P. Carrera, Morphine reward effects and morphine behavioral sensitization: The  
22 adventitious association of morphine activation of brain reward effects with ongoing  
23 spontaneous activity,

1 Pharmacology, Biochemistry and Behavior. 209 (2021) 173244. [https://doi:  
2 10.1016/j.pbb.2021.173244.](https://doi:10.1016/j.pbb.2021.173244)

3 [12] H.L. Fields, G.O. Hjelmstad, E.B. Margolis, S.M. Nicola, Ventral tegmental area  
4 neurons in learned appetitive behavior and positive reinforcement, Annual Review of  
5 Neuroscience. 30 (2007) 289-316. [https://doi:  
6 10.1146/annurev.neuro.30.051606.094341.](https://doi:10.1146/annurev.neuro.30.051606.094341)

7 [13] A.N. Fricks-Gleason, J.F. Marshall, Role of dopamine D1 receptors in the activation  
8 of nucleus accumbens extracellular signal-regulated kinase (ERK) by cocaine-paired  
9 contextual cues, Neuropsychopharmacology. 36 (2011) 434-444. [https://doi:  
10 10.1038/npp.2010.174.](https://doi:10.1038/npp.2010.174)

11 [14] J.A. Girault, E. Valjent, J. Caboche, D. Hervé, ERK2: a logical AND gate critical for  
12 drug- induced plasticity? Current Opinion in Pharmacology. 7 (2007) 77-85. [https://doi:  
13 10.1016/j.coph.2006.08.012.](https://doi:10.1016/j.coph.2006.08.012)

14 [15] G.R. Grice, The relation of secondary reinforcement to delayed reward in visual  
15 discrimination learning, Journal of Experimental Psychology. 38 (1948) 1-16. [https://doi:  
16 10.1037/h0061016.](https://doi:10.1037/h0061016)

17 [16] S.W. Johnson, R.A. North, Opioids excite dopamine neurons by hyperpolarization of  
18 local interneurons, Journal of Neuroscience. 12 (1992) 483-488. [https://doi:  
19 10.1523/JNEUROSCI.12-02-00483.1992.](https://doi:10.1523/JNEUROSCI.12-02-00483.1992)

20 [17] B. Juarez, M.H. Han, Diversity of Dopaminergic Neural Circuits in Response to Drug  
21 Exposure, Neuropsychopharmacology. 41 (2016) 2424-2446. [https://doi:  
22 10.1038/npp.2016.32.](https://doi:10.1038/npp.2016.32)

23 [18] R. Keiflin, H.J. Pribut, N.B. Shah, P.H. Janak, Ventral Tegmental Dopamine Neurons  
24 Participate in Reward Identity Predictions, Current Biology. 29 (2019) 93-103.e3.

1 <https://doi: 10.1016/j.cub.2018.11.050>.

- 1 [19] S. Kim, J.H. Kim, Time-dependent change of ERK phosphorylation levels in the  
2 nucleus accumbens during withdrawals from repeated cocaine, *Neuroscience Letters*.  
3 436 (2008) 107-110. [https://doi: 10.1016/j.neulet.2008.02.068](https://doi.org/10.1016/j.neulet.2008.02.068).
- 4 [20] J.B. Leite Júnior, J.M. de Mello Bastos, R.I. Samuels, R.J. Carey, M.P. Carrera,  
5 Reversal of morphine conditioned behavior by an anti-dopaminergic post-trial drug  
6 treatment during re- consolidation, *Behavioural Brain Research*. 359 (2019) 771-782.  
7 [https://doi: 10.1016/j.bbr.2018.08.009](https://doi.org/10.1016/j.bbr.2018.08.009).
- 8 [21] K.A. Leite-Morris, E.Y. Fukudome, M.H. Shoeb, G.B. Kaplan, GABA(B) receptor  
9 activation in the ventral tegmental area inhibits the acquisition and expression of opiate-  
10 induced motor sensitization, *Journal of Pharmacology and Experimental Therapeutics*.  
11 308 (2004) 667-78. [https://doi: 10.1124/jpet.103.058412](https://doi.org/10.1124/jpet.103.058412).
- 12 [22] L. Lu, E. Koya, H. Zhai, B.T. Hope, Y. Shaham, Role of ERK in cocaine addiction,  
13 *Trends in Neurosciences*. 29 (2006) 695-703. [https://doi: 10.1016/j.tins.2006.10.005](https://doi.org/10.1016/j.tins.2006.10.005).
- 14 [23] R.T. Matthews, D.C. German, Electrophysiological evidence for excitation of rat  
15 ventral tegmental area dopamine neurons by morphine, *Neuroscience*. 11 (1984) 617-  
16 625. [https://doi: 10.1016/0306-4522\(84\)90048-4](https://doi.org/10.1016/0306-4522(84)90048-4).
- 17 [24] C. Missale, S.R. Nash, S.W. Robinson, M. Jaber, M.G. Caron, Dopamine  
18 receptors: from structure to function, *Physiological Reviews*. 78 (1998) 189-225. [https://doi: 10.1152/physrev.1998.78.1.189](https://doi.org/10.1152/physrev.1998.78.1.189).
- 19  
20 [25] A.G. Nasello, C. Machado, J.F. Bastos, L.F. Felicio, Sudden darkness induces a  
21 high activity-low anxiety state in male and female rats, *Physiology & Behavior*. 63  
22 (1998) 451-454. [https://doi.org/10.1016/s0031-9384\(97\)00462-9](https://doi.org/10.1016/s0031-9384(97)00462-9).

- 1 [26] J.L. Neisewander, M.T. Bardo, Expression of morphine-conditioned hyperactivity is  
2 attenuated by naloxone and pimozide, *Psychopharmacology*. 93 (1987) 314-319.  
3 [https://doi: 10.1007/BF00187249](https://doi.org/10.1007/BF00187249).
- 4 [27] G. Paxinos, C. Watson, *The Rat Brain in Stereotaxic Coordinates*, 2005. 6th edition.  
5 Elsevier Academic Press, New York.
- 6 [28] K.R. Powell, S.G. Holtzman, Parametric evaluation of the development of  
7 sensitization to the effects of morphine on locomotor activity, *Drug and Alcohol*  
8 *Dependence*. 62 (2001) 83-90. [https://doi: 10.1016/s0376-8716\(00\)00167-8](https://doi.org/10.1016/s0376-8716(00)00167-8).
- 9 [29] K. Radwanska, J. Caboche, L. Kaczmarek, Extracellular signal-regulated kinases  
10 (ERKs) modulate cocaine-induced gene expression in the mouse amygdala, *European*  
11 *Journal of Neuroscience*. 22 (2005) 939-948. [https://doi: 10.1111/j.1460-](https://doi.org/10.1111/j.1460-9568.2005.04286.x)  
12 [9568.2005.04286.x](https://doi.org/10.1111/j.1460-9568.2005.04286.x).
- 13 [30] F.V. Sanguedo, C.V. Dias, F.R. Dias, R.I. Samuels, R.J. Carey, M.P. Carrera,  
14 Reciprocal activation/inactivation of ERK in the amygdala and frontal cortex is  
15 correlated with the degree of novelty of an open-field environment,  
16 *Psychopharmacology*. 233 (2016) 841-850. [https://doi: 10.1007/s00213-015-4163-z](https://doi.org/10.1007/s00213-015-4163-z).
- 17 [31] F.V. Sanguedo, F.R. Dias, E. Bloise, I.C. Cespedes, A. Giral-di-Guimarães, R.I.  
18 Samuels, R.J. Carey, M.P. Carrera, Increase in medial frontal cortex ERK activation  
19 following the induction of apomorphine sensitization, *Pharmacology, Biochemistry and*  
20 *Behavior*. 118 (2014) 60-68. [https://doi: 10.1016/j.pbb.2013.12.020](https://doi.org/10.1016/j.pbb.2013.12.020).
- 21 [32] F.V. Sanguedo, A.M. Figueiredo, R.J. Carey, R.I. Samuels, M.P. Carrera, ERK  
22 activation in the prefrontal cortex by acute apomorphine and apomorphine  
23 conditioned contextual stimuli,

1 Pharmacology, Biochemistry and Behavior. 159 (2017) 76-83. [https://doi:](https://doi:10.1016/j.pbb.2017.07.011)  
2 10.1016/j.pbb.2017.07.011.

3 [33] F.V. Sanguedo, R.I. Samuels, R.J. Carey, M.P. Carrera, Medial prefrontal cortex  
4 ERK and conditioning: Evidence for the association of increased medial prefrontal  
5 cortex ERK with the presence/absence of apomorphine conditioned behavior using a  
6 unique post-trial conditioning/extinction protocol, Behavioural Brain Research. 365  
7 (2019) 56-65. <https://doi:10.1016/j.bbr.2019.02.022>.

8 [34] K.R. Tan, C. Yvon, M. Turiault, J.J. Mirzabekov, J. Doehner, G. Labouèbe, K.  
9 Deisseroth,  
10 K.M. Tye, C. Lüscher, GABA neurons of the VTA drive conditioned place aversion, Neuron.  
11 73 (2012) 1173-1183. [https://doi: 10.1016/j.neuron.2012.02.015](https://doi:10.1016/j.neuron.2012.02.015).

13 [35] T.M. Tzschentke, W.J. Schmidt, N-methyl-D-aspartic acid-receptor antagonists  
14 block morphine-induced conditioned place preference in rats, Neuroscience Letters. 193  
15 (1995) 37-40. [https://doi: 10.1016/0304-3940\(95\)11662-g](https://doi:10.1016/0304-3940(95)11662-g).

16 [36] O. Valenti, A. Zambon, S. Boehm, Orchestration of Dopamine Neuron Population  
17 Activity in the Ventral Tegmental Area by Caffeine: Comparison With Amphetamine.  
18 International Journal of Neuropsychopharmacology. 24 (2021) 832-841. [https://doi:](https://doi:10.1093/ijnp/pyab049)  
19 10.1093/ijnp/pyab049.

20 [37] E. Valjent, C. Pagès, D. Hervé, J.A. Girault, J. Caboche, Addictive and non-  
21 addictive drugs induce distinct and specific patterns of ERK activation in mouse brain,  
22 European Journal of Neuroscience. 19 (2004) 1826-1836. [https://doi: 10.1111/j.1460-](https://doi:10.1111/j.1460-9568.2004.03278.x)  
23 9568.2004.03278.x.

24 [38] D. van der Kooy, R.F. Mucha, M. O'Shaughnessy, P. Buceniaks, Reinforcing effects  
25 of brain microinjections of morphine revealed by conditioned place preference, Brain  
26 Research. 243 (1982) 107-117. [https://doi: 10.1016/0006-8993\(82\)91124-6](https://doi:10.1016/0006-8993(82)91124-6).

- 1 [39] L.J. Vanderschuren, A.N. Schoffelmeer, A.H. Mulder, T.J. De Vries, Dopaminergic  
2 mechanisms mediating the long-term expression of locomotor sensitization following  
3 pre- exposure to morphine or amphetamine, *Psychopharmacology*. 143 (1999) 244-253.  
4 [https://doi: 10.1007/s002130050943](https://doi.org/10.1007/s002130050943).
- 5 [40] M.J. Wanat, A. Bonci, Dose-dependent changes in the synaptic strength on  
6 dopamine neurons and locomotor activity after cocaine exposure, *Synapse*. 62 (2008)  
7 790-795. [https://doi: 10.1002/syn.20546](https://doi.org/10.1002/syn.20546).
- 8 [41] A.R. Wang, A. Groome, L. Taniguchi, N. Eshel, B.S. Bentzley, The role of  
9 dopamine in reward-related behavior: shining new light on an old debate, *Journal of*  
10 *Neurophysiology*. 124 (2020) 309-311. [https://doi: 10.1152/jn.00323.2020](https://doi.org/10.1152/jn.00323.2020).



1 **Figure captions**

2  
3

4 **Fig. 1.** Timeline and scheme of injections.

5  
6

7 **Fig. 2.** Distance scores (M) during the induction phase of experiment 1. Data were  
8 represented as mean and S.E.M.; \*\*  $p < 0.01$  (higher locomotion) in comparison to all other  
9 groups; \*  $p < 0.05$  in comparison to APO-treated groups; +  $p < 0.05$  (lower locomotion) in  
10 comparison to all other groups; # denotes a significant difference ( $p < 0.05$ ) between the first  
11 and the last day of the induction phase for the same experimental group, according to the  
12 repeated measure two-way ANOVA followed by Tukey's post hoc test.

13  
14

15 **Fig. 3.** Distance scores (M) during the final test of experiment 1. Data were represented  
16 as mean and S.E.M.; \*\*  $p < 0.01$  (higher locomotion) in comparison with all other groups; &  
17 denotes the second higher locomotion score ( $p < 0.05$ ) except for the  
18 APO+VEH/VEH+VEH-treated group; §  $p < 0.05$  in comparison to VEH+VEH/VEH+VEH-  
19 treated group, according to a one-way ANOVA followed by Tukey's post hoc test.

20  
21

22 **Fig. 4.** ERK activation in the ventral tegmental area (VTA) from experiment 1. **A:**  
23 Quantification of immunohistochemical labeling for ERK phosphorylation in the VTA. Data  
24 represent the mean  
25  $\pm$  S.E.M.; \*\*  $p < 0.01$  in comparison to the other groups, according to one-way ANOVA  
26 followed by Tukey's post hoc test. **B-C:** Modified drawing of a brain parasagittal section  
27 displaying, at the vertical black bar, the location of the coronal midbrain section, obtained  
28 from the Paxinos and Watson atlas (2005). **D:** Representative photomicrographs of low  
29 (10X) and high (40X inserts)

1 magnification images of ERK-P-immunoreactive cells. The scale bar indicates 100  $\mu$ m in  
2 the lowmagnification images.

3  
4  
5 **Fig. 5.** ERK activation in the nucleus accumbens (NAc) from experiment 1. **A:**  
6 Quantification of immunohistochemical labelling for ERK phosphorylation in the nucleus  
7 accumbens (NAc). Data represent the mean  $\pm$  S.E.M.; \*\*  $p < 0.01$  in comparison to the other  
8 groups, according to a one-wayANOVA followed by Tukey's post hoc test. **B-C:** Modified  
9 drawing of sagittal section of the braindisplaying, at the vertical black bar, the location of  
10 the coronal section of forebrain, obtained fromthe Paxinos and Watson atlas (2005). **D:**  
11 Representative photomicrographs of low (10X) and high(40X inserts) magnification images  
12 of ERK-P-immunoreactive cells. The scale bar indicates 100  $\mu$ m in the low magnification  
13 images.

14  
15  
16 **Fig. 6.** Distance scores (M) during the induction phase from experiment 2. Data were  
17 representedas mean and S.E.M.; \*\* denotes higher locomotion ( $p < 0.01$ ) than all the other  
18 groups; # denotes asignificant difference ( $p < 0.05$ ) between the first and the last day of  
19 the induction phase for the same experimental group, according to the repeated measure  
20 two-way ANOVA followed by Tukey's post hoc test.

21  
22  
23 **Fig. 7.** Distance scores (M) during the final test of experiment 2. Data were represented  
24 as mean and S.E.M.; \*\* denotes higher locomotion ( $p < 0.01$ ) than all the other groups; +  
25 denotes lower locomotion ( $p < 0.05$ ) than all the other groups, according to a one-way  
26 ANOVA followed by Tukey's post hoc test.

1 **Fig. 8.** ERK activation in the ventral tegmental area (VTA) and in nucleus accumbens (NAc)  
2 from experiment 2. **A:** Quantification of immunohistochemical labelling for ERK  
3 phosphorylation in the VTA. **C:** Quantification of immunohistochemical labeling for ERK  
4 phosphorylation in the NAc. Data represent the mean  $\pm$  S.E.M.; \*\* denotes higher numbers  
5 ( $p < 0.01$ ) of immunoreactive nuclei than the other groups, according to a one-way ANOVA  
6 followed by Tukey's post hoc test. **B-D:** Representative photomicrographs of low (10X) and  
7 high (40X inserts) magnification images of ERK-P-immunoreactive cells. The scale bar  
8 indicates 100  $\mu$ m in the low magnification images.

9  
10  
11 **Fig. 9.** Distance scores (M) during the acute one-day experiment (experiment 3). Data  
12 represent the mean  $\pm$  S.E.M.; \*\* denotes higher locomotion than all the other groups  
13 ( $p < 0.05$ ; one-way ANOVA followed by the Tukey test).

14  
15  
16 **Fig. 10.** ERK activation in the ventral tegmental area (VTA) and in nucleus accumbens  
17 (NAc) from experiment 3. **A:** Quantification of immunohistochemical results for ERK  
18 phosphorylation in the VTA. **C:** Quantification of immunohistochemical labeling for ERK  
19 phosphorylation in the NAc. Data represent the mean  $\pm$  S.E.M.; \*\* denotes higher numbers  
20 ( $p < 0.01$ ) of immunoreactive nuclei than in the other groups, according to a one-way  
21 ANOVA followed by Tukey's post hoc test. **B-D:** Representative photomicrographs of low  
22 (10X) and high (40X inserts) magnification images of ERK-P-immunoreactive cells. The  
23 scale bar indicates 100  $\mu$ m in the low magnification images.

24

25

26

27

28

## 1 **Highlights**

- 2
- 3
- 4       • Acute MOR did not increase locomotion but repeated MOR induced
- 5       hyperactivity.
- 6       • Conditioned MOR hyperactivity was equivalent to MOR induced hyperactivity.
- 7
- 8       • APO pretreatment prevented the development and expression of MOR
- 9       conditioning.
- 10
- 11       • Acute MOR and conditioned MOR increased ERK activation in the VTA and
- 12       NAc.
- 13       • All MOR induced increases in ERK activity were eliminated by the APO
- 14       pretreatment.

15

16

**Figure 1: Timeline and scheme of injections.**

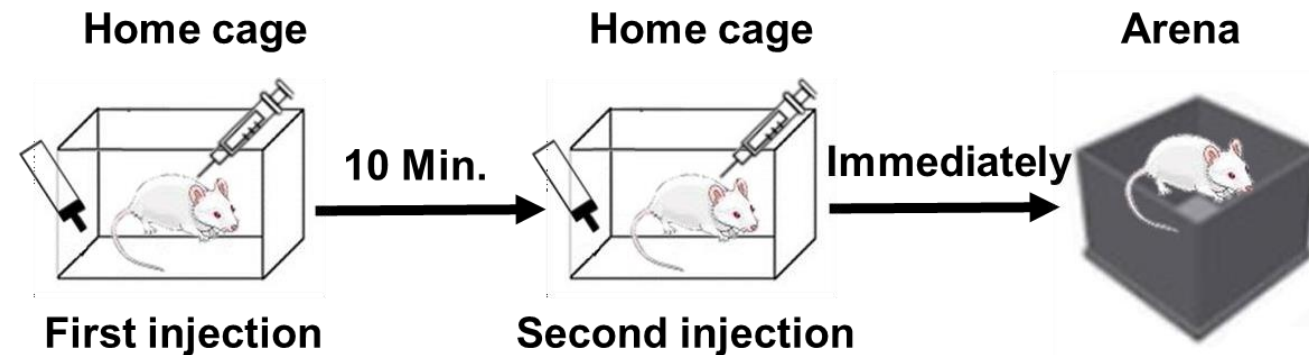
**A: Timeline – Experiment 1 and 2.**

Days	1	2	3	4	5	6	7	8	9
	Habituation			Induction Phase					Final Test

**B: Timeline – Experiment 3.**

Days	1	2	3	4
	Habituation			One Test Session

**C: Scheme of injections.**



**Table 1****Table 1: Groups of experiment 1.**

<b>Induction Phase</b>	<b>Final Test</b>	<b>Groups</b>
<b>5 Days</b>	<b>1 Day</b>	<b>Induction Phase / Final Test</b>
VEH + VEH	VEH + VEH	VEH+VEH / VEH+VEH (n=7)
VEH + VEH	VEH + <b>MOR</b>	VEH+VEH / VEH+ <b>MOR</b> (n=7)
VEH + <b>MOR</b>	VEH + VEH	VEH+ <b>MOR</b> / VEH+VEH (n=7)
VEH + <b>MOR</b>	VEH + <b>MOR</b>	VEH+ <b>MOR</b> / VEH+ <b>MOR</b> (n=7)
<b>APO</b> + VEH	VEH + VEH	<b>APO</b> +VEH / VEH+VEH (n=7)
<b>APO</b> + VEH	VEH + <b>MOR</b>	<b>APO</b> +VEH / VEH+ <b>MOR</b> (n=7)
<b>APO</b> + <b>MOR</b>	VEH + VEH	<b>APO</b> + <b>MOR</b> / VEH+VEH (n=7)
<b>APO</b> + <b>MOR</b>	VEH + <b>MOR</b>	<b>APO</b> + <b>MOR</b> / VEH+ <b>MOR</b> (n=7)

VEH=Vehicle; MOR=Morphine 10 mg/kg; APO=Apomorphine 0.05 mg/kg.

**Table 1****Table 2: Groups of experiment 2 (A) and 3 (B).****A**

<b>Induction Phase</b>	<b>Final Test</b>	<b>Groups</b>
<b>5 Days</b>	<b>1 Day</b>	<b>Induction Phase / Final Test</b>
VEH + VEH	VEH + VEH	VEH+VEH / VEH+VEH (n=7)
VEH + VEH	<b>APO</b> + VEH	VEH+VEH / <b>APO</b> +VEH (n=7)
VEH + <b>MOR</b>	VEH + VEH	VEH+ <b>MOR</b> / VEH+VEH (n=7)
VEH + <b>MOR</b>	<b>APO</b> + VEH	VEH+ <b>MOR</b> / <b>APO</b> +VEH (n=7)

**B**

<b>One Test Session</b>	<b>Groups</b>
VEH + VEH	VEH+VEH (n=6)
VEH + <b>MOR</b>	VEH+ <b>MOR</b> (n=6)
<b>APO</b> + VEH	<b>APO</b> +VEH (n=6)
<b>APO</b> + <b>MOR</b>	<b>APO</b> + <b>MOR</b> (n=6)

**VEH=Vehicle; MOR=Morphine 10 mg/kg; APO=Apomorphine 0.05 mg/kg.**

FIG. 2

EXPERIMENT 1: ACQUISITION

INDUCTION PHASE

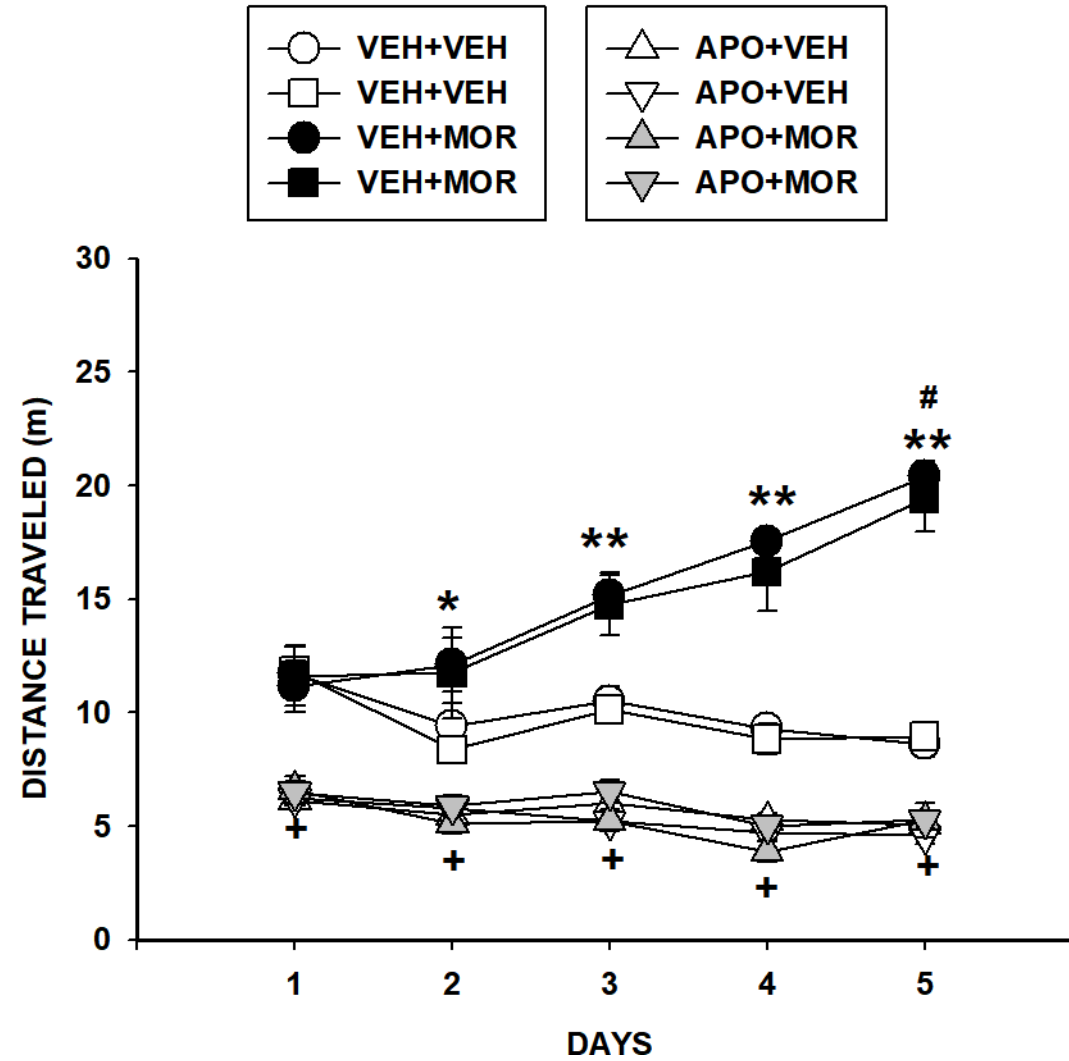




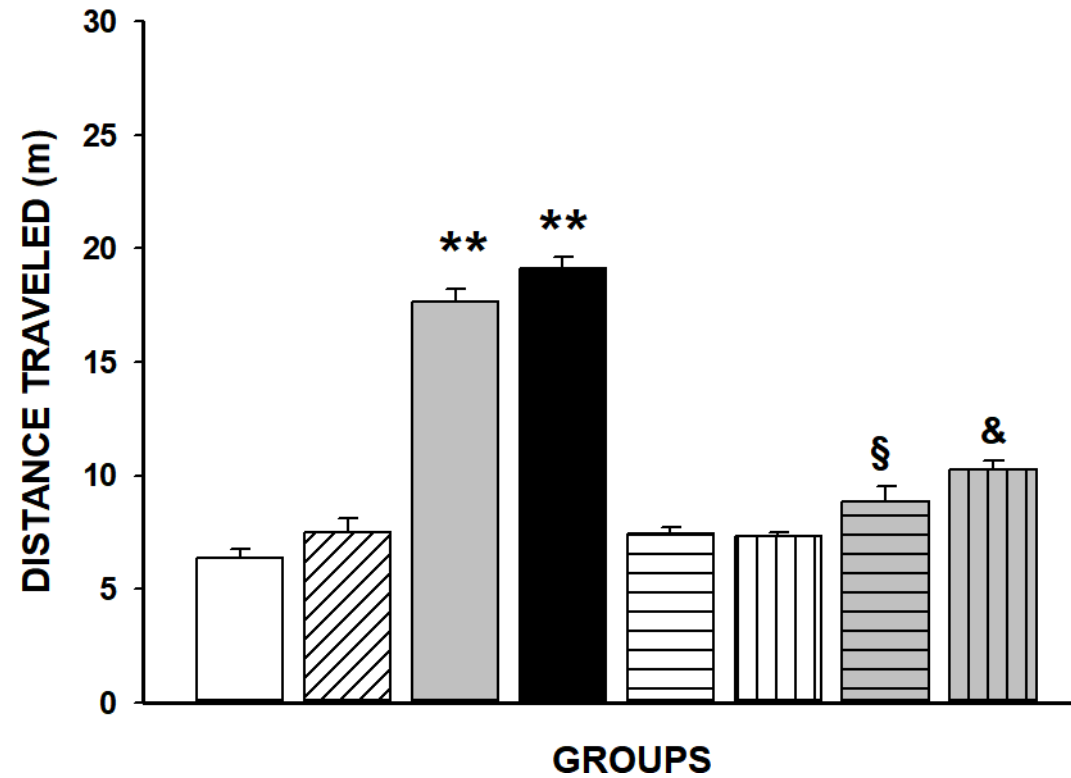
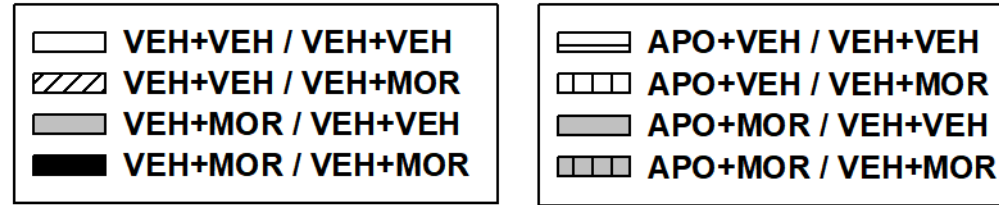
Fig. 3

# EXPERIMENT 1: ACQUISITION

## FINAL TEST

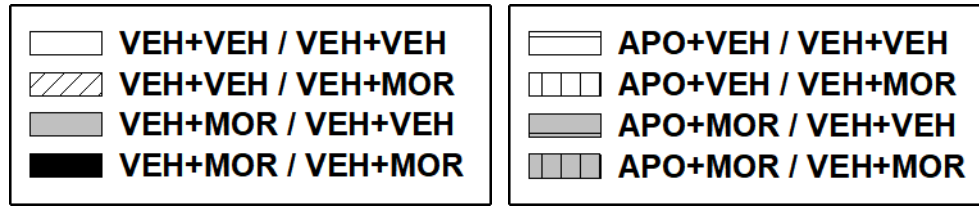
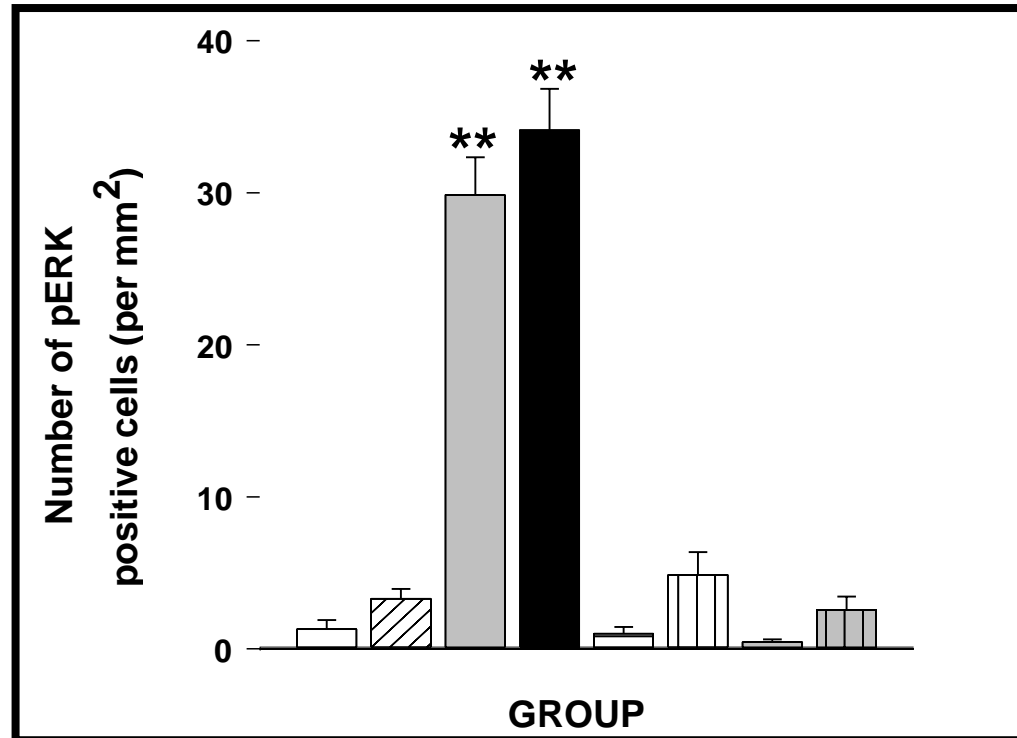
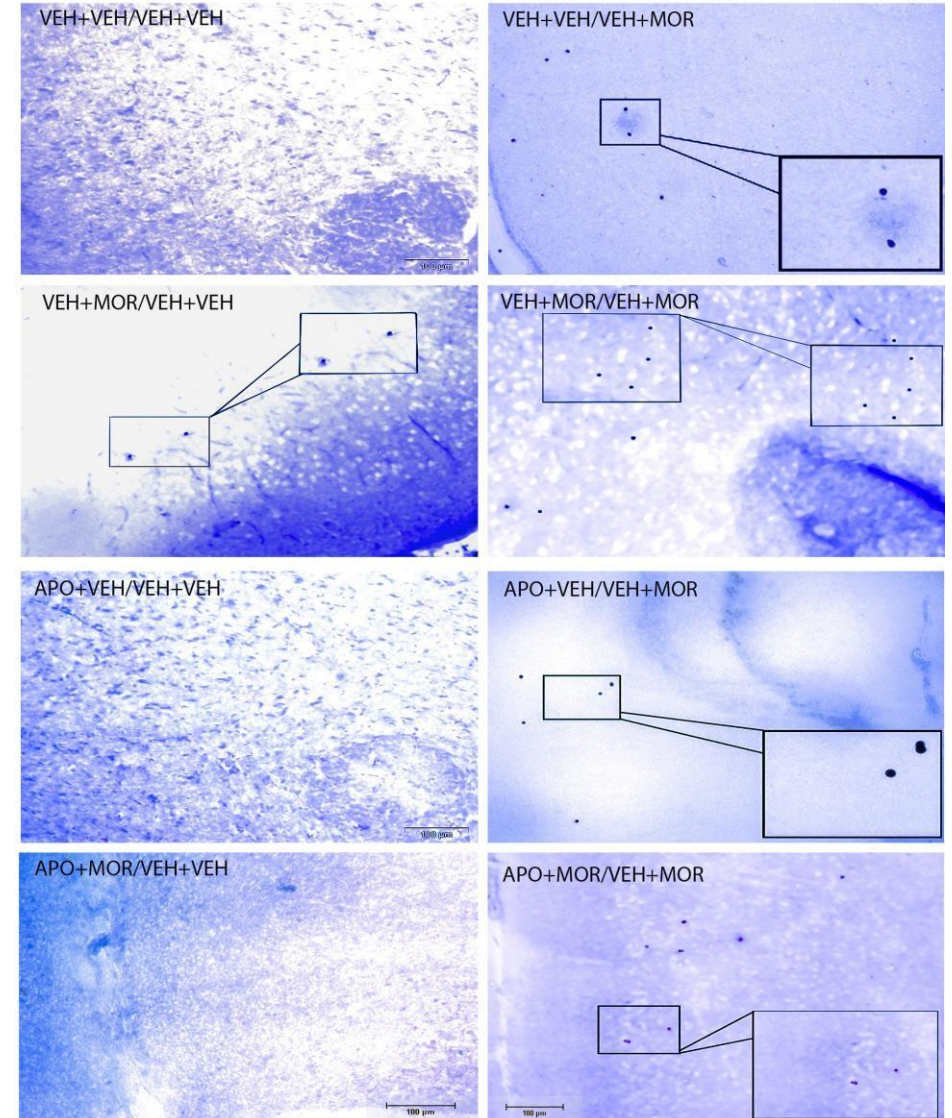
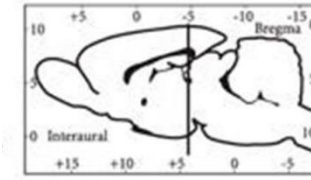
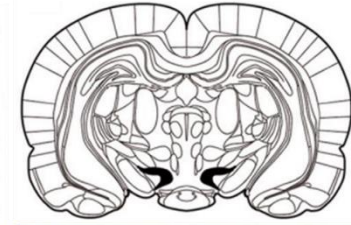
Group label:

Induction Phase Treatment / Final Test Treatment



**FIG. 4****EXPERIMENT 1: ACQUISITION****ERK ACTIVATION - VTA**

**Group label:  
Induction Phase / Final Test**

**A****D****B****C**

**FIG. 5**

**EXPERIMENT 1: ACQUISITION**

**ERK ACTIVATION - NAc**

**Group label:  
Induction Phase / Final Test**

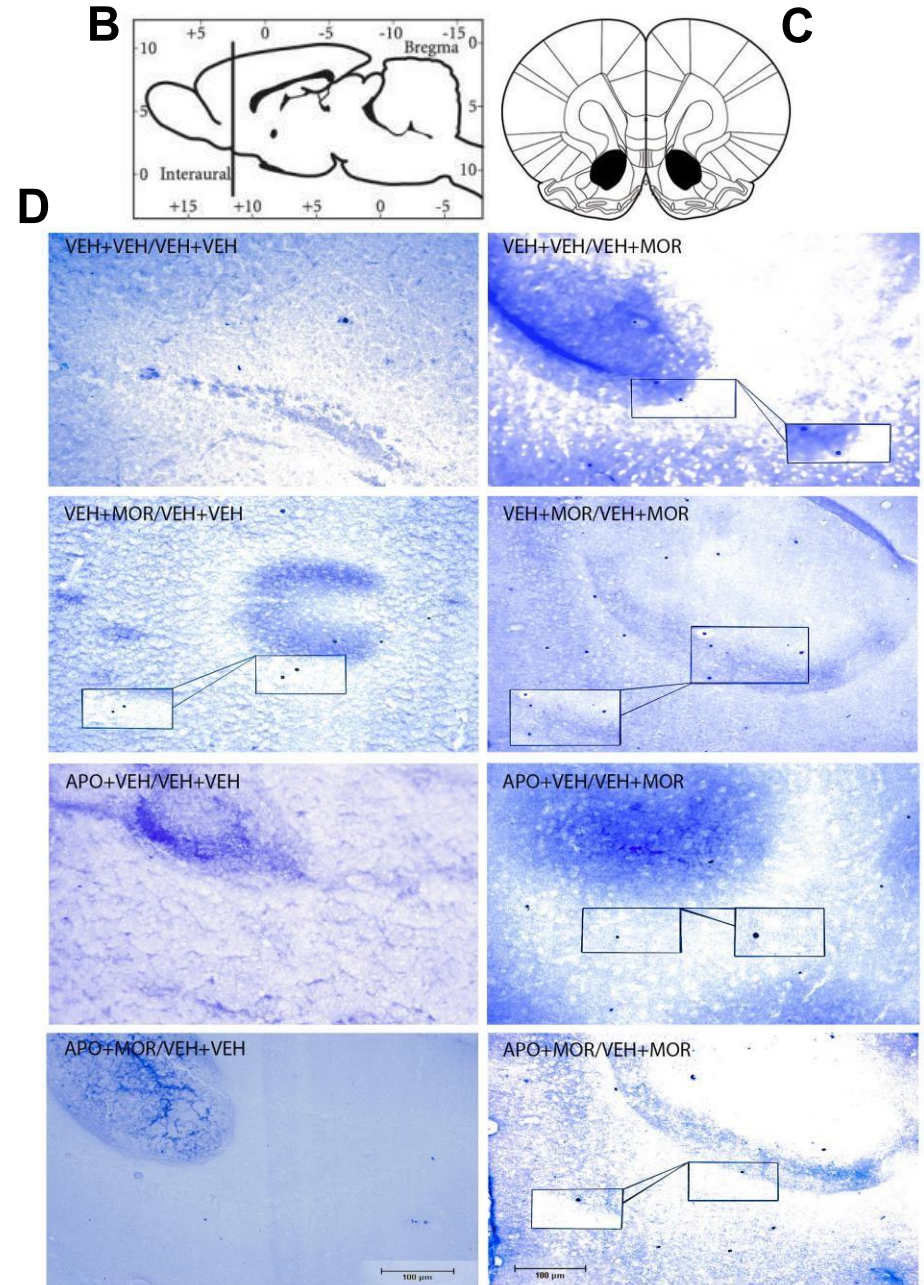
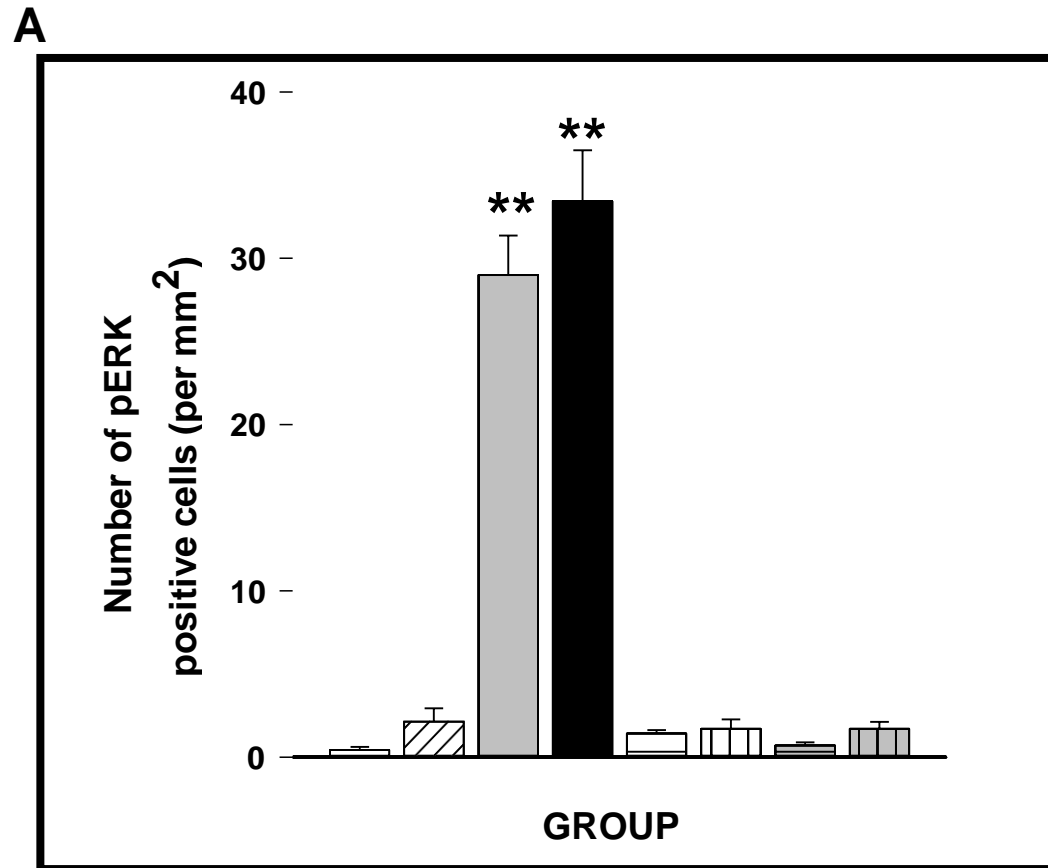
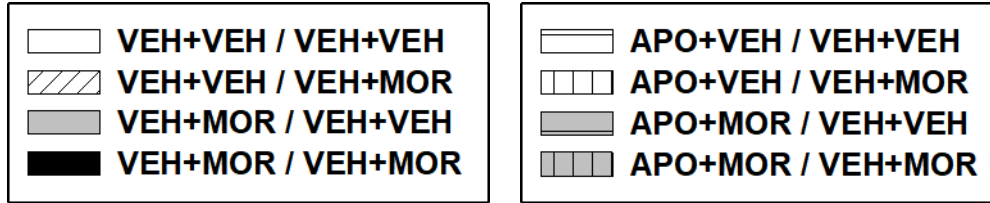


FIG. 6

EXPERIMENT 2: EXPRESSION

INDUCTION PHASE

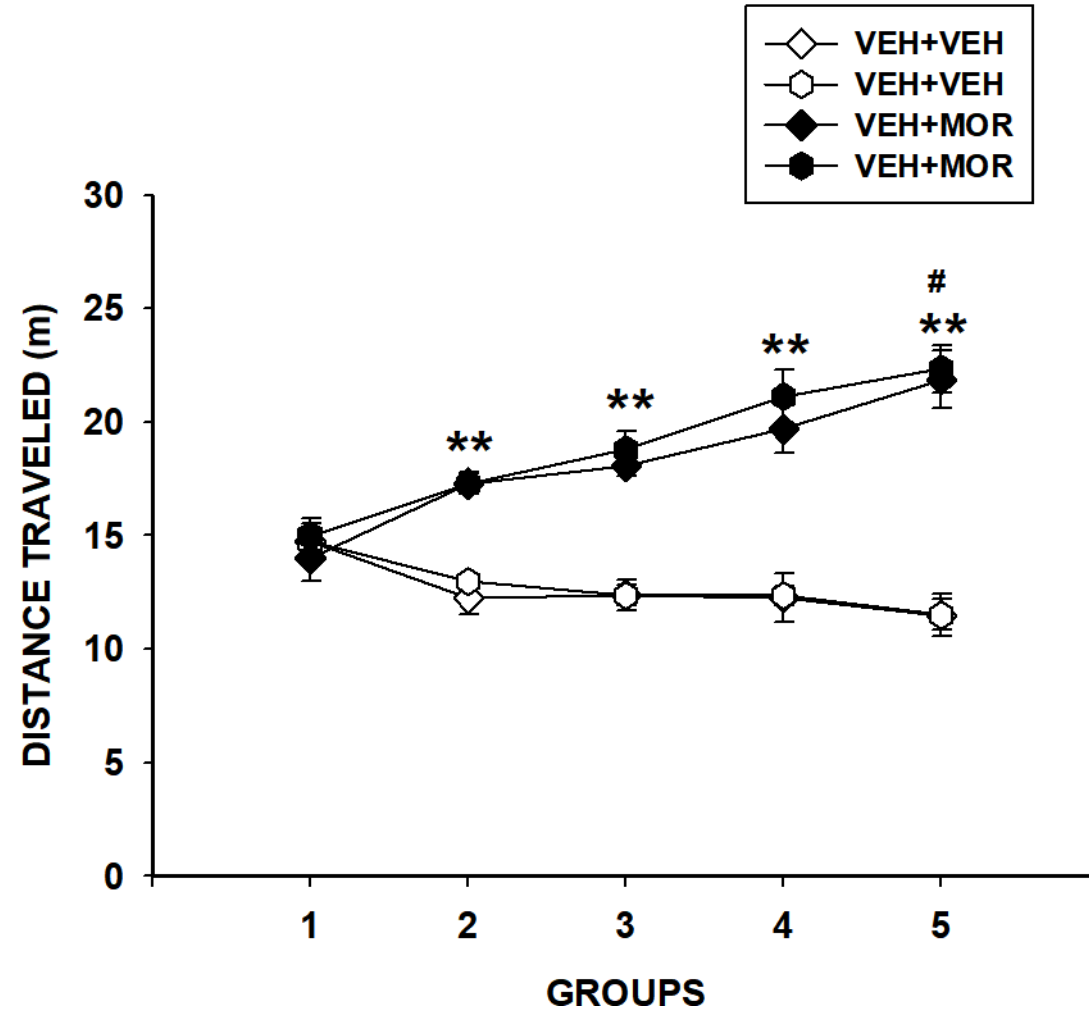


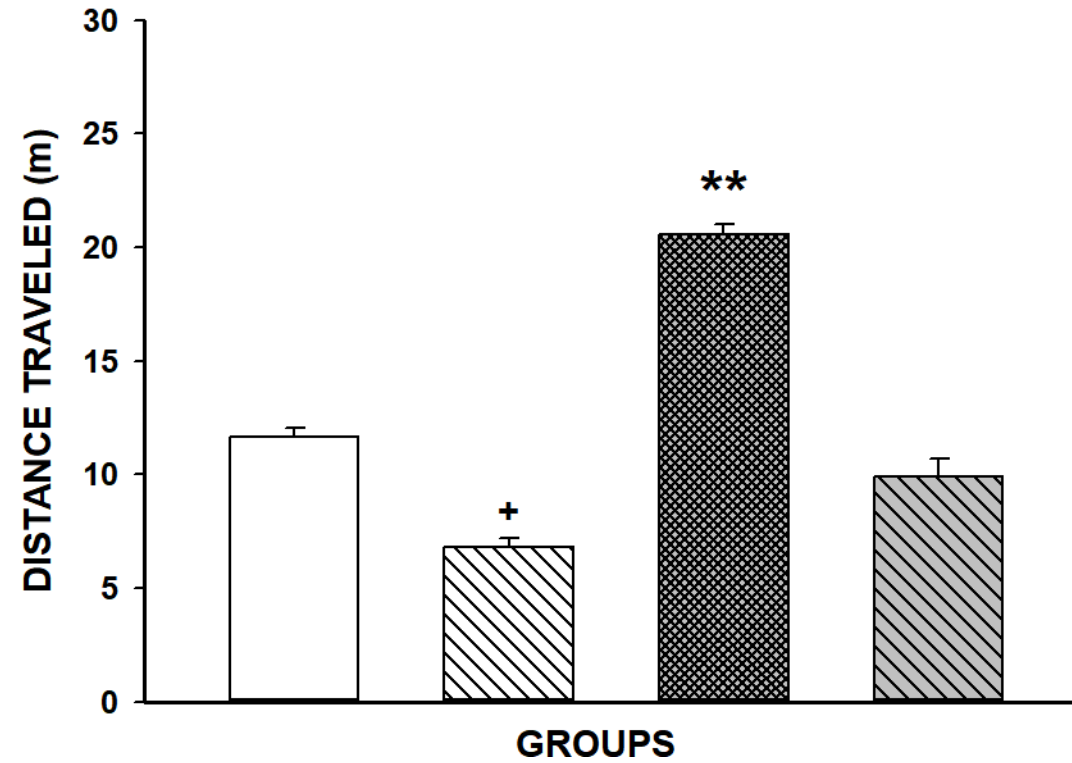
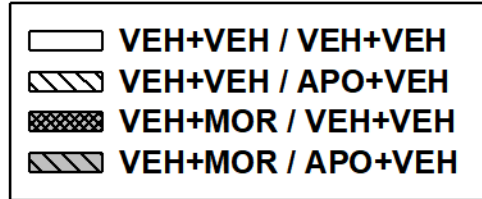
FIG.7

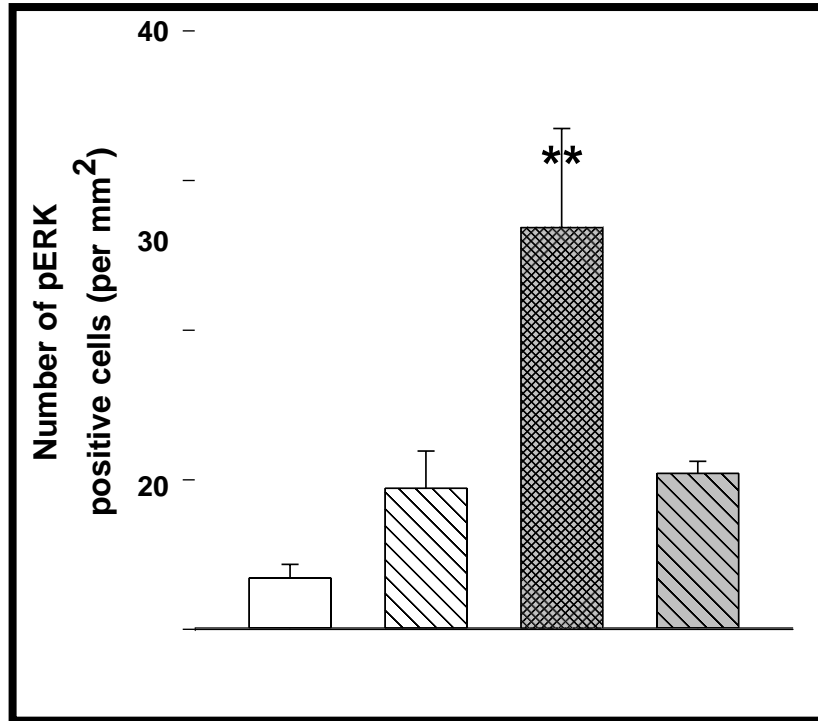
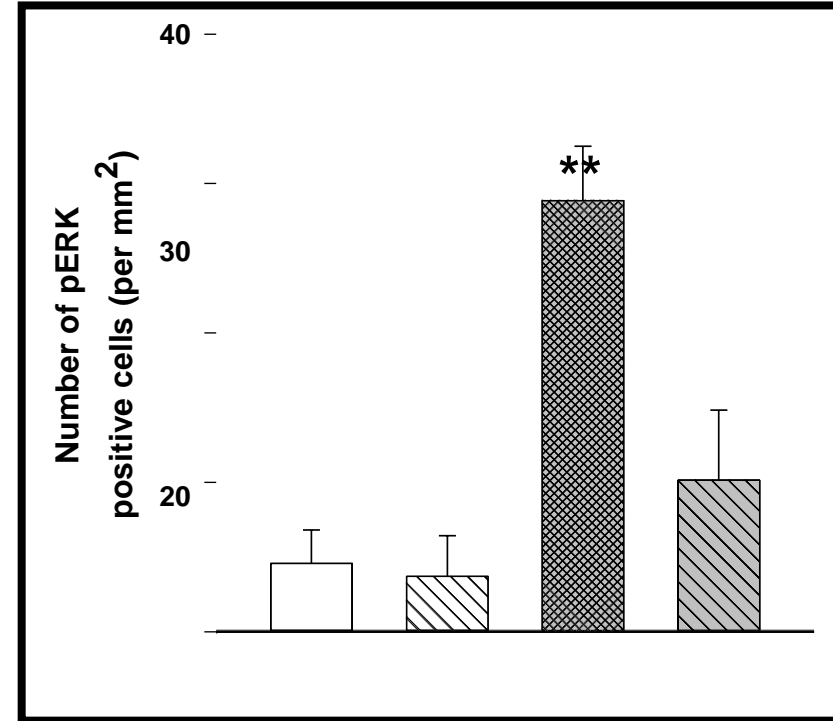
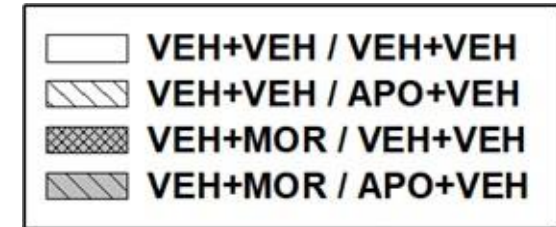
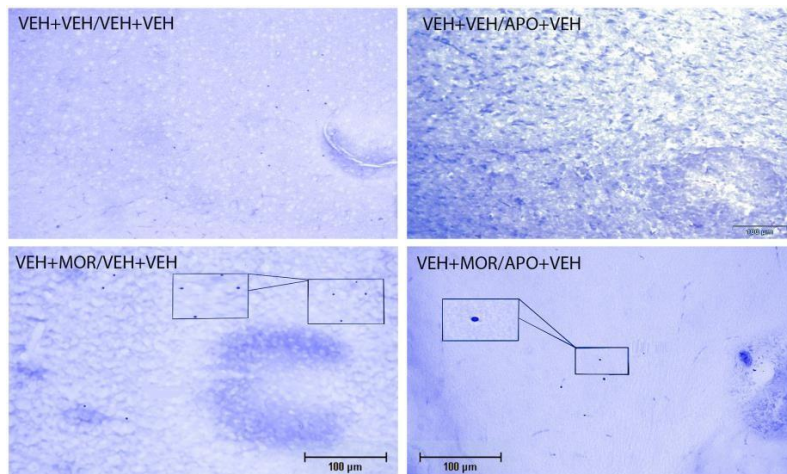
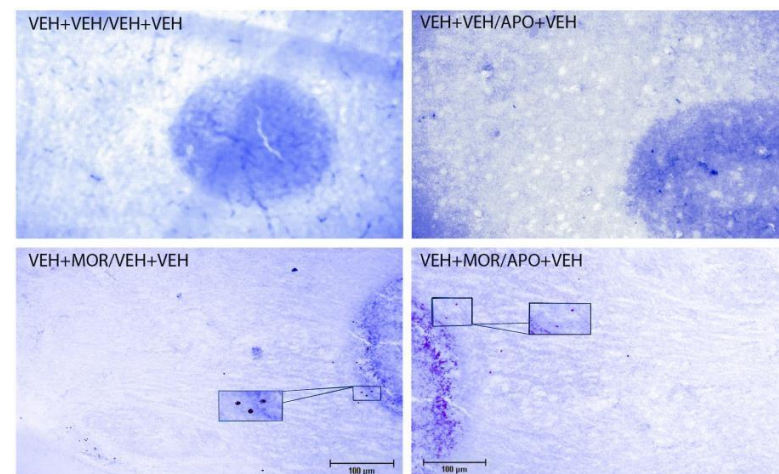
EXPERIMENT 2: EXPRESSION

FINAL TEST

Group label:

Induction Phase Treatment / Final Test Treatment



**FIG. 8****EXPERIMENT 2: EXPRESSION - ERK ACTIVATION****A****VTA****C****NAc****Group Legends:**  
Induction Phase / Final Test**B****D**

1

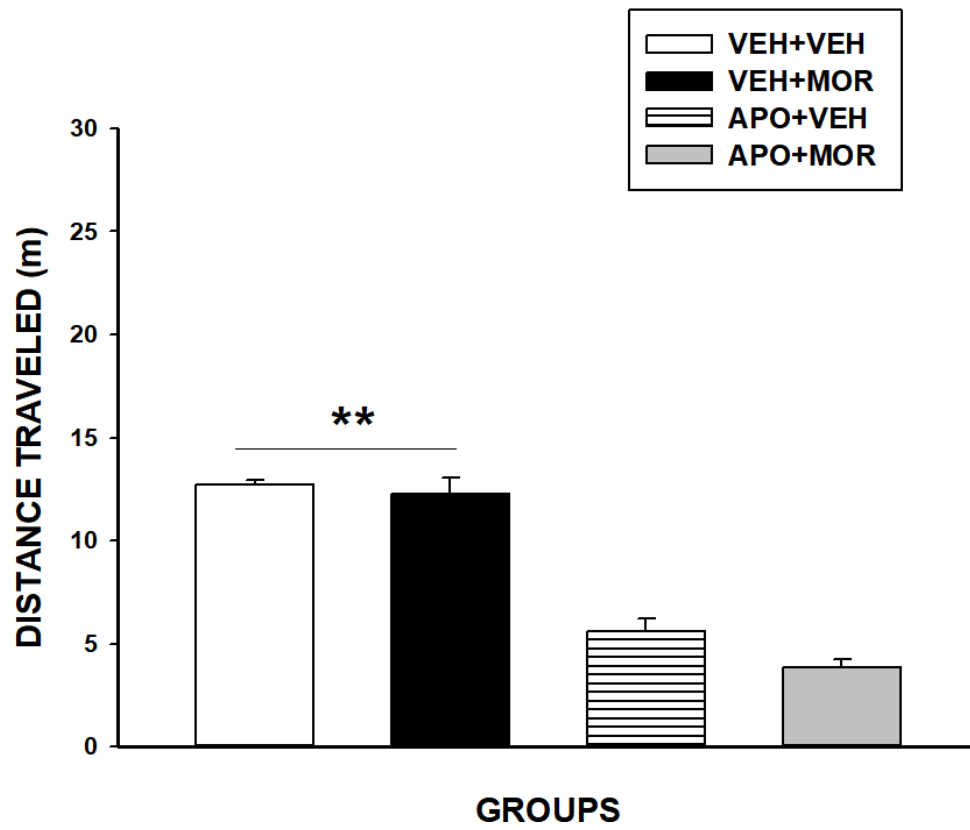
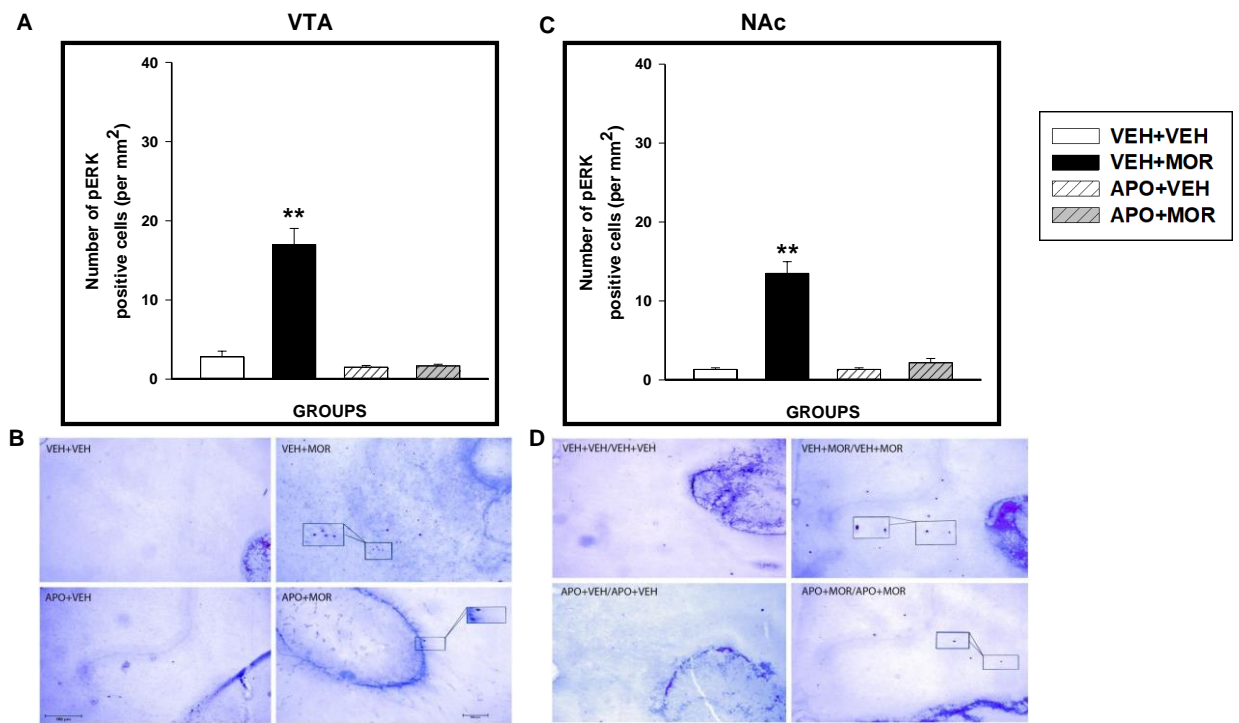
**FIG. 9****EXPERIMENT 3: ACUTE ONE DAY SESSION****LOCOMOTION**

FIG. 10

## EXPERIMENT 3: ACUTE ONE DAY SESSION - ERK ACTIVATION



1

2

3

4

5

6

7

8

9

10

11

12

13



1 **5.2 – Capítulo II**

2

3 **MK-801 induces dose dependent stimulant sensitization effects but dose**  
4 **independent conditioned stimulant effects: MK-801 effects on sensory**  
5 **information processing versus learning and memory.**

6 Joaquim Barbosa Leite Junior<sup>1</sup>, João Marcos de Mello Bastos<sup>1</sup>, Flávia Regina  
7 Cruz Dias<sup>1</sup>, Richard Ian Samuels<sup>2</sup>, Robert J Carey<sup>3</sup> and Marinete Pinheiro  
8 Carrera<sup>1</sup>

9

10 <sup>1</sup>Behavioral Pharmacology Group, Laboratory of Morphology and Pathology  
11 Animal Health, State University of North Fluminense Darcy Ribeiro, Avenida  
12 Alberto Lamego, 2000, Campos dos Goytacazes, 28013-600, RJ, Brazil.

13 <sup>2</sup>Department of Entomology and Plant Pathology, State University of North  
14 Fluminense Darcy Ribeiro, Campos dos Goytacazes, RJ, Brazil

15 <sup>3</sup>Department of Psychiatry SUNY Upstate Medical University, 800 Irving Avenue,  
16 Syracuse, NY 13210, USA

17

18 Corresponding author: Marinete Pinheiro Carrera, Behavioral Pharmacology  
19 Group, Laboratory of Morphology and Pathology Animal, State University of  
20 North Fluminense Darcy Ribeiro, Avenida Alberto Lamego, 2000, Campos dos  
21 Goytacazes, 28013-600, RJ, Brazil. Fax: +55-22-27397197; e-mail:  
22 marinete@uenf.br

23

24 Acknowledgements: This research was supported by UENF and FAPERJ (grant  
25 nº E-26/211.903/2021). J.B.L.J. and J.M.M.B. are recipients of fellowships from  
26 CAPES-Brazil. R.I.S. and M.P.C. are CNPq research fellows.

27 Conflict of interest: All authors declare no conflicts of interest.

## 1 Abstract

2 Rationale: The NMDA receptor antagonist MK-801 can prevent sensitization  
3 induced by dopaminergic stimulant drugs and also induces sensitization.

4 Objectives: To determine whether repeated MK-801 treatments can induce dose  
5 dependent locomotor sensitization and conditioned effects.

6 Methods: Initially, rats received vehicle (VEH) prior to the 30 minutes open-field  
7 sessions for 3 days (habituation) and locomotion was measured. Afterwards,  
8 groups were formed with comparable locomotion and the pharmacological phase  
9 was initiated with the treatments administered immediately prior to the five daily  
10 30 min. test sessions. Three groups received one of the three MK-801 doses  
11 (0.025, 0.1 and 1.0 mg/kg) and one group received VEH. Two days later there  
12 was a conditioning test where rats received VEH immediately prior to a single 5  
13 min. (experiment 1) test session or a 30 min. test session (experiment 2).

14 Results: A habituation response occurred in all groups. In experiment 1,  
15 sensitization effects were induced in the 1.0 mg/kg MK-801 group but not in the  
16 0.025 mg/kg group. Surprisingly, equivalent conditioned effects were observed in  
17 all MK-801 groups. Experiment 2 confirmed the responses obtained in  
18 experiment 1. Additionally, the conditioned and habituation day 3 locomotor  
19 responses were equivalent in all MK-801 groups, whereas for the VEH group, the  
20 conditioned response was lower than on habituation day 3.

21 Conclusions: We relate the conditioned response to sensory neglect effects  
22 induced by MK-801. The impact of MK-801 on the sensory processes is  
23 independent of the motor stimulant responses and can mimic effects on memory.

24 Key-words: MK-801, NMDA receptor, motor stimulation, learning and memory,  
25 sensory neglect, drug stimuli.

## 26 Introduction

27

1 The importance of the basal ganglia in movement processes has long been  
2 established. Indeed, it is the interplay between glutamate and dopamine (DA)  
3 systems in the basal ganglia that mediates normative movement wherein  
4 glutamatergic inhibitory activity is balanced by dopaminergic excitatory activity  
5 (Dai and Carey 1995). Thus, excessive movement can occur either by decreasing  
6 glutamate activity or increasing dopamine activity. Numerous studies have  
7 shown that in rats direct as well as indirect acting dopamine agonists can induce  
8 locomotor hyperactivity and that these effects undergo conditioning and  
9 sensitization (Anagnostaras and Robinson 1996; Bloise *et al.* 2007; Braga *et al.*  
10 2009; de Matos *et al.* 2010; Keller and Delius 2001; Mattingly *et al.* 1997, 1988;  
11 Rowlett *et al.* 1997). Similarly, antagonism at the glutamate N-methyl-D-aspartate  
12 (NMDA) receptor by drugs such as dizocilpine (MK-801) can induce hyperactivity  
13 in rats and that with repeated treatments this hyperactivity becomes exaggerated,  
14 indicative of sensitization (Carey *et al.* 1995).

15 Interestingly, the sensitization and conditioned effects induced by dopamine  
16 agonists can be prevented if the glutamate NMDA receptor non-competitive  
17 antagonist MK-801 is given in combination with DA agonists such as  
18 amphetamine and cocaine (Stewart and Druhan 1993; Vezina and Queen 2000;  
19 Zweifel *et al.* 2008), including the hyper motoric stimulation induced by L-DOPA  
20 in an animal model of Parkinson's (Pinheiro-Carrera *et al.* 1995). In that  
21 behavioral drug sensitization and conditioning are persistent and lasting effects,  
22 and that NMDA receptors are important for neuronal changes such as long-term  
23 potentiation (LTP), drug induced activation of the NMDA receptor site has  
24 provided a possible explanation for drug sensitization and conditioning. While a  
25 substantial body of evidence supports the efficacy of NMDA antagonists in  
26 preventing the development of sensitization to other drugs, it is also the case that  
27 repeated MK-801 treatments can induce MK-801 behavioral sensitization effects  
28 (Carey *et al.* 1995). Thus, while MK-801 can prevent the sensitization induced by  
29 dopaminergic stimulant drugs it also induces behavioral stimulant effects that  
30 also undergo sensitization with repeated treatments. Thus, MK -801 has the  
31 seemingly puzzling capability to both prevent and induce stimulant sensitization

1 effects. In that MK-801 can prevent the Pavlovian conditioning of dopaminergic  
2 drug induced hyperactivity (Damianopoulos and Carey 1995), the question arises  
3 as to whether MK-801 induced hyperactivity can undergo Pavlovian conditioning  
4 as occurs with dopamine agonist induced hyperactivity. Specifically, the present  
5 study was undertaken to determine whether repeated MK-801 treatments that  
6 can induce dose dependent behavioral stimulant sensitization effects and also  
7 induce conditioned stimulant effects. In this study, a replication experimental  
8 design was used in which the experiment was conducted twice in two separate  
9 independent experiments to enhance the reliability of the findings.

10

## 11 Methods

12

### 13 Subjects

14 Male Wistar albino rats provided by the State University of North  
15 Fluminense Darcy Ribeiro, initially weighing 200-300 g, were housed in individual  
16 plastic cages (25 X 18 X 17 cm) until the end of the experiment. Food and water  
17 were always available. The vivarium was maintained at a constant temperature  
18 ( $22 \pm 2$  °C) and a 12h/12h light/dark cycle (lights on at 07:00h and off at 19:00h).  
19 All experiments occurred between 8:00 and 12:00h. For 7 days prior to all  
20 experimental procedures, each animal was weighed and handled daily for 5 min.  
21 This process included being placed in a transport cage and taken to the injection  
22 administration bench located in an anteroom adjacent to the experimental testing  
23 room. The animals were given vehicle (VEH) injections. In this way the animals  
24 were familiarized with the handling and injection aspects of the testing protocol.  
25 The experimental procedures were in accordance with the recommendations of  
26 the Commission of Ethics in Animal Experimentation (CEUA) of the University of  
27 North Fluminense Darcy Ribeiro-UENF (process 395/2018). These  
28 recommendations are in agreement with the ethical principles of animal research  
29 adopted by the Brazilian College of Animal Experimentation (COBEA) and by the  
30 National Council for Animal Experimentation Control (CONCEA).

## 1 Drugs

2

3 Dizocilpine [(+)-MK-801; (+)-5-methyl-10,11-dihydro-5H-dibenzo[a,d]cyclohepten-  
4 5,10-imine maleate]—further referred to as MK-801—was purchased from  
5 Sigma–Aldrich (Sigma, St Louis, USA) was dissolved in saline (0.9% NaCl  
6 solution) at doses of 0.025 mg/kg, 0.1 mg/kg and 1.0 mg/kg (Carey *et al.* 1998;  
7 Frantz and Van Hartesveldt 1999; Qi *et al.* 2008). Saline alone was administered  
8 to vehicle controls. MK-801 or vehicle was subcutaneously (s.c.) injected in a  
9 volume of 1.0 ml/kg body weight.

10

## 11 Apparatus and behavioral measurements

12

13 The behavioral measurements were conducted in a black open field chamber (60  
14 X 60 X 45 cm). A closed-circuit camera (IKEGAMI, model ICD-49) mounted 60  
15 cm above the arena was used to record behavioral data. Locomotion, measured  
16 as distance traveled (m), was automatically analyzed using software EthoVision  
17 (Noldus, the Netherlands). The complete test procedure was conducted  
18 automatically without the presence of the experimenter in the test room. All  
19 behavioral testing was conducted under dim red light that favors exploratory  
20 behavior and avoids the possible aversive quality of white light (Nasello *et al.*  
21 1998) as well as to enhance the contrast between the white subject and the dark  
22 background of the test chamber and reduce the animal's shadow.

23

## 24 Experimental procedure

25

26 Two similar experiments were conducted. In experiment 1 rats were initially given  
27 three 30 min tests in the open field arena on 3 successive days (habituation  
28 period). Immediately prior to each of the habituation test days, the subjects

1 received vehicle (VEH) injections. The locomotor distance scores were used to  
2 form four groups (n=5 for each group) with equivalent distance scores.  
3 Subsequently the groups were injected with VEH, 0.025, 0.1 or 1.0 mg/kg MK-  
4 801 immediately before 30 min test sessions in the open field arena on 5  
5 successive days (pharmacological treatment phase). Following 2 days of non-  
6 testing (withdrawal period), all groups were given VEH injections and tested for 5  
7 min to assess possible conditioned drug effects (conditioning test; CT).

8 Experiment 2 was undertaken because of the finding in experiment 1 that all MK-  
9 groups expressed similar conditioned stimulant responses in the conditioning test  
10 despite having very different locomotor stimulant responses in the five days of  
11 drug treatment phase. In that the number of animals per group in experiment 1  
12 was modest and to have a more substantial conditioning test the protocol used in  
13 experiment 1 was repeated and the conditioning test session was expanded to  
14 30 minutes (n=6-7 per group). Table 1 present, respectively, the experimental  
15 timeline (A) and the treatment protocols (B).

16

## 17 Statistics

18

19 For the drug treatment phase, a repeated two-way analysis of variance (ANOVA)  
20 was used to determine the group effect, day effect, as well as the interactions  
21 between both factors. When a significant effect of interaction was recorded, data  
22 were further statistically evaluated using a one-way ANOVA followed by Tukey's  
23 test with  $p < 0.05$  as the criterion for statistical significance. For the 5 and 30 min  
24 conditioning test a one-way ANOVA followed by Tukey's post-hoc test were used  
25 with  $p < 0.05$  as the criterion for statistical significance. For the within-treatment  
26 assessment of the behavioral activity during conditioning test of the experiment  
27 2, the total activity score was divided into 6 intervals of 5 min each and a repeated  
28 measure two-way ANOVA was used to determine treatments, intervals, and  
29 interactions between factors. When a significant effect of treatment group and

1 intervals was obtained, data were further evaluated by one-way ANOVA followed  
2 by Tukey's post-hoc tests using  $p < 0.05$  as the criterion for statistical significance.

3

## 4 Results

5

### 6 Pre-treatment habituation effects

7

8 Prior to the initiation of each experiment, a three-day habituation procedure was  
9 conducted and locomotion was measured. In experiment 1, a one-way ANOVA  
10 showed there was an effect of days of treatment [ $F(2, 63) = 24.0$ ;  $p < 0.01$ ] and  
11 the Tukey's test showed that day 1 had higher locomotion than the day 2 and day  
12 3 ( $p < 0.05$ ) (data not shown). In experiment 2, a one-way ANOVA showed that  
13 there was an effect of days of treatment [ $F(2, 69) = 18.0$ ;  $p < 0.01$ ] and the Tukey's  
14 test showed that day 1 had higher locomotion than day 2 and day 3 ( $p < 0.05$ )  
15 (data not shown).

16

### 17 Experiment 1: 5 minute conditioning test

18

19 Figure 1 shows the locomotor activity over the course of days 1-5 of the induction  
20 phase of the first experiment. For the total locomotor activity over the course of  
21 days 1-5 (induction phase), a repeated two-way ANOVA showed a group X days  
22 interaction [ $F(12, 72) = 7.64$ ;  $p < 0.01$ ], an effect of groups [ $F(3, 18) = 22.25$ ;  
23  $p < 0.01$ ] and an effect of days of treatment [ $F(4, 72) = 7.33$ ;  $p < 0.01$ ]. A one-way  
24 ANOVA followed by Tukey test to further analyze the interaction of group X days  
25 showed that on day 1, the MK-1.0 group had higher locomotion than the VEH  
26 group ( $p < 0.05$ ). There was no difference among the MK-1.0, MK-0.1 and MK-  
27 0.025 groups ( $p > 0.05$ ). On days 2-5, the MK-1.0 group had higher locomotion  
28 than all other groups ( $p < 0.05$ ). However, on day 4 and day 5, the MK-0.1 had

1 higher locomotion than the VEH group ( $p < 0.05$ ). There was no difference  
2 between the MK-0.1 and MK-0.025 groups ( $p > 0.05$ ).

3 Figure 2 shows the locomotor activity during the conditioning test for the first  
4 experiment. A one-way ANOVA showed that there were differences among the  
5 groups [ $F(3, 18) = 8.0$ ;  $p < 0.01$ ] and the Tukey test showed that the MK groups,  
6 (MK-0.025, MK-0.1 and MK-1.0 groups), had higher locomotion than all VEH  
7 groups ( $p < 0.05$ ) and were not significantly different from each other ( $p > 0.05$ ).

8

9 Experiment 2: 30 minute conditioning test

10

11 Figure 3 shows the locomotor activity over the course of days 1-5 of the induction  
12 phase of the second experiment. A repeated two-way ANOVA showed an  
13 interaction group  $\times$  day [ $F(12, 80) = 7.54$ ;  $p < 0.01$ ], an effect of groups [ $F(3, 20)$   
14  $= 50.80$ ;  $p < 0.01$ ] and an effect of days of treatment [ $F(4, 80) = 11.25$ ;  $p < 0.01$ ]. A  
15 one-way ANOVA followed by the Tukey test to further analyze the interaction of  
16 group  $\times$  days, showed on day 1, that the MK-1.0 group had higher locomotion  
17 than the VEH group ( $p < 0.05$ ). There was no significant difference among the  
18 VEH, MK-0.025 and MK-0.1 groups ( $p > 0.05$ ). Also, there was no significant  
19 difference among the MK-0.025, MK-0.1 and MK-1.0 groups ( $p > 0.05$ ). From day  
20 2 until day 5, the MK-1.0 groups had higher locomotion than all groups ( $p < 0.05$ ).  
21 However, from day 2 until day 4, the MK-0.1 group had higher locomotion than  
22 the VEH group ( $p < 0.05$ ). There was no difference between the MK-0.1 and MK-  
23 0.25 groups ( $p > 0.05$ ) and no difference between the VEH and MK-0.025 groups  
24 ( $p > 0.05$ ). On day 5, the MK-0.1 group had higher locomotion than the VEH and  
25 MK-0.025 groups ( $p < 0.05$ ). There was no difference between the VEH and MK-  
26 0.025 groups ( $p > 0.05$ ).

27 Figure 4 shows the within session locomotor activity during the 5 induction days  
28 of the second experiment. For induction day 1 (Fig. 4A), a repeated two-way  
29 ANOVA showed an interaction group  $\times$  interval [ $F(15, 100) = 7.32$ ;  $p < 0.01$ ], an  
30 effect of groups [ $F(3, 20) = 3.35$ ;  $p < 0.05$ ] and an effect of intervals [ $F(5, 100) =$



1 31.53;  $p < 0.01$ ]. A one-way ANOVA followed by the Tukey test to further analyze  
2 the interaction showed that on interval 1 [ $F(3, 20) = 14.63$ ;  $p < 0.01$ ], showed that  
3 there was difference among the groups and Tukey's test showed that the MK-1.0  
4 group had higher locomotion than all other groups ( $p < 0.05$ ). On interval 2 [ $F(3,$   
5  $20) = 5.0$ ;  $p < 0.01$ ], there was difference among the groups and Tukey's test  
6 showed that the MK-1.0 group had higher locomotion than the VEH group  
7 ( $p < 0.05$ ). There was no difference among the MK-1.0, MK-0.1 and MK-0.25  
8 groups ( $p > 0.05$ ) and no differences among the VEH, MK-0.1 and MK-0.25 groups  
9 ( $p > 0.05$ ). On interval 3 [ $F(3, 20) = 0.25$ ;  $p > 0.05$ ], interval 4 [ $F(3, 20) = 1.80$ ;  
10  $p > 0.05$ ], interval 5 [ $F(3, 20) = 1.90$ ;  $p > 0.05$ ] and interval 6 [ $F(3, 20) = 1.45$ ;  
11  $p < 0.05$ ], there were no differences among the groups.

12 For induction day 2 (Fig. 4B), a repeated two-way ANOVA showed an interaction  
13 group X interval [ $F(15, 100) = 5.91$ ;  $p < 0.01$ ], an effect of groups [ $F(3, 20) =$   
14  $34.13$ ;  $p < 0.01$ ] and an effect of intervals [ $F(5, 100) = 22.02$ ;  $p < 0.01$ ]. A one-way  
15 ANOVA followed by the Tukey test to further analyze the interaction, showed that  
16 on interval 1 [ $F(3, 20) = 2.22$ ;  $p > 0.05$ ], there was no difference among the groups.  
17 On interval 2 [ $F(3, 20) = 23.53$ ;  $p < 0.01$ ], there was a difference among the groups  
18 and Tukey's test showed that the MK-1.0 groups had higher locomotion than all  
19 other groups ( $p < 0.05$ ). On interval 3 [ $F(3, 20) = 16.0$ ;  $p < 0.01$ ], there was a  
20 difference among the groups and Tukey's test showed that the MK-1.0 group had  
21 higher locomotion than all other groups ( $p < 0.05$ ). The results also showed that  
22 the MK-0.1 group had higher locomotion than the VEH, and MK-0.25 groups  
23 ( $p < 0.05$ ). There was no difference between the MK-0.1 and MK-0.25 groups  
24 ( $p > 0.05$ ) and no difference between VEH and MK-0.25 groups ( $p > 0.05$ ). On  
25 interval 4 [ $F(3, 20) = 1.50$ ;  $p > 0.05$ ], there was no difference among the groups.  
26 On interval 5 [ $F(3, 20) = 8.54$ ;  $p < 0.01$ ], there was difference among the groups  
27 and Tukey's test showed that the MK-1.0 groups had higher locomotion than all  
28 the other groups ( $p < 0.05$ ), except the MK-0.1 group in which there was no  
29 difference between these groups ( $p > 0.05$ ). However, the MK-0.1 group had  
30 higher locomotion than the VEH group ( $p < 0.05$ ), but there was no difference  
31 between MK-0.1 and MK-0.025 groups ( $p > 0.05$ ) and no difference between the

1 MK-0.025 and VEH groups ( $p>0.05$ ). On interval 6 [ $F(3, 20) = 0.70$ ;  $p>0.05$ ],  
2 there was no significant difference among the groups.

3 For induction day 3 (Fig. 4C), a repeated two-way ANOVA showed an interaction  
4 group X interval [ $F(15, 100) = 5.10$ ;  $p<0.01$ ], an effect of groups [ $F(3, 20) = 28.0$ ;  
5  $p<0.01$ ] and an effect of intervals [ $F(5, 100) = 18.30$ ;  $p<0.01$ ]. A one-way ANOVA  
6 followed by the Tukey test to further analyze the interaction, showed that on  
7 interval 1 [ $F(3, 20) = 20.70$ ;  $p<0.01$ ], there was a difference among the groups  
8 and Tukey's test showed that the MK-1.0 group had higher locomotion than all  
9 other groups ( $p<0.05$ ). The results also showed that the MK-0.1 and MK-0.025  
10 groups had higher locomotion than the VEH group ( $p<0.05$ ), there was no  
11 difference between the MK-0.1 and MK-0.025 groups ( $p>0.05$ ). On interval 2 [ $F$   
12  $(3, 20) = 23.00$ ;  $p<0.01$ ] and interval 3 [ $F(3, 20) = 15.50$ ;  $p<0.01$ ], the MK-1.0  
13 group had higher locomotion than all other groups ( $p<0.05$ ). On interval 4 [ $F(3,$   
14  $20) = 5.0$ ;  $p<0.01$ ] and interval 5 [ $F(3, 20) = 5.50$ ;  $p<0.01$ ], there was a difference  
15 among the groups and Tukey's test showed that the MK-1.0 group had higher  
16 locomotion than all the other groups ( $p<0.05$ ), except the MK-0.1 group. There  
17 was no difference between the MK-1.0 and MK-0.1 groups ( $p>0.05$ ) and no  
18 difference among VEH, MK-0.1 and MK-0.25 groups ( $p>0.05$ ). On interval 6 [ $F$   
19  $(3, 20) = 4.40$ ;  $p<0.05$ ], the MK-1.0 and MK-0.1 groups had higher locomotion  
20 than the VEH group ( $p<0.05$ ). There was no difference between the MK-1.0, MK-  
21 0.1 and MK-0.025 groups ( $p>0.05$ ) and no differences between the VEH and MK-  
22 0.025 groups ( $p>0.05$ ).

23 For induction day 4 (Fig. 4D), a repeated two-way ANOVA showed an interaction  
24 group X interval [ $F(15, 100) = 5.23$ ;  $p<0.01$ ], an effect of groups [ $F(3, 20) = 27.0$ ;  
25  $p<0.01$ ] and an effect of intervals [ $F(5, 100) = 14.21$ ;  $p<0.01$ ]. A one-way ANOVA  
26 followed by the Tukey test to further analyze the interaction, showed on interval  
27 1 [ $F(3, 20) = 14.21$ ;  $p<0.01$ ], that there was a difference among the groups and  
28 Tukey's test showed that the MK-1.0 group had higher locomotion than all the  
29 other groups ( $p<0.05$ ). On interval 2 [ $F(3, 20) = 45.10$ ;  $p<0.01$ ], there was a  
30 difference among the groups and Tukey's test showed that the MK-1.0 group had  
31 higher locomotion than all the other groups ( $p<0.05$ ). The results also showed

1 that the MK-0.1 and MK-0.025 groups had higher locomotion than the VEH group  
2 ( $p < 0.05$ ), and there were no differences between the MK-0.1 and MK-0.025 group  
3 ( $p > 0.05$ ). On interval 3 [ $F(3, 20) = 12.20$ ;  $p < 0.01$ ] and interval 4 [ $F(3, 20) = 8.53$ ;  
4  $p < 0.01$ ], the MK-1.0 group had higher locomotion than all the other groups  
5 ( $p < 0.05$ ). On interval 5 [ $F(3, 20) = 6.0$ ;  $p < 0.01$ ] and interval 6 [ $F(3, 20) = 5.10$ ;  
6  $p < 0.01$ ], there was a difference among the groups and Tukey's test showed that  
7 the MK-1.0 group had higher locomotion than the VEH and MK-0.025 groups  
8 ( $p < 0.05$ ). There was no difference between the MK-1.0 and MK-0.1 groups  
9 ( $p > 0.05$ ) and no difference among VEH, MK-0.1 and MK-0.25 groups ( $p > 0.05$ ).

10 For induction day 5 (Fig. 4E), a repeated two-way ANOVA showed an interaction  
11 group X interval [ $F(15, 100) = 3.32$ ;  $p < 0.01$ ], an effect of groups [ $F(3, 20) =$   
12  $27.10$ ;  $p < 0.01$ ] and an effect of intervals [ $F(5, 100) = 13.20$ ;  $p < 0.01$ ]. A one-way  
13 ANOVA followed by the Tukey test showed that on interval 1 [ $F(3, 20) = 8.70$ ;  
14  $p < 0.01$ ], there was difference among the groups and Tukey's test showed that  
15 the MK-1.0, MK-0.1 and MK-0.025 groups had higher locomotion than the VEH  
16 group ( $p < 0.05$ ). There was no difference among the MK-1.0, MK-0.1 and MK-  
17 0.025 groups ( $p > 0.05$ ). On interval 2 [ $F(3, 20) = 15.73$ ;  $p < 0.01$ ], there was a  
18 difference among the groups and Tukey's test showed that the MK-1.0 group had  
19 higher locomotion than all the other groups ( $p < 0.05$ ). The results also showed  
20 that MK-0.1 had higher locomotion than the VEH group ( $p < 0.05$ ). There was no  
21 difference between MK-0.1 and MK-0.025 groups ( $p > 0.05$ ) and no difference  
22 between MK-0.025 and VEH groups ( $p > 0.05$ ). On interval 3 [ $F(3, 20) = 12.60$ ;  
23  $p < 0.01$ ], the MK-1.0 groups had higher locomotion than all the other groups  
24 ( $p < 0.05$ ). On interval 4 [ $F(3, 20) = 12.30$ ;  $p < 0.01$ ], the MK-1.0 group had higher  
25 locomotion than the VEH and MK-0.025 groups ( $p < 0.05$ ). There was no  
26 difference between the MK-1.0 and MK-0.1 groups ( $p > 0.05$ ). The results also  
27 showed that the MK-0.1 group had higher locomotion than the VEH group  
28 ( $p < 0.05$ ). There was no difference between the MK-0.1 and MK-0.025 groups  
29 ( $p > 0.05$ ) and no difference between the VEH and MK-0.025 groups ( $p > 0.05$ ). On  
30 interval 5 [ $F(3, 20) = 4.83$ ;  $p < 0.01$ ] there was difference among the groups and  
31 Tukey's test showed that the MK-1.0 group had higher locomotion than the VEH

1 group ( $p < 0.05$ ). There was no difference among the MK-1.0, MK-0.1 and MK-  
2 0.025 groups ( $p > 0.05$ ) and no difference among VEH, MK-0.1 and MK-0.25  
3 groups ( $p > 0.05$ ). On interval 6 [ $F(3, 20) = 4.71$ ;  $p < 0.01$ ], the MK-1.0 group had  
4 higher locomotion than the VEH and MK-0.025 groups ( $p < 0.05$ ). There was no  
5 difference between the MK-1.0 and MK-0.1 groups ( $p > 0.05$ ) and no difference  
6 among VEH, MK-0.1 and MK-0.025 groups ( $p > 0.05$ ).

7 Figures 5 A, B and C show the locomotor activity during the 30 min. conditioning  
8 test in experiment 2. For the first 5 min. interval (Fig. 5A), a one-way ANOVA  
9 showed that there was a difference among the groups [ $F(3, 200) = 7.0$ ;  $p < 0.01$ ]  
10 and the Tukey test showed that the MK groups, i. e., MK-0.025, MK-0.1 and MK-  
11 1.0, had higher locomotion than the VEH group ( $p < 0.05$ ). For the 30 min. total  
12 locomotion measurements (Fig. 5B), a one-way ANOVA showed that there was  
13 a difference among the groups [ $F(3, 20) = 6.0$ ;  $p < 0.01$ ] and the Tukey test  
14 showed that the MK-1.0, MK-0.1 and MK-0.025 groups had higher locomotion  
15 than the VEH group ( $p < 0.05$ ). There were no differences among the MK-801  
16 groups ( $p > 0.05$ ). For the within session analysis, the 30 min total locomotion  
17 evaluation was divided into 6 intervals of 5 min. each (Fig. 5C). A repeated two-  
18 way ANOVA showed an interaction group X interval [ $F(15, 100) = 3.0$ ;  $p < 0.01$ ],  
19 an effect of groups [ $F(3, 20) = 5.70$ ;  $p < 0.01$ ] and an effect of intervals [ $F(5, 100)$   
20  $= 60.0$ ;  $p < 0.01$ ]. A one-way ANOVA followed by Tukey test to further analyze the  
21 interaction of groups X intervals showed that on interval 1 [ $F(3, 20) = 7.0$ ;  $p < 0.01$ ]  
22 and interval 2 [ $F(3, 20) = 10.40$ ;  $p < 0.01$ ], the MK-1.0, MK-0.1 and MK-0.025  
23 groups had higher locomotion than the VEH group ( $p < 0.05$ ). There was no  
24 difference among the MK-1.0, MK-0.1 and MK-0.025 groups ( $p > 0.05$ ). On interval  
25 3 [ $F(3, 20) = 1.12$ ;  $p > 0.05$ ], interval 4 [ $F(3, 20) = 0.24$ ;  $p > 0.05$ ] and interval 5 [ $F$   
26  $(3, 20) = 0.40$ ;  $p > 0.05$ ], there was no difference among the groups ( $p > 0.05$ ). On  
27 interval 6 [ $F(3, 20) = 3.44$ ;  $p < 0.01$ ], there was only a difference between the MK-  
28 1.0 and MK-0.025 groups, i. e., the MK-0.025 group had higher locomotion than  
29 the MK-1.0 group ( $p < 0.05$ ).

30 To assess the possibility of possible differential habituation changes between the  
31 MK and VEH groups in experiment 2, Figures 6A and 6B present the changes in

1 locomotor activity during the habituation and the day 9 conditioning test. Figure  
2 6A compares activity levels on habituation day 1 versus habituation day 3. A  
3 repeated two-way ANOVA showed that there was only an effect of days of  
4 treatment [ $F(1, 20) = 65.0$ ;  $p < 0.01$ ]. There was no effect of groups [ $F(3, 20) =$   
5  $0.74$ ;  $p > 0.05$ ] and no interaction groups X days [ $F(3, 20) = 0.52$ ;  $p > 0.05$ ]. The  
6 paired t-test showed that habituation day 1 had higher locomotion than  
7 habituation day 3 [ $t(23) = 8.32$ ;  $p < 0.01$ ]. Thus, all groups showed comparable  
8 habituation. Figure 6B compares habituation day 3 versus conditioning test  
9 session 9. A repeated two-way ANOVA showed that there was a groups X days  
10 interaction [ $F(3, 20) = 3.14$ ;  $p < 0.05$ ] and an effect of days of treatment [ $F(1, 20)$   
11  $= 8.0$ ;  $p < 0.01$ ] but no effect of groups [ $F(3, 20) = 2.40$ ;  $p > 0.05$ ]. The one-way  
12 ANOVA showed on habituation day 3 [ $F(3, 20) = 1.30$ ;  $p > 0.05$ ], that there was  
13 no difference among the groups. However, for the conditioning test on session 9,  
14 there was a difference among the groups [ $F(3, 20) = 5.70$ ;  $p < 0.01$ ] and the  
15 Tukey's test showed that all MK-801 groups had higher locomotion than the VEH  
16 group ( $p < 0.05$ ). The paired t-tests showed that for the VEH group, the activity  
17 level on during the conditioning test day was lower than for habituation day 3 [ $t$   
18  $(5) = 3.04$ ;  $p < 0.05$ ], whereas for all MK-801 groups, there were no differences  
19 ( $p > 0.05$ ) between habituation day 3 and the ninth test session.

20 In that Fig. 6A and Fig. 6B suggest that the MK treatments prevented the  
21 progression of habituation that occurred in the VEH group, Figure 7 presents the  
22 locomotor activity for the 30 minute conditioning test including the 30 min day 3  
23 habituation test for the VEH group. For the within analysis, the 30 min total  
24 locomotion was divided into 6 intervals of 5 min. each. A repeated two-way  
25 ANOVA showed an interaction group X interval [ $F(20, 125) = 2.23$ ;  $p < 0.01$ ], an  
26 effect of groups [ $F(4, 25) = 3.10$ ;  $p < 0.05$ ] and an effect of intervals [ $F(5, 125) =$   
27  $70.74$ ;  $p < 0.01$ ]. A one-way ANOVA followed by the Tukey test to further analyze  
28 the interaction of groups X intervals, showed that on interval 1 [ $F(4, 25) = 5.45$ ;  
29  $p < 0.01$ ], the MK-1.0, MK-0.1 and MK-0.025 groups had higher locomotion than  
30 the VEH group ( $p < 0.05$ ). There was no difference among the VEH-HAB-DAY3,  
31 MK-1.0, MK-0.1 and MK-0.025 groups ( $p > 0.05$ ) On interval 2 [ $F(4, 25) = 4.13$ ;

1 p<0.01], the MK-1.0, MK-0.1 and VEH-HAB-DAY3 groups had higher locomotion  
2 than the VEH group (p<0.05). There was no difference among the VEH-HAB-  
3 DAY3, MK-1.0, MK-0.1 and MK-0.025 groups (p>0.05). On interval 3 [F (4, 25) =  
4 1.43; p>0.05], interval 4 [F (4, 25) = 0.20; p>0.05], interval 5 [F (4, 25) = 0.35;  
5 p>0.05] and interval 6 [F (4, 25) = 2,50; p>0.05], there was no difference among  
6 the groups (p>0.05).

7

## 8 Discussion

9

10 In line with previous reports (Carey *et al.* 1995; Lefevre *et al.* 2016; Wolf and  
11 Khansa 1991), the present study found that MK-801 can induce locomotor  
12 sensitization with repeated treatments. During the pharmacological treatment  
13 phase, sensitization effects were dose related and largely manifested in the MK-  
14 801 1.0 mg/kg group. While the 30 min session totals indicated a rather smooth  
15 increase in locomotion with repeated treatments, the analysis of the locomotor  
16 stimulant effects in successive five-minute intervals during the 30 min test session  
17 indicated a more complicated progression. In fact, the initial treatment (day 1 of  
18 the pharmacological treatment phase) only induced a modest stimulant effect and  
19 only at the highest dose level (1.0 mg/kg) that was limited to the initial five minutes  
20 of the post-injection period. By the second treatment however, potent locomotor  
21 stimulant effects emerged and after only five minutes post-injection. These  
22 stimulant effects expanded to later intervals with additional MK-801 treatments  
23 but the peak remained during the 5-10 min interval. Essentially, the same pattern  
24 of hyperlocomotion was observed in the second experiment. In addition to the  
25 potent stimulant effects induces by the 1.0 MK-801 dose after 4 treatments, a  
26 modest locomotor stimulation developed following the 0.1 mg/kg MK-801  
27 treatments, but no locomotor stimulation occurred in the 0.025 MK-801 treatment  
28 group. The behavioral responses to the different MK-801 doses, which ranged  
29 from no difference from the vehicle injections (0.025 mg/kg MK-801), to the major  
30 hyperlocomotion pattern with the 1.0 mg/kg MK-801 treatment dose, provided the

1 opportunity to investigate the capacity of MK-801 to induce a conditioned drug  
2 response. In Pavlovian conditioning the conditioned response is an attenuated  
3 replica of the unconditioned response. In that the 0.025 dose did not induce a  
4 locomotor response, no conditioned stimulant response was expected while the  
5 profound behavioral effect of the 1.0 mg/kg MK-801 dose would be expected to  
6 induce some type of locomotor stimulant response. On the other hand, if the  
7 NMDA antagonism interfered with the formation of an association, then no  
8 conditioning would be expected in any of the MK-801 groups. Surprisingly in both  
9 experiments, all MK groups expressed equivalent conditioned stimulant  
10 responses.

11 In that the result obtained in the conditioning test does not make sense in terms  
12 of a conditioned drug response, we looked for other possible explanations. While  
13 all groups were given three habituation test sessions prior to the start of drug  
14 treatments, there were an additional five days of testing for the vehicle groups.  
15 This additional testing would have permitted an enhanced degree of habituation  
16 for the vehicle groups. When we examined the three-pre-drug habituation  
17 sessions we found that a strong habituation response occurred in all experimental  
18 groups. Interestingly, when we compared activity levels in the day three  
19 habituation session with the conditioning session, we found that the activity levels  
20 in all MK-801 groups in the conditioning test were the same as their third  
21 habituation session level whereas, the conditioning test activity level of the  
22 vehicle group in the conditioning test was significantly less than the activity level  
23 in the third habituation session. These findings indicated that the MK-801  
24 treatments had interfered with the acquisition of additional habituation over the  
25 course of the 5 MK-801 treatment sessions. It is important to note that the MK-  
26 801 treatment did not reverse the habituation acquired between habituation day  
27 1 and day 3, indicating that MK-801 did not interfere with previously acquired  
28 habituation but rather that MK-801 interfered with the acquisition of new additional  
29 habituation. One possible explanation for the efficacy of MK-801 in preventing the  
30 acquisition of additional habituation is that the exposures to the test environment  
31 with MK-801 were drug state dependent, so that the possible habituation acquired

1 under the MK-801 drug state did not transfer to the non-drug vehicle state. This  
2 explanation suggests that the drug state cues in the 0.025 MK-801 group were  
3 sufficient to induce a complete drug state dependent effect. Another possibility is  
4 suggested by the report (Dai and Carey 1994) that low doses of MK-801 (e.g.  
5 0.03 mg/kg) block attention to external stimuli. In this study the low dose MK-801  
6 treatments that did not affect activity levels prevented the spontaneous  
7 responding of rats to an object placed in an open-field. Whereas non-drug  
8 animals would stop and briefly interact with the object, the MK-801 treated rats  
9 which had similar contacts with the object, did not stop and interact with the object  
10 but rather seemingly ignored the object. In that the MK-801 treated rats' behavior  
11 at the location of the object was the same with or without the object being present  
12 suggested an inattention to the external environment. This could also be viewed  
13 as a general behavioral neglect of external stimuli. This characterization would fit  
14 with other reported effects of MK-801 (Baker and Azorlosa 1996), in which MK-  
15 801 blocked the effects of extinction exposures to conditioned stimuli. Also,  
16 consistent with an MK-801 induced neglect of external stimuli, it has been  
17 reported that MK-801 behavioral sensitization is context independent (Lefevre *et*  
18 *al.* 2016). In that MK-801 generates interoceptive drug stimuli (Grant *et al.* 1996;  
19 Koek *et al.* 1990), then the sensitization would be exclusively associated with the  
20 MK-801 drug stimuli. When MK-801 sensitization is viewed in this way, the  
21 external contextual stimuli paired with the MK-801 sensitization would be ignored  
22 and irrelevant. Consequently, the drug induced response would therefore only be  
23 paired with the drug stimuli. An implication of this analysis is that other drugs that  
24 generate drug stimuli that can substitute for MK-801 in drug discrimination  
25 experiments (Grant *et al.* 1996; Koek *et al.* 1990) seemingly could substitute for  
26 the MK-801 drug stimuli and evoke the behavioral sensitization response (Carey  
27 and Damianopoulos 1993). In addition, the conditioning results obtained in the  
28 present study, wherein the non-motoric 0.025 mg/kg MK-801 dose was as  
29 effective as the hyper-motoric 1.0 mg/kg dose in terms of the conditioned  
30 response, suggests that rats sensitized to the 1.0 MK-801 dose would also  
31 express this sensitization if tested with the 0.025 dose. This would suggest that



1 MK-801 sensitization is associated with MK-801 drug stimuli is a testable  
2 possibility.

3 In conclusion, the behavioral properties of MK-801, such as drug stimuli and MK-  
4 801 induced inattention to external stimuli, provide potential explanations for  
5 important effects of MK-801 that do not require invoking the NMDA receptor in  
6 the mediation of memory processes (Chan *et al.* 2019; Jackson *et al.* 1992). The  
7 reports showing that combined MK-801 dopaminergic drug treatments prevent  
8 the expression of context sensitization to the dopaminergic drugs is readily  
9 explicable by the removal of MK-801 drug stimuli in tests with the dopaminergic  
10 drug. In fact, there have been several reports in which drugs with stimuli different  
11 from dopaminergic drugs that are paired with dopaminergic drug treatments can  
12 acquire control over the behavioral expression of the dopaminergic drug  
13 sensitization effects so that the dopaminergic drug sensitization effects only occur  
14 when the non-dopaminergic drug stimuli are present (Carey *et al.* 1999; Carey *et al.*  
15 *et al.* 2005). While the possible linkage of the NMDA receptor with memory  
16 processes has had heuristic value in initiating behavioral pharmacology  
17 investigations with drugs such as MK-801, the behavioral properties of drugs  
18 such as interoceptive drug stimuli, remain of fundamental importance in the  
19 elucidation of the behavioral drug effects.

20

21 Figure Captions

22

23 Fig. 1: The Means and SEMs for distance scores (M) during the five 30 minute  
24 pharmacological treatment sessions in which VEH or MK-801 (0.025, 0.1 and 1.0  
25 mg/kg) were administered immediately before testing sessions. \*\* denotes  
26 difference for all groups, \*denotes difference from the VEH group. + denotes  
27 difference from VEH group ( $p < 0.05$ ; repeated two-way ANOVA followed by the  
28 Tukey test).

29

1 Fig. 2: The Means and SEMs for distance scores during the 5 minute conditioning  
2 test sessions in which all groups received VEH before testing. \*\* denotes  
3 difference from the VEH group ( $p < 0.05$ ; one-way ANOVA followed by the Tukey  
4 test).

5

6 Fig. 3: The Means and SEMs for distance scores (M) during the five 30 minute  
7 pharmacological treatment sessions in which VEH or MK-801 (0.025, 0.1 and 1.0  
8 mg/kg) were administered immediately before testing sessions. \*\* denotes  
9 difference for all groups, \*denotes difference from the VEH group. ++ denotes  
10 difference for all groups, except the MK-1.0 group. + denotes difference from the  
11 VEH group ( $p < 0.05$ ; repeated two-way ANOVA followed by the Tukey test).

12

13 Fig. 4: The Means and SEMs for distance scores (M) during the within session  
14 locomotor activity during the 5 pharmacological treatment days (A-E) in  
15 experiment 2. The 30 min. total activity score was divided into 6 intervals of 5 min  
16 each. \*\* denotes difference for all groups, \*denotes difference from the VEH  
17 group. ++ denotes difference for all groups, except the MK-1.0 group. + denotes  
18 difference from the VEH group ( $p < 0.05$ ; repeated two-way ANOVA followed by  
19 the Tukey test).

20

21 Fig. 5: The Means and SEMs for distance scores during the 30 minute  
22 conditioning test session. A. Distance scores during the first 5 min. interval.

23 B. Distance scores during the 30 min. total session.

24 C. Within group comparisons in which the 30 min. total activity score was divided  
25 into 6 intervals of 5 min. each.

26 \*denotes difference from the VEH group. + denotes the MK-0.025 group had  
27 higher locomotion than the MK-1.0 group ( $p < 0.05$ ; repeated two-way ANOVA  
28 followed by the Tukey test).

1 Fig. 6: The Means and SEMs for distance scores for changes in locomotor activity  
2 during habituation and the conditioning test (day 9).

3 A. Comparison between habituation day 1 versus habituation day 3.

4 B. Comparison between habituation day 3 versus the conditioning test session  
5 9.

6 \*denotes difference from the VEH group. # denotes difference between the days  
7 ( $p < 0.05$ ; repeated two-way ANOVA followed by the Tukey test or the paired t-  
8 test).

9

10 Fig. 7: The Means and SEMs for distance scores for locomotor activity in the 30  
11 minute conditioning test including the 30 minute habituation day 3 of the VEH  
12 group. For the within analysis, the 30 min. total locomotion was divided into 6  
13 intervals of the 5 min. each. \*\* denotes difference for all groups. \*denotes  
14 difference from VEH group ( $p < 0.05$ ; repeated two-way ANOVA followed by the  
15 Tukey test).

16

## 17 References

18

19 Anagnostaras SG, Robinson TE (1996) Sensitization to the psychomotor  
20 stimulant effects of amphetamine: modulation by associative learning. *Behav*  
21 *Neurosci* 110:1397-1414. <https://doi:10.1037//0735-7044.110.6.1397>.

22

23 Baker JD, Azorlosa JL (1996) The NMDA antagonist MK-801 blocks the  
24 extinction of Pavlovian fear conditioning. *Behav Neurosci* 110:618-620.  
25 <https://doi:10.1037//0735-7044.110.3.618>.

26

1 Bloise E, Carey RJ, Carrera MP (2007) Behavioral sensitization produced by a  
2 single administration of apomorphine: implications for the role of Pavlovian  
3 conditioning in the mediation of context-specific sensitization. *Pharmacol*  
4 *Biochem Behav* 86:449-457. <https://doi:10.1016/j.pbb.2007.01.002>.

5

6 Braga PQ, Dias FR, Carey RJ, Carrera MP (2009) Behavioral sensitization to  
7 dopaminergic inhibitory and stimulatory effects induced by low vs. high dose  
8 apomorphine treatments: an unconventional dose and response reversal  
9 sensitization challenge test reveals sensitization mechanisms. *Behav Brain Res*  
10 204:169-174. <https://doi:10.1016/j.bbr.2009.06.001>.

11

12 Carey RJ, Damianopoulos E, DePalma G (1999) Issues in the pharmacological  
13 modification of cocaine conditioning: evidence that the stimulus properties of  
14 drugs can interact with contextual cues to activate or inactivate cocaine  
15 conditioned stimuli. *Behav Brain Res* 101:189-206. [https://doi:10.1016/s0166-](https://doi:10.1016/s0166-4328(98)00149-1)  
16 [4328\(98\)00149-1](https://doi:10.1016/s0166-4328(98)00149-1).

17

18 Carey RJ, Dai H, Gui J (1998) Effects of dizocilpine (MK-801) on motor activity  
19 and memory. *Psychopharmacology* 137:241-246.  
20 <https://doi:10.1007/s002130050616>.

21

22 Carey RJ, Dai H, Krost M, Huston JP (1995) The NMDA receptor and cocaine:  
23 evidence that MK-801 can induce behavioral sensitization effects. *Pharmacol*  
24 *Biochem Behav* 51:901-908. [https://doi:10.1016/0091-3057\(95\)00074-7](https://doi:10.1016/0091-3057(95)00074-7).

25

26 Carey RJ, Damianopoulos EM (1993) Activation of an apomorphine behavior  
27 pattern by cocaine: a new behavioral test for drug substitution. *J Neurosci*  
28 *Methods* 46:1-10. [https://doi:10.1016/0165-0270\(93\)90135-e](https://doi:10.1016/0165-0270(93)90135-e).

1 Carey RJ, DePalma G, Damianopoulos E, Shanahan A (2005) Stimulus gated  
2 cocaine sensitization: interoceptive drug cue control of cocaine locomotor  
3 sensitization. *Pharmacol Biochem Behav* 82:353-360.  
4 <https://doi:10.1016/j.pbb.2005.09.005>.

5

6 Chan M, Austen JM, Eacott MJ, Easton A, Sanderson DJ (2019) The NMDA  
7 receptor antagonist MK-801 fails to impair long-term recognition memory in mice  
8 when the state-dependency of memory is controlled. *Neurobiol Learn Mem*  
9 161:57-62. <https://doi:10.1016/j.nlm.2019.03.006>.

10

11 Dai H, Carey RJ (1994) The NMDA antagonist MK-801 can impair attention to  
12 exteroceptive stimuli. *Behav Brain Res* 62:149-156. [https://doi:10.1016/0166-](https://doi:10.1016/0166-4328(94)90022-1)  
13 [4328\(94\)90022-1](https://doi:10.1016/0166-4328(94)90022-1).

14

15 Dai H, Carey RJ (1995) Behavioural interaction between the NMDA antagonist  
16 MK-801 and the dopaminergic antagonist haloperidol: support for a balance  
17 model. *J Psychopharmacol* 9:9-15. <https://doi:10.1177/026988119500900102>.

18

19 Damianopoulos EN, Carey RJ (1995) Evidence for N-methyl-D-aspartate  
20 receptor mediation of cocaine induced corticosterone release and cocaine  
21 conditioned stimulant effects. *Behav Brain Res* 68:219-28.  
22 [https://doi:10.1016/0166-4328\(94\)00175-f](https://doi:10.1016/0166-4328(94)00175-f).

23

24 de Matos LW, Carey RJ, Carrera MP (2010) Apomorphine conditioning and  
25 sensitization: the paired/unpaired treatment order as a new major determinant of  
26 drug conditioned and sensitization effects. *Pharmacol Biochem Behav* 96:317-  
27 324. <https://doi:10.1016/j.pbb.2010.05.025>.

28

- 1 Frantz K, Van Hartesveldt C (1999) Locomotion elicited by MK801 in developing  
2 and adult rats: temporal, environmental, and gender effects. *Eur J Pharmacol*  
3 369:145-157. [https://doi:10.1016/s0014-2999\(99\)00070-9](https://doi:10.1016/s0014-2999(99)00070-9).
- 4
- 5 Grant KA, Colombo G, Grant J, Rogawski MA (1996) Dizocilpine-like  
6 discriminative stimulus effects of low-affinity uncompetitive NMDA antagonists.  
7 *Neuropharmacology* 35:1709-19. [https://doi:10.1016/s0028-3908\(96\)00147-5](https://doi:10.1016/s0028-3908(96)00147-5).
- 8
- 9 Jackson A, Koek W, Colpaert FC (1992) NMDA antagonists make learning and  
10 recall state-dependent. *Behav Pharmacol* 3:415-421.  
11 <https://doi:10.1097/00008877-199208000-00018>.
- 12
- 13 Keller S, Delius JD (2001) Discriminative learning occasioned by the  
14 administration of a dopamine agonist. *Psychopharmacology* 157:320-3.  
15 <https://doi:10.1007/s002130100847>.
- 16
- 17 Koek W, Woods JH, Colpaert FC (1990) N-methyl-D-aspartate antagonism and  
18 phencyclidine-like activity: a drug discrimination analysis. *J Pharmacol Exp Ther*  
19 253:1017-1025.
- 20
- 21 Lefevre EM, Eyles DW, Burne TH (2015) Behavioural sensitisation to MK-801 is  
22 dose-dependent and independent of environmental context. *Behav Brain Res*  
23 298:241-245. <https://doi:10.1016/j.bbr.2015.11.014>.
- 24
- 25 Mattingly BA, Gotsick JE, Marin C (1988) Locomotor activity and stereotypy in  
26 rats following repeated apomorphine treatments at 1-, 3-, or 7-day intervals.

1 Pharmacol Biochem Behav 31:871-875. [https://doi:10.1016/0091-](https://doi:10.1016/0091-3057(88)90398-x)  
2 3057(88)90398-x.

3

4 Mattingly BA, Koch C, Osborne FH, Gotsick JE (1997) Stimulus and response  
5 factors affecting the development of behavioral sensitization to apomorphine.  
6 Psychopharmacology 130:109-116. <https://doi:10.1007/s002130050217>.

7

8 Nasello AG, Machado C, Bastos JF, Felicio LF (1998) Sudden darkness induces  
9 a high activity-low anxiety state in male and female rats. *Physiol Behav* 63:451-  
10 454. [https://doi:10.1016/s0031-9384\(97\)00462-9](https://doi:10.1016/s0031-9384(97)00462-9).

11

12 Pinheiro-Carrera M, Tomaz C, Huston JP, Carey RJ (1995) NMDA antagonist  
13 effects on the development of L-dopa behavioral sensitization in rats. *Behav*  
14 *Neurosci* 109:34-42. <https://doi:10.1037//0735-7044.109.1.34>.

15

16 Qi C, Zou H, Zhang R, Zhao G, Jin M, Yu L (2008) Age-related differential  
17 sensitivity to MK-801-induced locomotion and stereotypy in C57BL/6 mice. *Eur J*  
18 *Pharmacol* 580:161-168. <https://doi:10.1016/j.ejphar.2007.07.071>.

19

20 Rowlett JK, Mattingly BA, Bardo MT (1997) Locomotor activity and dopamine  
21 synthesis following 1 and 15 days of withdrawal from repeated apomorphine  
22 treatments. *Pharmacol Biochem Behav* 57:13-8. [https://doi:10.1016/s0091-](https://doi:10.1016/s0091-3057(96)00397-8)  
23 3057(96)00397-8.

24

25 Stewart J, Druhan JP (1993) Development of both conditioning and sensitization  
26 of the behavioral activating effects of amphetamine is blocked by the non-

1 competitive NMDA receptor antagonist, MK-801. *Psychopharmacology* 110:125-  
2 132. [https://doi: 10.1007/BF02246961](https://doi.org/10.1007/BF02246961).

3

4 Vezina P, Queen AL (2000) Induction of locomotor sensitization by amphetamine  
5 requires the activation of NMDA receptors in the rat ventral tegmental area.  
6 *Psychopharmacology* 15:184-91. [https://doi:10.1007/s002130000463](https://doi.org/10.1007/s002130000463).

7

8 Zweifel LS, Argilli E, Bonci A, Palmiter RD (2008) Role of NMDA receptors in  
9 dopamine neurons for plasticity and addictive behaviors. *Neuron* 59:486-496.  
10 [https://doi:10.1016/j.neuron.2008.05.028](https://doi.org/10.1016/j.neuron.2008.05.028).

11

12 Wolf ME, Khansa MR (1991) Repeated administration of MK-801 produces  
13 sensitization to its own locomotor stimulant effects but blocks sensitization to  
14 amphetamine. *Brain Res* 562:164-68. [https://doi:10.1016/0006-8993\(91\)91202-](https://doi.org/10.1016/0006-8993(91)91202-c)  
15 c.

## 16 TABLE. 1

17

**Table 1: Experimental procedure**

**A: Timeline.**

Days	1	2	3	4	5	6	7	8	9	10	11
	Habituation			Pharmacological Treatments					Withdrawal Period		CT

**B: Treatment protocols.**

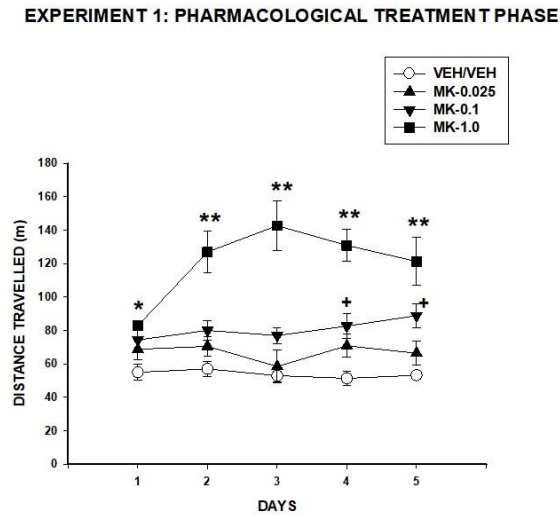
Groups	Pharmacological Treatment	Conditioning Test (CT)
<b>Experiment 1</b>		
	30 min. arena	5 min. arena
VEH (n=5)	VEH	VEH
MK-0.025 (n=5)	MK-0.025	VEH
MK-0.1 (n=5)	MK-0.1	VEH
MK-1.0 (n=5)	MK-1.0	VEH
<b>Experiment 2</b>		
	30 min. arena	30 min. arena
VEH (n=7)	VEH	VEH
MK-0.025 (n=6)	MK-0.025	VEH
MK-0.1 (n=6)	MK-0.1	VEH
MK-1.0 (n=6)	MK-1.0	VEH

CT=Conditioning Test; VEH=vehicle; MK=MK-801.

18



1 FIG. 1

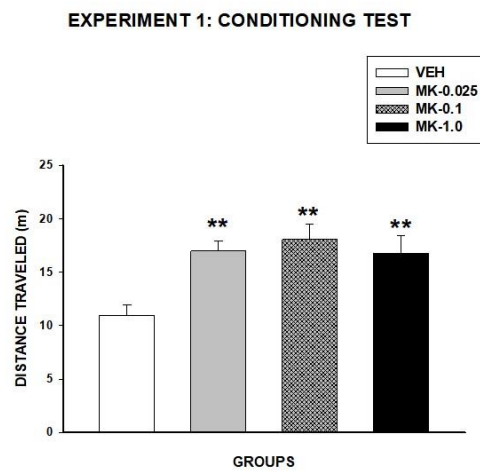


2

3

4

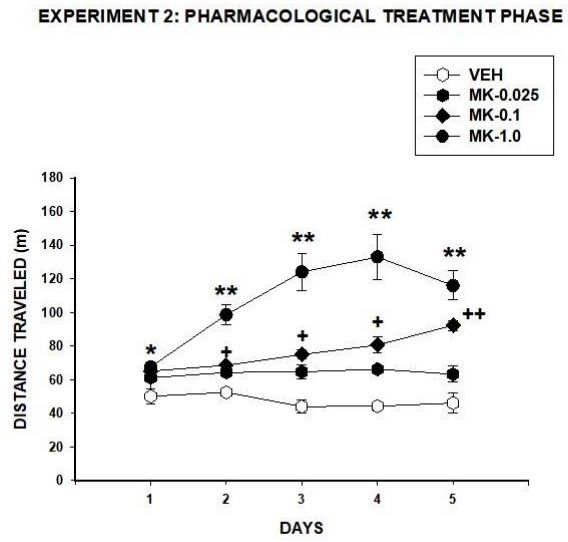
5 FIG. 2



6

7

1 FIG. 3



2

3

4

5

6

7

8

9

10

11

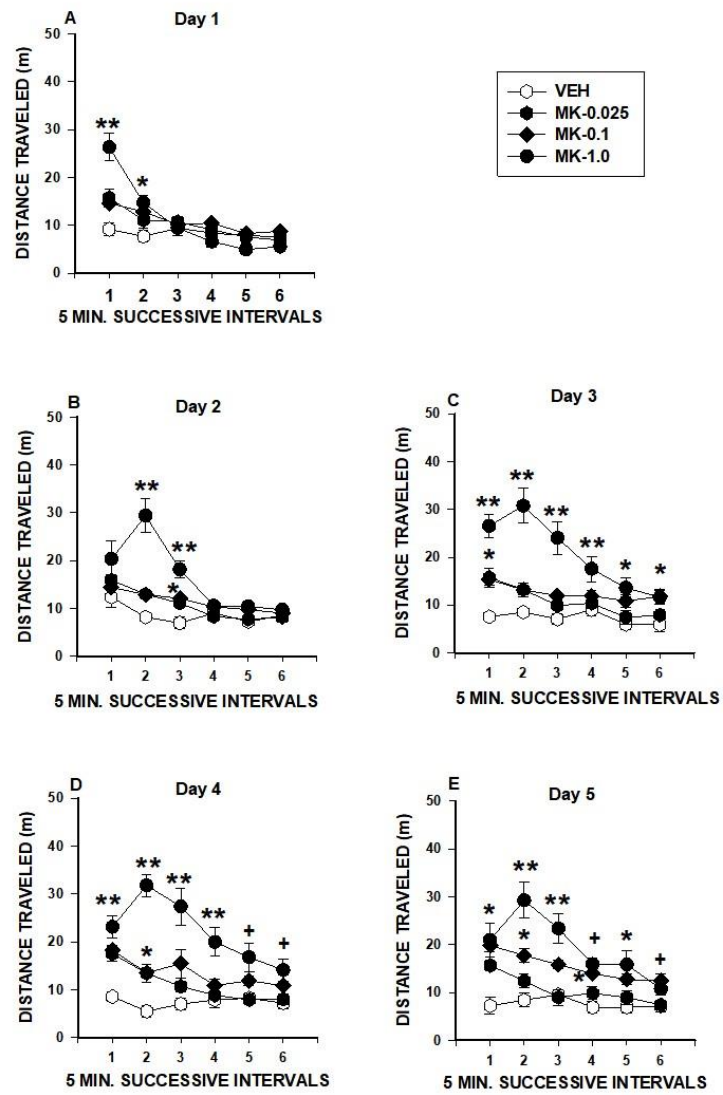
12

13

14

## 1 FIG. 4

## EXPERIMENT 2: PHARMACOLOGICAL TREATMENT PHASE



2

3

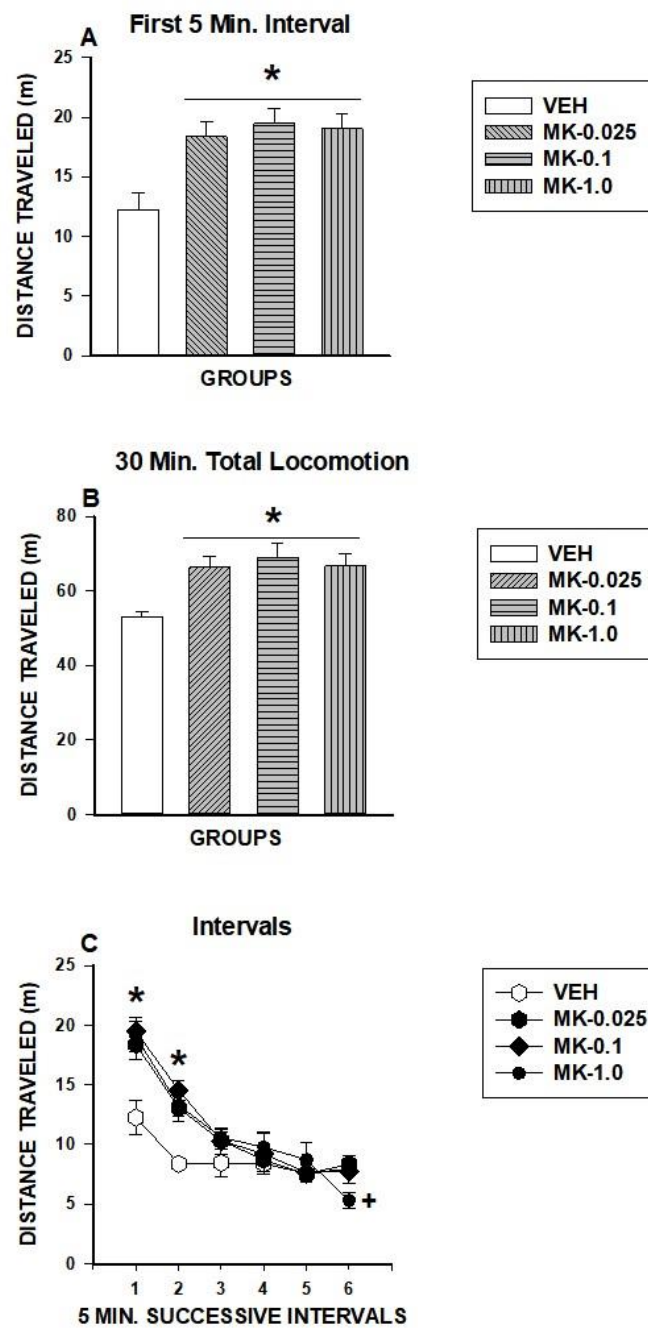
4

5

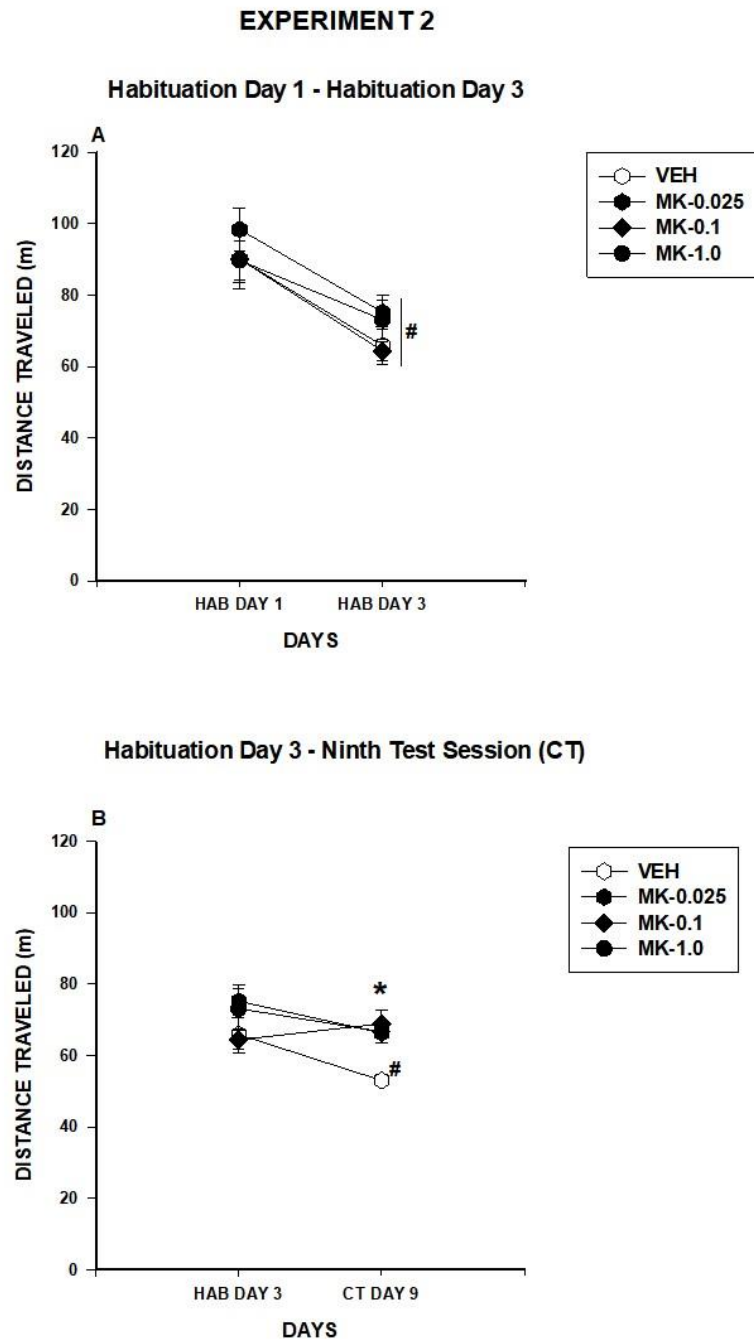
6

1 FIG. 5

## EXPERIMENT 2: CONDITIONING TEST

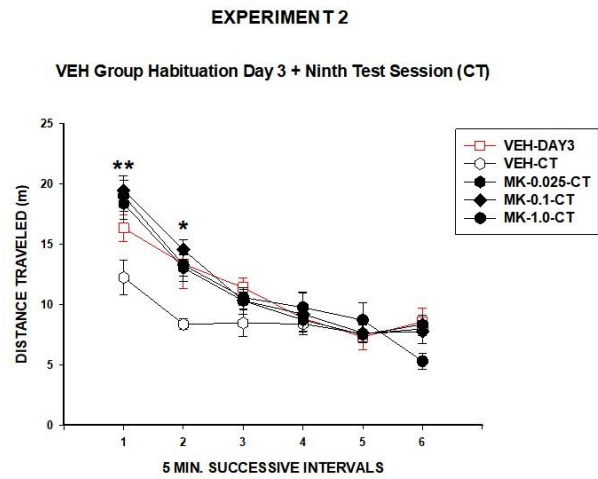


1 FIG. 6



2

3

1 **FIG. 7**

2

3

4

5

6

7

8

9

10

11

12

13

14

15

## 1 **6 - DISCUSSÃO GERAL**

2

3 Curiosamente, com apenas uma exposição ao ambiente experimental, a  
4 resposta comportamental de todos os animais do atual trabalho, incluindo os  
5 grupos dos dois conjuntos experimentais foi semelhante. Em estudos anteriores  
6 (SANTOS e colaboradores 2015) e (LEITE JUNIOR e colaboradores 2019) foram  
7 mostrados os mesmos padrões de resposta.

8 A primeira etapa de cada um dos conjuntos experimentais, a habituação,  
9 foi eficaz em reduzir a locomoção de todos os animais, deixando-os habituados  
10 à arena experimental, pois o efeito da novidade causa um acréscimo nos níveis  
11 circulantes de dopamina, gerando hiperatividade e aumento da locomoção  
12 (COSTA *et al.*, 2014). A morfina e apomorfina são duas drogas com capacidade  
13 de alterar os níveis dopaminérgicos, nesse caso, a influência de uma novidade  
14 pode confundir os resultados (CERBONE e SADILE, 1994).

15 A morfina se mostrou uma droga boa para o estudo da dependência  
16 química, os grupos tratados por 5 dias com morfina 10 mg/kg, tiveram um  
17 aumento da locomoção e sensibilização comportamental, como mostrado nas  
18 figuras 2, 3, 6 e 7 do conjunto experimental 1. Nos testes com apenas uma dose  
19 de morfina (figura 9), não houve aumento da locomoção quando comparado aos  
20 resultados do grupo veículo, mas houve hipolocomoção tanto nos animais do  
21 grupo tratados com Apomorfina e veículo, quanto nos grupos tratados com  
22 apomorfina e morfina, os mesmos resultados foram descritos por Leite Junior e  
23 colaboradores 2019 e Crespo e colaboradores 2022.

24 Após a avaliação do condicionamento e da sensibilização, figura 2 e 3 do  
25 primeiro conjunto, os grupos pré-tratados com apomorfina tiveram o bloqueio dos  
26 efeitos comportamentais comuns no tratamento com morfina, quando o  
27 tratamento foi feito antes da injeção de morfina, da mesma forma essa supressão  
28 foi descrita por Santos e colaboradores 2015; Sanguedo e colaboradores 2015  
29 e 2017 e De Mello e colaboradores 2020. No teste final (figura 7), os tratamentos

1 com apenas uma dose de apomorfina foram eficazes em evitar o aparecimento  
2 dos efeitos do condicionamento previamente registrados com morfina.

3 Algumas estruturas encefálicas foram usadas nesse trabalho para avaliar  
4 posteriormente o condicionamento e sensibilização, sendo que as regiões do  
5 NAc e do VTA mostraram esses detalhes, descrito por Kobrin e colaboradores  
6 2017. A morfina foi capaz de promover uma maior ativação da resposta ERK 1/2  
7 no VTA e NAc. Conforme os dados expostos nas figuras 4, 5, 8 e 10 do conjunto  
8 experimental 1 nos mostram que, houve maior ativação de ERK nos grupos  
9 experimentais que receberam morfina na fase de indução (figura 4 e 5), mesmo  
10 naqueles grupos que receberam veículo no teste final, essa observação nos  
11 mostra que o condicionamento gerado pela morfina é duradouro e pode ocorrer  
12 na ausência da droga, na forma de hiperlocomoção que reflete numa maior  
13 ativação da ERK (DIAS *et al.*, 2021). A ativação da ERK foi bloqueada nos  
14 animais que receberam pré-tratamentos com apomorfina, sendo essa  
15 quantificação similar à encontrada no grupo veículo. No processo do uso de  
16 drogas e dependência ocorre uma maior ativação da via mesocorticolímbica, que  
17 se traduz em maior plasticidade em regiões responsáveis pelo prazer como o  
18 NAc (VOLLSTÄDT-KLEIN *et al.*, 2011; KANTAK e c DHONNCHADHA *et al.*,  
19 2011). Dessa forma o paciente tem dificuldade em ter sucesso no tratamento pra  
20 inibir os estímulos de saliência deixados pelo uso de psicoativos, como por  
21 exemplo os opioides (THOMPSON *et al.*, 2020). Quando foi usado um protocolo  
22 experimental com apenas uma injeção de morfina (figuras 9 e 10 - experimento  
23 3), não foi possível observar aumento da locomoção, mas houve maior ativação  
24 de ERK. Curiosamente pode ser que ocorra um aumento da plasticidade  
25 neuronal, que antecede o aumento da locomoção, ou seja, o reflexo ao uso de  
26 drogas não ocorre de forma aguda no comportamento, mas ocorre mudanças  
27 em regiões encefálicas como o VTA e NAc, similar ao observado por Bender e  
28 Torregrossa, 2021 e Baimel e colaboradores 2019.

29 Aqui, os dados mostram que o MK-801 induz a sensibilização locomotora  
30 após tratamentos repetidos, o que corrobora com os resultados encontrados  
31 (WOLF E KHANSA 1991; CAREY *et al.*, 1995 e LEFEVRE *et al.*, 2016). Durante



1 o tratamento farmacológico, fase de indução mostrada nos gráficos das figuras  
2 1 e 4, os efeitos de sensibilização foram relacionados à dose alta e amplamente  
3 manifestados no grupo MK-801 1,0 mg/kg, como vistos em (TANG *et al.*, 2006).  
4 Antagonistas NMDA como exemplo o MK-801, podem emular os sintomas  
5 psicóticos da Esquizofrenia, como a hiper locomoção, que também são comuns  
6 ao dependente químico (SNYDER e GAO, 2013). Quanto aos diferentes  
7 tratamentos com MK-801, houve diferença na locomoção dos animais que  
8 receberam as doses altas (1.0 mg/kg) e os animais tratados com as doses  
9 médias (0,1 mg/kg). A sensibilização locomotora nos animais tratados com 1.0  
10 mg/kg ocorreu logo ao 3º dia, de tratamento de indução, e continuou sendo  
11 observada até o 5º dia, por outro lado os animais do grupo 0,1 mg/kg mostraram  
12 sensibilização locomotora mais demorada, essa só ocorreu no quinto dia de  
13 experimentação.

14 O fenômeno da sensibilização é muito importante para que um indivíduo  
15 se torne um dependente químico, essa sensibilização foi encontrada também por  
16 Cui e colaboradores 2014, com o uso de altas doses de MK-801 (0,5 mg/kg).  
17 Entretanto os animais que receberam MK-801 em baixa dose (0,025 mg/kg), não  
18 houve diferença na locomoção se comparados ao grupo veículo, essa ausência  
19 de efeito locomotor também foi observada recentemente por Chen e  
20 colaboradores 2022.

21 Um estudo Imunoistoquímico das estruturas encefálicas como o HIPO,  
22 VTA, NAc, CPF e AM, relacionadas à dependência assim como foi feito com o  
23 uso de apomorfina e morfina, se faz necessário para que os dados quanto ao  
24 uso do MK-801 sejam melhor compreendidos.

25

26

27

28

29

## 1 **7 - CONCLUSÃO**

2

3 Os tratamentos com morfina 10 mg/kg, de forma associada ao contexto  
4 ambiental, com uma duração de 5 dias, produziram tanto o condicionamento  
5 quanto a sensibilização comportamental;

6 A morfina 10 mg/kg produz uma memória associativa, desenvolvida pela  
7 associação do contexto ambiental como os efeitos da droga e também maior  
8 ativação da ERK no VTA e Nac;

9 Os pré-tratamentos com apomorfina eliminaram tanto os efeitos  
10 comportamentais da morfina no condicionamento e sensibilização, quanto a  
11 ativação da morfina da resposta ERK no VTA e NAc;

12 Os tratamentos pré-arena com MK-801 na dose de 1.0 mg/kg, produziram  
13 tanto sensibilização quanto condicionamento ambiental;

14 Para os efeitos da morfina, uma maior atividade do sistema  
15 dopaminérgico é dependente dos efeitos desinibidores do GABA;

16 Os resultados dos tratamentos com apomorfina fornecem suporte  
17 substancial para a importância o nível de atividade dos neurônios  
18 dopaminérgicos para a eficácia da desinibição da morfina nos sistemas de  
19 recompensa da dopamina na mediação da sensibilização comportamental e dos  
20 efeitos condicionados;

21 Portanto, para uma melhor compreensão dos dados relacionados ao MK-  
22 801, se faz necessário um estudo Imunoistoquímico da ERK nas estruturas  
23 encefálicas como o HIPO, VTA, NAc, CPF e AM, que estão relacionadas à  
24 dependência, assim como foi feito com o uso de apomorfina e morfina. Este  
25 questionamento é de grande interesse para investigações em experimentos  
26 futuros.

27

28

1 **8 - REFERÊNCIAS BIBLIOGRÁFICAS**

2

3 ADINOFF, B. Neurobiologic processes in drug reward and addiction. **Harvard**  
4 **Review of Psychiatry**. v. 12, n. 06, p. 305 – 320, 2004.

5

6 ALEXANDER, M.D., NITESH, K.J., MOHIT, G. Opioid use disorder. **StatPearls**  
7 **Publishing**, Treasure Island, FL, USA. 2020.

8

9 ANAGNOSTARAS, S.G., ROBINSON, T.E. Sensitization to the psychomotor  
10 stimulant effects of amphetamine: modulation by associative learning.  
11 **Behavioral Neuroscience**. V. 110, p. 1397-1414, 1996.

12

13 ASENSIO, V.J., MIRALLES, A., GARCÍA-SEVILLA, J.A. Stimulation of mitogen-  
14 activated protein kinase kinases (MEK1/2) by  $\mu$ -,  $\delta$ - and  $\kappa$ -opioid receptor  
15 agonists in the rat brain: Regulation by chronic morphine and opioid withdrawal.  
16 **European Journal of Pharmacology**. v. 539, n. 01 - 02, p. 49 – 56, 2006.

17

18 ARAÚJO, M.R., MOREIRA, F.G., SILVEIRA, D.X., MOREIRA, F.G. Histórias das  
19 drogas, Panorama atual de drogas e dependências. **São Paulo: Atheneu**. Ed.  
20 01, n.01, p. 09 – 14, 2006.

21

22 BAIMEL, C., MCGARRY, L.M., CARTER, A.G. The Projection Targets of Medium  
23 Spiny Neurons Govern Cocaine-Evoked Synaptic Plasticity in the Nucleus  
24 Accumbens. **Cell Reports**. v. 28, n. 09, p. 2256 - 2263, 2019.

25

26 BARAKA, A. Historical aspects of opium. **Middle East Journal of**  
27 **Anesthesiology**. v. 15, n. 04, p. 423 – 436, 2000.

28

29 BARDO, M.T., BEVINS, R.A. Conditioned place preference: what does it add to  
30 our preclinical understanding of drug reward? **Psychopharmacology (Berl)**. v.  
31 153, n. 01, p. 31 – 43, 2000.

32

- 1 BENDER, B.N., TORREGROSSA, M.M. Dorsolateral striatum dopamine-  
2 dependent cocaine seeking is resistant to pavlovian cue extinction in male and  
3 female rats. **Neuropharmacology**. v. 182, n. 108403, 2021.
- 4
- 5 BERKE, J.D. What does dopamine mean? **Nature Neuroscience**. v. 21, n. 06,  
6 p.787 – 793, 2018.
- 7
- 8 BLOISE, E., CAREY, R.J., CARRERA, M.P. Behavioral sensitization produced  
9 by a single administration of apomorphine: implications for the role of Pavlovian  
10 conditioning in the mediation of context-specific sensitization. **Pharmacology**  
11 **Biochemistry and Behavior** v. 86, p. 449-457, 2007.
- 12
- 13 BONSI, P., CUOMO, D., DE PERSIS, C., CENTONZE, D., BERNARDI, G.,  
14 CALABRESI, P., PISANI, A. Modulatory action of metabotropic glutamate  
15 receptor (mGluR) 5 on mGluR1 function in striatal cholinergic interneurons.  
16 **Neuropharmacology**. v. 49, n. 01, p. 104 – 113, 2005.
- 17
- 18 BOWIE, D. Ionotropic Glutamate Receptors & CNS Disorders. **CNS &**  
19 **Neurological Disorders - Drug Targets**. v. 07, n. 02, p. 129 – 143, 2008.
- 20
- 21 BRAGA, P.Q., DIAS, F.R., CAREY, R.J., CARRERA, M.P. Behavioral  
22 sensitization to dopaminergic inhibitory and stimulatory effects induced by low vs.  
23 high dose apomorphine treatments: an unconventional dose and response  
24 reversal sensitization challenge test reveals sensitization mechanisms.  
25 **Behavioural Brain Research**. v. 204, n. 01, p. 169 – 174, 2009.
- 26
- 27 BROWNSTEIN, M.J. A brief history of opiates, opioid peptides, and opioid  
28 receptors. **Proceedings of the National Academy of Sciences**. v. 90, n. 12, p.  
29 5391 – 5393, 1993.
- 30
- 31 CADOR, M., BJIJOU, Y., STINUS, L. Evidence of a complete Independence of  
32 the neurobiological substrates for the induction and expression of behavioral  
33 sensitization to amphetamine. **Neuroscience**. v.65, n. 02, p. 385 – 395, 1995.
- 34

- 1 CAHILL, E., SALERY, M., VANHOUTTE, P., CABOCHE, J. Convergence of  
2 dopamine and glutamate signaling onto striatal ERK activation in response to  
3 drugs of abuse. **Frontiers in Pharmacology**. v. 04, n. 172, pages: 13, 2014.
- 4
- 5 CAMARGO, B., DELLA SERA, L., CAMARGO, M. Participacion del núcleo  
6 entopeduncular (globo pálido interno) en los procesos de aprendizaje. **Revista**  
7 **médica de Panamá**. v. 06, n. 03, p. 221 – 232, 1981.
- 8
- 9 CAREY, R.J., DAI, H., KROST, M., HUSTON, J.P. The NMDA receptor and  
10 cocaine: evidence that MK-801 can induce behavioral sensitization effects.  
11 **Pharmacology Biochemistry and Behavior**. 51: 901-908, 1995.
- 12
- 13 CAREY, R.J., CARRERA, M.P., DAMIANOPOULOS, E.N. A new proposal for  
14 drug conditioning with implications for drug addiction: The Pavlovian two-step  
15 from delay to trace conditioning. **Behavioural Brain Research**. v. 275, p. 150 –  
16 156, 2014.
- 17
- 18 CARRERA, M.P, TOMAZ, C., HOUSTON, J.P., CAREY, R.J. NMDA Antagonist  
19 Effects on the Development of L-DOPA Behavioral Sensitization in Rats.  
20 **Behavioral Neuroscience**. v. 109, n. 01, p. 34 – 42, 1995.
- 21
- 22 CARRERA, P.M., CAREY, R.J., DIAS, F.R.C., SAMPAIO, F.S., MATOS, L.W.  
23 Post trial apomorphine at an autoreceptor dose level can eliminate apomorphine  
24 conditioning and sensitization: Support for the critical role of dopamine in re-  
25 consolidation. **Behavioural Brain Research**. v. 236, p. 244 –250, 2013.
- 26
- 27 CATANIA, A.C. **Aprendizagem: comportamento, linguagem e cognição**  
28 **Porto Alegre: Artmed**. Capítulo 12: Comportamento Respondente:  
29 Condicionamento, 1999.
- 30
- 31 CEUA-UNIFESP. Guia de anestesia e analgesia em animais de laboratório.  
32 **Unifesp - São Paulo**, SP. n. 01, 2017.
- 33

- 1 CERBONE, A., SADILE, A.G. Behavioral habituation to spatial novelty:  
2 interference and noninterference studies. **Neuroscience & Biobehavioral**  
3 **Reviews**. v. 18, n. 04, p. 497 – 518, 1994.
- 4
- 5 CFMV, Guia Brasileiro de Boas Práticas em Eutanásia em Animais - Conceitos  
6 e Procedimentos Recomendados – **CFMV - Brasília, DF**. 2012.
- 7
- 8 CHANG, H.Y., KHARRAZI, H., BODYCOMBE, D., WEINER, J.P., ALEXANDER,  
9 G.C. Healthcare costs and utilization associated with high-risk prescription opioid  
10 use: a retrospective cohort study. **BMC Medicine**. v. 16, n. 01, p. 01 – 11, 2018.
- 11
- 12 CHARTOFF, E.H., CONNERY, H.S. It's MORE exciting than mu: crosstalk  
13 between mu opioid receptors and glutamatergic transmission in the mesolimbic  
14 dopamine system. **Frontiers in Pharmacology**. v. 05, n. 116, p. 01 – 22, 2014.
- 15
- 16 CHEN, M., ZHAO, Y., YANG, H., LUAN, W., SONG, J., CUI, D., DONG, Y., LAI,  
17 B., MA, L., ZHENG, P. Morphine disinhibits glutamatergic input to VTA dopamine  
18 neurons and promotes dopamine neuron excitation. **eLife**. v. 04, pages: 25, 2015.
- 19
- 20 CHEN, P.A., WANG, H.Y., SUN, C.L., CHEN, M.L., CHEN, Y.C. Neurobehavioral  
21 Differences of Valproate and Risperidone on MK-801 Inducing Acute  
22 Hyperlocomotion in Mice. *Behavioural Neurology*. v. 1048463, 2022.
- 23
- 24 CICCARELLI, A., CALZA, A., SANTORU, F., GRASSO, F., CONCAS, A.,  
25 SASSOÈ-POGNETTO, M., GIUSTETTO, M. Morphine withdrawal produces  
26 ERK-dependent and ERK-independent epigenetic marks in neurons of the  
27 nucleus accumbens and lateral septum. **Neuropharmacology**. v. 70, n. 01, p.  
28 168 – 179, 2013.
- 29
- 30
- 31 COLLINS, E.D., WARD, A.S., MCDOWELL, D.M., FOLTIN, R.W., FISCHMAN,  
32 M.W. The effects of memantine on the subjective, reinforcing and cardiovascular  
33 effects of cocaine in humans. **Behavioural Pharmacology**. v. 09, n. 07, p. 587  
34 – 598, 1998.

- 1
- 2 COHEN, J.J. Disorders of potassium balance. **Journal Hospital Practice**. v. 14,  
3 n. 01, p. 119 – 128, 1979.
- 4
- 5 CONTI, F. EAAC1, a high-affinity glutamate transporter, is localized to astrocytes  
6 and gabaergic neurons besides pyramidal cells in the rat cerebral cortex.  
7 **Cerebral Cortex**. v. 08, n. 02, p. 108 – 116, 1998.
- 8
- 9 COSTA, V.D., TRAN, V.L., TURCHI, J., AVERBECK, B.B. Dopamine modulates  
10 novelty seeking behavior during decision making. **Behavioral Neuroscience**. v.  
11 128, n. 05, p. 556 – 566, 2014.
- 12
- 13 CRESPO, L.G.S.C., LEITE JÚNIOR, J.B., DEMELLO BASTOS, J.M., SAMUELS,  
14 R.I., COIMBRA, N.C., CAREY, R.J., CARRERA, M.P. Context evoked morphine  
15 conditioned effects can be equivalent to morphine induced drug effects in terms of  
16 behavioral response and ERK activation in reward associated subcortical brain  
17 structures. **Pharmacology Biochemistry And Behavior**. v.214, p.173356,  
18 2022.
- 19
- 20 CUNHA P.J., NOVAES M. A. Neurocognitive assessment in alcohol abuse and  
21 dependence: implications for treatment. **Brazilian Journal of Psychiatry**. v. 26,  
22 n. 01, p. 23 – 27, 2004.
- 23
- 24 DAHAN, A., KEST B., WAXMAN, A.R., SARTON, E. Sex-specific responses to  
25 opiates: Animal and human studies. **Anesthetic Pharmacology Preclinical  
26 Pharmacology**. v. 107, n. 01, p. 83 – 95, 2008.
- 27
- 28 DAI, H., CAREY, R.J. Behavioural interaction between the NMDA antagonist MK-  
29 801 and the dopaminergic antagonist haloperidol: support for a balance model.  
30 **Journal of Psychopharmacology** 9:9-15, 1995.
- 31
- 32 DAVIE, J.R., SPENCER, V.A. Signal transduction pathways and the modification  
33 of chromatin structure. **Progress in Nucleic Acid Research and Molecular  
34 Biology**. v. 65, n. 01, p. 299 – 340, 2000.

- 1 DE MATOS, L.W., CAREY, R.J., CARRERA, M.P. Apomorphine conditioning  
2 and sensitization: the paired/unpaired treatment order as a new major  
3 determinant of drug conditioned and sensitization effects. **Pharmacology,**  
4 **Biochemistry, and Behavior**, n. 96, p. 317-324, 2010.
- 5
- 6 DE MELLO BASTOS, J.M., DIAS, F.R., ALVES, V.H., CAREY RJ, CARRERA  
7 MP. Drug memory substitution during re-consolidation: a single inhibitory  
8 autoreceptor apomorphine treatment given during psychostimulant memory re-  
9 consolidation replaces psychostimulant conditioning with conditioned inhibition  
10 and reverses psychostimulant sensitization. **Behavioural Brain Research** n.  
11 260, p.139 - 47, 2014.
- 12
- 13 DE MELLO BASTOS J.M., LEITE JUNIOR J.B., SAMUELS R.I., CAREY R.J.,  
14 CARRERA M.P. Post-trial low dose apomorphine prevents the development of  
15 morphine sensitization. **Behavioural Brain Research**. v. 380, n. 112398, 2020.
- 16
- 17 DE MORAIS, N.A., RAFFAELLI, M., KOLLER, S.H. Adolescentes em situação  
18 de vulnerabilidade social e o continuum risco-proteção. **Avances en Psicología**  
19 **Latinoamericana**. v. 30, n. 1, p. 118–136, 2012.
- 20
- 21 DHAWAN, B.N., CESSSELIN, F., RAGHUBIR, R., REISINE, T., BRADLEY P.B.,  
22 PORTOGHESE, P.S., HAMON, M. International Union of Pharmacology. XII.  
23 Classification of opioid receptors. **Pharmacological Reviews**. v. 48, n. 04, p.  
24 567-92, 1996.
- 25
- 26 DIAS F.P., CRESPO L.G.S.C., LEITE JUNIOR J.B., SAMUELS R.I., COIMBRA  
27 N.C., CAREY R.J., CARRERA M.P. Morphine reward effects and morphine  
28 behavioral sensitization: The adventitious association of morphine activation of  
29 brain reward effects with ongoing spontaneous activity, **Pharmacology,**  
30 **Biochemistry and Behavior**. v. 209, n. 173244, 2021.
- 31
- 32 DIAZ, J. How Drugs Influence Behavior: A Neurobehavioral Approach. **Pearson**  
33 **College Division**; 1<sup>o</sup> edition, pages: 285, 1996.
- 34



- 1 DOBI, A., MARGOLIS, E.B., WANG, H.L., HARVEY, B.K., MORALES, M.  
2 Glutamatergic and Nonglutamatergic Neurons of the Ventral Tegmental Area  
3 Establish Local Synaptic Contacts with Dopaminergic and Nondopaminergic  
4 Neurons. **Journal of Neuroscience**. v. 30, n. 01, p. 218 – 229, 2010.  
5
- 6 D'SOUZA, M.S. Glutamatergic transmission in drug reward: implications for drug  
7 addiction. **Frontiers in Neuroscience**. v. 09, n. 404, pages: 27, 2015.  
8
- 9 DUARTE, D.F. Opium and opioids: a brief history. **Revista Brasileira de**  
10 **Anestesiologia**. v. 55, n. 01, p. 135 – 146, 2005.  
11
- 12 DUNCAN, C.P. Habit reversal induced by electroshock in the rat. **Journal of**  
13 **Comparative and Physiological Psychology**. v. 41, n. 01, p. 11 – 16, 1948.  
14
- 15 DUPONT, R.L. Cérebro, Álcool e Drogas, O Cérebro Egoísta: Aprender Com as  
16 Dependências. **Instituto Piaget**. Ed. 12, n. 01, p. 01 – 544, 2005.  
17
- 18 EISCH, A.J., BARROT, M., SCHAD, C.A., SELF, D.W., NESTLER, E.J. Opiates  
19 inhibit neurogenesis in the adult rat hippocampus. **Proceedings of the National**  
20 **Academy of Sciences of the United States of America**. v. 97, n. 13, p. 7579  
21 – 7584, 2000.  
22
- 23 ENGELHARDT, E. O sistema glutamatérgico e desordens neuropsiquiátricas:  
24 aspectos básicos. **Revista Brasileira de Neurologia**. v. 39, n. 02, p. 05 – 19.  
25 2003.  
26
- 27 FEATHERSTONE, D.E. Intercellular Glutamate Signaling in the Nervous System  
28 and Beyond. **ACS Chemical Neuroscience**. v. 01, n. 01, p. 04 – 12, 2010.  
29
- 30 FERREIRA, M.P., LEITE, M.C., HOCHGRAF, P.B., ZILBERMAN, M.L.,  
31 CORDÁS, T.A., MORENO, R.A. Dependências Químicas, Conduas em  
32 psiquiatria. **Lemos Editorial**. Ed. 04, n. 01, p. 319 – 348, 2001.  
33

- 1 FONTES, A., FIGLIE, N.B., LARANJEIRA, R. O comportamento de beber entre  
2 dependentes de álcool: estudo de seguimento. **Revista de psiquiatria clínica**,  
3 v. 33, n. 06, p. 304-312, 2006.
- 4
- 5 FORD, L.M., NORMAN, A.B., SANBERG, P.R. The topography of MK-801-  
6 induced locomotor patterns in rats. **Physiology & Behavior**. 46, 755–758, 1989.
- 7
- 8 FOX, C.A., ANDRADE, A.N., DU QUI, I.J., RAFOLS, J.A. The primate globus  
9 pallidus: a Golgi and electron microscopic study. **Journal fur Hirnforschung**. v.  
10 15, n. 01, p. 95 – 126, 1974.
- 11
- 12 FRANTZ, K., VAN HARTESVELDT, C. Locomotion elicited by MK801 in  
13 developing and adult rats: temporal, environmental, and gender effects.  
14 **European Journal of Pharmacology**. v. 369, n. 02, p. 145 – 157, 1999.
- 15
- 16 GARCIA-BORREGUERO, D., USHA, K.M., KATILA, J., KUMAR, S., PRASAD,  
17 S., ANNE-MARIE, W., LARROSA, O. Treatment of restless legs syndrome with  
18 pregabalin: A double-blind, placebo-controlled study. **American Academy of**  
19 **Neurology**. v. 76, n.04, p. 408 – 409, 2011.
- 20
- 21 GIRAULT, J., VALJENT, E., CABOCHE, J., HERVE, D. ERK2: a logical AND  
22 gate critical for drug-induced plasticity? **Current Opinion in Pharmacology**. v.  
23 07, n. 01, p. 77 – 85, 2007.
- 24
- 25 GLARE, P.A., WALSH, T.D. Clinical pharmacokinetics of morphine. **Therapeutic**  
26 **Drug Monitoring**. v. 13. n. 01, p. 01 – 23, 1991.
- 27
- 28 GOLDBERG, J.H., FARRIES, M.A., FEE, M.S. Basal ganglia output to the  
29 thalamus: still a paradox. **Trends In Neurosciences**. v. 36, n. 12, p. 695 – 705,  
30 2013.
- 31
- 32 GRAY, A.M., RAWLS, S.M., SHIPPENBERG, T.S., & MCGINTY, J.F. The K-  
33 Opioid Agonist, U-69593, Decreases Acute Amphetamine-Evoked Behaviors and

- 1 Calcium-Dependent Dialysate Levels of Dopamine and Glutamate in the Ventral  
2 Striatum. **Journal of Neurochemistry**. v. 73, n. 03, p. 1066 – 1074, 2001.
- 3
- 4 HEMMINGS, H.C., JEVTOVIC-TODOROVIC, V. Special issue on anaesthetic  
5 neurotoxicity and neuroplasticity. **British Journal of Anesthesia**. v. 110, n. 01,  
6 p. 01 – 02, 2013.
- 7
- 8 HENKE, J., HABERSTROH, J., SAGER, M., BECKER, K., EBERSPÄCHER, E.,  
9 BERGADANO, A., ZAHNER, D., ARRAS, M. Pain management for laboratory  
10 animals. **Committee on Anaesthesia of the GV Solas**. v. 01, 69 pages, 2016.
- 11
- 12 HINDLEY, A., KOLCH, W. Extracellular signal regulated kinase (ERK)/mitogen  
13 activated protein kinase (MAPK)-independent functions of Raf kinases. **Journal**  
14 **of Cell Science**. v. 115, n. 08, p. 1575 – 1581, 2002.
- 15
- 16 HOFFORD, R.S., HODGSON, S.R., ROBERTS, K.W., BRYANT, C.D., EVANS,  
17 C.J., EITAN, S. Extracellular signal-regulated kinase activation in the amygdala  
18 mediates elevated plus maze behavior during opioid withdrawal. **Behavioural**  
19 **Pharmacology**. v. 20, v. 07, p. 576 – 583, 2009.
- 20
- 21 HUECKER, M.R., LEAMING, J.M. Opioid Addiction. **StatPearls - Treasure**  
22 **Island (FL)**. 2020.
- 23
- 24 HYMAN, S.E., MALENKA, R.C. Addiction and the brain: the neurobiology of  
25 compulsion and its persistence. **Nature Reviews Neuroscience**. v. 02, n. 10, p.  
26 695 – 703, 2001.
- 27
- 28 IZQUIERDO, L.A., BARROS, D.M., VIANNA, M.R., COITINHO, A., DEDAVID,  
29 E., SILVA, T., CHOI, H., MOLETTA, B., MEDINA, J.H., IZQUIERDO, I. Molecular  
30 pharmacological dissection of short-and long-term memory. **Celular and**  
31 **Molecular Neurobiology**. v. 22, n. 03, p. 269 – 287, 2002.
- 32

- 1 IZQUIERDO, I., BEVILAQUA, L.R., ROSSATO, J.I., BONINI, J.S., MEDINA, J.H.,  
2 CAMMAROTA, M. Different molecular cascades in different sites of the brain  
3 control memory Consolidation. **Trends in Neuroscience**. v. 29, n. 09, p. 496 –  
4 505, 2006.
- 5
- 6 JACOBS, E.H., SMITH, A.B., De VRIES, T.J., SCHOFFELMEER, A. N.  
7 Neuroadaptive effects of active versus passive drug administration in addiction  
8 research. **Trends Pharmacological Sciences**. v. 24, n. 11, p. 566 – 573, 2003.
- 9
- 10 JOHNSON, G.L., LAPADAT, R. Mitogen-Activated Protein Kinase Pathways  
11 Mediated by ERK, JNK, and p38 Protein Kinases. **Science**. v. 298, n. 5600, p.  
12 1911 – 1912, 2002.
- 13
- 14 JONAS, P., BISCHOFBERGER, J., AND SANDKUHNER, J. Corelease of two  
15 fast neurotransmitters at a central synapse. **Science**. v. 281, p. 419 - 424, 1998.
- 16
- 17 KANTAK, K.M., NICHONCHADHA, B.Á. Pharmacological enhancement of  
18 drug cue extinction learning: translational challenges. **Annals of the New York  
19 Academy of Sciences**. v. 1216, p. 122 - 137, 2011.
- 20
- 21 KELLER, S., DELIUS, J.D. Discriminative learning occasioned by the  
22 administration of a dopamine agonist. **Psychopharmacology** n. 157, p. 320-330,  
23 2001.
- 24
- 25 KENNA, G.A., NIELSEN, D.M., MELLO, P., SCHIESL, A., SWIFT, R. M.  
26 Pharmacotherapy of dual substance abuse and dependence. **CNS Drugs**. v. 21,  
27 n. 03, p. 213 – 237, 2007.
- 28
- 29 KIM, P.C., YOO, J.W., COCHRAN, C.R., PARK, S.M., CHUN, S., LEE, Y.J.,  
30 SHEN, J.J. Trends and associated factors of use of opioid, heroin, and cannabis  
31 among patients for emergency department visits in Nevada: 2009–2017.  
32 **Observatory Study – Medicine (Baltimore)**. v. 98, n. 47, pages: 09, 2019.
- 33

- 1 KOBRIK, K.L., ARENA, D.T., HEINRICHS, S.C.; NGUYEN, O.H., KAPLAN, G.B.  
2 Dopamine D1 receptor agonist treatment attenuates extinction of morphine  
3 conditioned place preference while increasing dendritic complexity in the nucleus  
4 accumbens core. **Behavioural Brain Research**. v. 322, p. 18 - 28, 2017.
- 5
- 6 KRAWCZYK, N., GREENE, M.C., ZORZANELLI, R., BASTOS, F.I. Rising Trends  
7 of Prescription Opioid Sales in Contemporary Brazil, 2009–2015. **American**  
8 **Journal of Public Health**. v. 108, n. 05, p. 666 – 668, 2018.
- 9
- 10 KREEK, M.J., PATRICK, E., HAGGERTY, B.M. Opioids, dopamine, stress, and  
11 the addictions. **Dialogues in Clinical Neuroscience**. v. 09, n. 04, p. 363 – 378,  
12 2007.
- 13
- 14 KOLODNY, A., COURTWRIGHT, D.T., HWANG, C.S., KREINER, P., EADIE,  
15 J.L., CLARK, T.W., ALEXANDER, G.C. The Prescription Opioid and Heroin  
16 Crisis: A Public Health Approach to an Epidemic of Addiction. **Annual Review of**  
17 **Public Health**. v. 36, n. 01, p. 559 – 574, 2015.
- 18
- 19 LEFEVRE, E.M., EYLES, D.W., BURNE, T.H. Behavioural sensitisation to MK-  
20 801 is dose-dependent and independent of environmental context. **Behavioural**  
21 **Brain Research**. v. 298, p. 241 - 245, 2015.
- 22
- 23 LEITE JUNIOR J.B., DE MELLO BASTOS J.M., SAMUELS R.I., CAREY R.J.,  
24 CARRERA M.P., Reversal of morphine conditioned behavior by an anti-  
25 dopaminergic post-trial drug treatment during reconsolidation, **Behavioural**  
26 **Brain Research**. v. 359, p. 771 - 782, 2019.
- 27
- 28 LEWIS, T.S., SHAPIRO, P.S., AHN, N.G. Signal Transduction through MAP  
29 Kinase Cascades. **Advances in Cancer Research**. v. 74, n. 01, p. 49 – 139,  
30 1998.
- 31
- 32 LI, C.T., YANG, K.C., LIN, W.C. Glutamatergic Dysfunction and Glutamatergic  
33 Compounds for Major Psychiatric Disorders: Evidence From Clinical  
34 Neuroimaging Studies. **Frontiers in Psychiatry**. v. 09, n. 767, pages: 11, 2019.

- 1 LÖTSCH, J., GEISLINGER, G. Morphine-6-glucuronide: an analgesic of the  
2 future? **Clinical Pharmacokinetics**. v. 40, n. 07, p. 485 – 499, 2001.
- 3
- 4 LUGO, R.A., KERN, S.E. Clinical pharmacokinetics of morphine. **Journal of Pain  
5 & Palliative Care Pharmacotherapy**. v. 16, n. 04, p. 05 – 18, 2002.
- 6
- 7 MANN, K., GÜNTHER, A., STETTER, F., ACKERMANN, K. Rapid recovery from  
8 cognitive deficits in abstinent alcoholics: a controlled test-retest study. **Alcohol  
9 and Alcoholism**. v. 34, n. 04, p. 567 – 574, 1999.
- 10
- 11 MARTINS, E.R.C., CORREA, A.K. Dealing with psychoactive substances: the  
12 meaning for nursing workers. **Revista Latino-Americana de Enfermagem**. v.  
13 12, n. 01, p.398 – 405, 2004.
- 14
- 15 MARTINS, R.T., ALMEIDA, D.B. de, MONTEIRO, F.M. do R., KOWACS, P.A.;  
16 RAMINA, R. Receptores opioides até o contexto atual. **Instituto de Neurologia  
17 de Curitiba**. v. 13, n. 01, p. 75 – 79, 2012.
- 18
- 19 MATTINGLY, B.A., GOTSICK, J.E., MARIN, C. Locomotor activity and stereotypy  
20 in rats following repeated apomorphine treatments at 1-, 3-, or 7-day intervals.  
21 **Pharmacology Biochemistry and Behavior** 31:871-875, 1988.
- 22
- 23 MATTINGLY, B.A., KOCH, C., OSBORNE, F.H., GOTSICK, J.E. Stimulus and  
24 response factors affecting the development of behavioral sensitization to  
25 apomorphine. **Psychopharmacology**, v. 130, p. 109-116, 1997.
- 26
- 27 MATSUMOTO K., YOSHIDA M., ANDERSSON K., HEDLUND P. Effects in vitro  
28 and in vivo by apomorphine in the rat corpus cavernosum. **British Journal of  
29 Farmacology**. v. 146, n. 02, p. 259 – 267, 2005.
- 30
- 31 MELDRUM, B.S. Glutamate as a Neurotransmitter in the Brain: Review of  
32 Physiology and Pathology. **The Journal of Nutrition**. v. 130, n. 04, p. 1007 –  
33 1015, 2000.

- 1 MICZEK, K.A., NIKULINA, E.M., SHIMAMOTO, A., COVINGTON, H.E.  
2 Escalated or Suppressed Cocaine Reward, Tegmental BDNF, and Accumbal  
3 Dopamine Caused by Episodic versus Continuous Social Stress in Rats. **Journal**  
4 **of Neuroscience**. v. 31, n. 27, p. 9848 – 9857, 2011.
- 5 MILTON, A.L., EVERITT, B.J. The persistence of maladaptative memory:  
6 addiction, drug memories and anti-relapsed treatments. **Neuroscience &**  
7 **Biobehavioral Reviews**, v. 36, p. 1119-1139, 2012.
- 8
- 9 MITRA, S., SINATRA, R.S. Perioperative Management of Acute Pain in the  
10 Opioid-dependent Patient. **Anesthesiology**. v. 101, n. 01, p. 212 – 227, 2004.
- 11
- 12 MORGANE, P.J., GALLER, J.R., MOKLER, D.J. A review of systems and  
13 networks of the limbic forebrain/limbic midbrain. **Progress in Neurobiology**. v.  
14 75, n. 02, p. 143 – 160, 2005.
- 15
- 16 MORRISON, D.K. MAP Kinase Pathways. **Cold Spring Harbor Perspectives in**  
17 **Biology**. v. 04, n. 11, pages: 06, 2012.
- 18
- 19 NABEKURA, J., KATSURABAYASHI, S., KAKAZU, Y., SHIBATA, S.,  
20 MATSUBARA, A., JINNO, S., MIZOGUCHI, Y., SASAKI, A., AND ISHIBASHI, H.  
21 Developmental switch from GABA to glycine release in single central synaptic  
22 terminals. **Nature Neuroscience**. v. 07, p. 17–23, 2004.
- 23
- 24 NARITA, M., HASHIMOTO, K., AMANO, T., NARITA, M., NIIKURA, K.,  
25 NAKAMURA, A., SUZUKI, T. Post-synaptic action of morphine on glutamatergic  
26 neuronal transmission related to the descending antinociceptive pathway in the  
27 rat thalamus. **Journal of Neurochemistry**. v. 104, n. 02, p. 469 – 478, 2008.
- 28
- 29 NASCIMENTO, P.F.C. do. Efeitos da exposição ao etanol em camundongos  
30 adolescentes e adultos: comportamentos relacionados à recompensa,  
31 sensibilização comportamental e o papel dos sistemas dopaminérgico e  
32 glutamatérgicos. São Paulo, **Instituto de Ciências Biomédicas da**  
33 **Universidade de São Paulo**. 2011.
- 34

- 1 NERY FILHO, A., MACRAE, E., TAVARES, L.A., RÊGO, M. Toxicomanias:  
2 incidências clínicas e socioantropológicas. **EDUFBA Drogas: clínica e cultura**  
3 **collection**. 1º ed, n. 01, p. 01 – 308, 2009.
- 4
- 5 NESTLER, E.J. From neurobiology to treatment: progress against addiction.  
6 **Nature Neuroscience**. v. 05, n. 01, p. 1076 – 1079, 2002.
- 7
- 8 NESTLER, E.J. Historical review: Molecular and cellular mechanisms of opiate  
9 and cocaine addiction. **Trends in Pharmacological Sciences**. v. 25, n. 04, p.  
10 210 – 218, 2004.
- 11
- 12 NIKULINA, E., COVINGTON, H., GANSCHOW, L., HAMMER, R., MICZEK, K.  
13 Long-term behavioral and neuronal cross-sensitization to amphetamine induced  
14 by repeated brief social defeat stress: Fos in the ventral tegmental area and  
15 amygdala. **Neuroscience**. v. 134, n. 04, p. 857 – 865, 2004.
- 16
- 17 NODA, Y., MIYAMOTO, Y., MAMIYA, T., KAMEI, H., FURUKAWA, H.,  
18 NABESHIMA, T. Involvement of dopaminergic system in phencyclidine-induced  
19 place preference in mice pretreated with phencyclidine repeatedly. **The Journal**  
20 **of Pharmacology and Experimental Therapeutics**. v. 288, n. 01, p. 44 – 51,  
21 1998.
- 22
- 23 NODA, Y., NABESHIMA, T. Involvement of signal transduction cascade via  
24 dopamine-D1 receptors in phencyclidine dependence. **ANNALS**. v. 1025, n. 01,  
25 p. 62 – 68, 2004.
- 26
- 27 NUNES, L., JÓLLUSKIN, G., O uso de drogas breve análise histórica e social,  
28 revista da faculdade de ciências humanas e sociais. **Revista da Faculdade de**  
29 **Ciências Humanas e Sociais**. v. 04, n. 01, p. 230 – 237, 2007.
- 30
- 31 O'BRIEN, C.P., CHILDRESS, A.R., EHRMAN, R., ROBBINS, S.J. Conditioning  
32 factors in drug abuse: can they explain compulsion? **Journal Of**  
33 **Psychopharmacology**. v. 12. n. 01, p. 14 – 22, 1998.
- 34



- 1 ÖGREN, S.-O., GOLDSTEIN, M. Phencyclidine- and dizocilpine-in-duced  
2 hyperlocomotion are differentially mediated. **Neuropsychopharmacology** 11,  
3 167–177, 1994.
- 4
- 5 OLIVE, M.F., CLEVA, R.M., KALIVAS, P.W., MALCOLM, R.J. Glutamatergic  
6 medications for the treatment of drug and behavioral addictions. **Pharmacology**  
7 **Biochemistry and Behavior**. v. 100, n. 04, p. 801 – 810, 2012.
- 8
- 9 ORDOÑEZ, R.A.S., Reconsolidação da Memória e a Dependência de Estado:  
10 Mecanismos da Atualização. Dissertação Mestrado. **Universidade Federal do**  
11 **Rio Grande do Sul**. Porto Alegre – RS, 2012.
- 12
- 13 PATHAN, H., WILLIAMS, J. Basic opioid pharmacology: an update. **British**  
14 **Journal of Pain**. v. 06, n. 01, p. 11 – 16, 2012.
- 15
- 16 PAVLOV I. P. Conditioned Reflexes. **Oxford, Oxford University Press**. 1927.
- 17
- 18 PÉREZ-CAJARAVILLE, J., ABEJÓN, D., ORTIZ, J.R., PÉREZ, J.R. El dolor y su  
19 tratamiento a través de la historia. **Revista de la Sociedad Española del Dolor**.  
20 v. 12, n. 06, p. 373 – 384, 2005.
- 21
- 22 PERT, C.B., SNYDER, S.H. Opiate receptor: demonstration in nervous tissue.  
23 **Science**. v. 179, n. 4077, p. 1011 – 1014, 1973.
- 24
- 25 PETROFF, O.A. GABA and glutamate in the human brain. **The Neuroscientist**.  
26 v. 08, n. 06, p. 562 – 573, 2002.
- 27
- 28 PIERCE, R.C., BARI, A.A. The Role of Neurotrophic Factors in Psychostimulant-  
29 induced Behavioral and Neuronal Plasticity. **Reviews in the Neurosciences**. v.  
30 12, n. 02, p. 95 – 110, 2001.
- 31

- 1 RAITH, K., HOCHHAUS, G. Drugs used in the treatment of opioid tolerance and  
2 physical dependence: a review. **Journal of Clinical Pharmacology and**  
3 **Therapeutics**. v. 42, n. 04, p. 191 – 203, 2004.
- 4
- 5 RAWLS, S.M., MCGINTY, J.F. Delta opioid receptors regulate calcium-  
6 dependent, amphetamine-evoked glutamate levels in the rat striatum: an in vivo  
7 microdialysis study. **Brain Research**. v. 861, n. 02, p. 296 – 304, 2000.
- 8
- 9 REEDS, P.J., BURRIN, D.G., STOLL, B., JAHOOOR, F. Intestinal Glutamate  
10 Metabolism. **The Journal of Nutrition**. v. 130, n. 04, p. 978 – 982, 2000.
- 11
- 12 REINER, A., LEVITZ, J. Glutamatergic Signaling in the Central Nervous System:  
13 Ionotropic and Metabotropic Receptors in Concert. **Neuron**. v. 98. N. 06, p. 1080  
14 – 1098, 2018.
- 15
- 16 REHMAN, I., REHMAN, C.I. Classical Conditioning. **Treasure Island,**  
17 **StatPearls**. 2018.
- 18
- 19 RIBEIRO, F.M.L., MINAYO, M.C.S. Religious therapeutic communities in  
20 recovering drug users: the case of Manguinhos, RJ. **Interface**. v. 19, n. 54, p.  
21 515 – 526, 2015.
- 22
- 23 RIVERA, A., GAGO, B., FUXE, K., BRENÉ, S., DÍAZ-CABIALE, Z., REINA-  
24 SÁNCHEZ, M.D., SUÁREZ-BOOMGAARD, D., ROALES-BUJÁN,  
25 R.; VALDERRAMA-CARVAJAL, A., DE LA CALLE, A. Early modulation by the  
26 dopamine D4 receptor of morphine-induced changes in the opioid peptide  
27 systems in the rat caudate putamen. **Journal of Neuroscience Research**. v. 91,  
28 n. 12, p. 1533 – 1540, 2013.
- 29
- 30 ROBBINS, T.W., EVERITT, B.J. Drug addiction: bad habits add up. **Nature**. v.  
31 398, p. 567 – 70, 1999.
- 32

- 1 ROBBINS, T.W., EVERITT, B.J. Limbic-striatal memory systems and drug  
2 addiction. **Neurobiology of Learning na Memory**. v. 78, n. 03, p. 625 – 636,  
3 2002.
- 4
- 5 ROBINSON, T.E., BECKER, J.B. Enduring changes in brain and behavior  
6 produced by chronic amphetamine administration: a review and evaluation of  
7 animal models of amphetamine psychosis. **Brain Research Reviews**. v. 396, n.  
8 02, p. 157 – 189, 1986.
- 9
- 10 ROBINSON, T.E., BERRIDGE, K.C. The neural basis of drug craving: An  
11 incentive-sensitization theory of addiction. **Brain Research Reviews**. v. 18, n.  
12 03, p. 247 – 291, 1993.
- 13
- 14 ROBINSON, T.E., KOLB, B. Alterations in the morphology of dendrites and  
15 dendritic spines in the nucleus accumbens and prefrontal cortex following  
16 repeated treatment with amphetamine or cocaine. **European Journal of**  
17 **Neuroscience**. v. 11, n. 05, p. 1598 – 1604, 1999.
- 18
- 19 ROWLETT, J.K., MATTINGLY, B.A., BARDO, M.T. Locomotor activity and  
20 dopamine synthesis following 1 and 15 days of withdrawal from repeated  
21 apomorphine treatments. **Pharmacology Biochemistry and Behavior**, v. 57, p.  
22 13-8, 1997.
- 23
- 24 RUGGIERO, R.N., BUENO-JÚNIOR, L.S., ROSS, J.B. DE, FACHIM, H.A.,  
25 PADOVAN-NETO, F.E., MERLO, S., ROHNER, C.J.S., IKEDA, E.T., BRUSCO,  
26 J., MOREIRA, J.E. Neurotransmissão glutamatérgica e plasticidade sináptica:  
27 aspectos moleculares, clínicos e filogenéticos. **Revista Medicina Ribeirão**  
28 **Preto, SP**. v. 44, n. 02, p. 143 – 156, 2011.
- 29
- 30 SAKAE, D.Y., MARTI, F., LECCA, S., VORSPAN, F., MARTÍN-GARCÍA, E.,  
31 MOREL, L.J., HENRION, A., GUTIÉRREZ-CUESTA, J., BESNARD, A., HECK,  
32 N., HERZOG, E., BOLTE, S., PRADO, V.F., PRADO, M.A.M., BELLIVIER, F.,  
33 EAP, C.B., CRETTOL, S., VANHOUTTE, P., CABOCHE, J., GRATTON, A.,  
34 MOQUIN, L., GIROS, B., MALDONADO, R., DAUMAS, S., MAMELI, M.,  
35 JAMAIN, S., EL MESTIKAWY, S. The absence of VGLUT3 predisposes to

- 1 cocaine abuse by increasing dopamine and glutamate signaling in the nucleus  
2 accumbens. **Molecular Psychiatry**. v. 20, p. 1448 – 1459, 2015.
- 3
- 4 SALAMONE, J.D., CORREA, M. The Mysterious Motivational Functions of  
5 Mesolimbic Dopamine. **Neuron**. v. 76, n. 03, p. 470 – 485, 2012.
- 6
- 7 SANGUEDO, F.V., DIAS, C.V.B., DIAS, F.R.C., SAMUELS, R.I., CAREY, R.J.,  
8 CARRERA, M.P. Reciprocal activation/inactivation of ERK in the amygdala and  
9 frontal cortex is correlated with the degree of novelty of an open-field  
10 environment. **Psychopharmacology**. v. 233, n. 05, p. 841 – 850, 2015.
- 11
- 12 SANGUEDO, F.V., FIGUEIREDO, A.M., CAREY, R.J., SAMUELS, R.I.,  
13 CARRERA, M.P. ERK activation in the prefrontal cortex by acute apomorphine  
14 and apomorphine conditioned contextual stimuli. **Pharmacology Biochemistry  
15 and Behavior**. v. 159, n. 01, p. 76 – 83, 2017.
- 16
- 17 SANTOS, B.G., CAREY, R.J., CARRERA, M.P. Post-trial induction of  
18 conditioned apomorphine stimulant and inhibitory response effects: evidence for  
19 potent trace conditioning of drug effects. **Pharmacology Biochemistry and  
20 Behavior**. v.129 p. 79 - 86, 2015.
- 21 SCHOLL, L., SETH, P., KARIISA, M., WILSON, N., BALDWIN, G. Drug and  
22 opioid-involved overdose deaths. **Morbidity and Mortality Weekly Report**. v.  
23 67, n. 5152, p. 1419 – 1427, 2019.
- 24
- 25 SCHROLL, H., HAMKER, F.H. Computational models of basal-ganglia pathway  
26 functions: focus on functional neuroanatomy. **Frontiers In Systems  
27 Neuroscience**. v. 07, n. 122, p. 01 – 18, 2013.
- 28
- 29 SCHULTZ, D.P., SCHULTZ, S.E. **História da Psicologia Moderna**. Tradução  
30 de Adail Ubirajara Sobral e Marta Stela Gonçalves, 9<sup>o</sup>. Ed., Editora Cultrix, 1992.
- 31
- 32 SCHULTZ, W., APICELLA, P., LJUNGBERG, T. Responses of monkey  
33 dopamine neurons to reward and conditioned stimuli during successive steps of

- 1 learning a delayed response task. **The Journal of Neuroscience**. v. 13, n. 03,  
2 p. 900 – 913, 1993.
- 3
- 4 SCHULTZ, W. Dopamine signals for reward value and risk: basic and recent data.  
5 **Behavioral and Brain Functions**. v. 06, n. 01, pages: 09, 2010.
- 6
- 7 SCHWARTZ, T.L., SACHDEVA, S., STAHL, S.M. Glutamate Neurocircuitry:  
8 Theoretical Underpinnings in Schizophrenia. *Frontiers in Pharmacology*. v. 03, n.  
9 195, pages: 11, 2012.
- 10
- 11 SEE, R.E. Neural substrates of conditioned-cued relapse to drug-seeking  
12 behavior. **Pharmacology Biochemistry and Behavior**. v. 71, n. 03, p. 517 –  
13 529, 2002.
- 14
- 15 SHALEV, U., GRIMM, J.W., YAVIN SHAHAM, Y. Neurobiology of Relapse to  
16 Heroin and Cocaine Seeking: A Review. **Pharmacological Reviews**. v. 54, n.  
17 01, p. 01 – 49, 2002.
- 18
- 19 SHIFLETT, M.W., BALLEINE, B.W. Contributions of ERK signaling in the striatum  
20 to instrumental learning and performance. **Behavioural Brain Research**. v. 218,  
21 n. 01, p. 240 – 247, 2011.
- 22
- 23 SHOBLOCK, J.R., WICHMANN, J., MAIDMENT, N.T. The effect of a systemically  
24 active ORL-1 agonist, Ro 64-6198, on the acquisition, expression, extinction, and  
25 reinstatement of morphine conditioned place preference. **Neuropharmacology**.  
26 v. 49, n. 04, p. 439 – 446, 2005.
- 27
- 28 SIEGEL, S. Pavlov Conditioning and drug overdose: when tolerances fail.  
29 **Addiction Research & Theory**, v. 09, n. 05, p. 503 – 513, 2001.
- 30
- 31 SJØGREN, P., JENSEN, N.H., JENSEN, T.S. Disappearance of morphine-  
32 induced hyperalgesia after discontinuing or substituting morphine with other  
33 opioid agonists. **Pain**. v. 59, n. 02, p. 313 – 316, 1994.

- 1 SMITH, M.T. Neuroexcitatory effects of morphine and hydromorphone: evidence  
2 implicating the 3-glucuronide metabolites. **Clinical and Experimental**  
3 **Pharmacology and Physiology**. v. 27, n. 07, p. 524 – 528, 2000.
- 4
- 5 SNYDER, M. A.; GAO, W. NMDA hypofunction as a convergence point for  
6 progression and symptoms of schizophrenia. **Frontiers in Cellular**  
7 **Neuroscience**. v. 7, n. 31, p. 1 – 12, 2013.
- 8
- 9 SOMALWAR, A.R., CHOUDHARY, A.G., SHARMA, P.R., SAGARKAR, B.N.,  
10 SAKHARKAR, A.J., SUBHEDAR, N.K., KOKARE, D.M. Cocaine- and  
11 amphetamine-regulated transcript peptide (CART) induced reward behavior is  
12 mediated via G(i/o) dependent phosphorylation of PKA/ERK/CREB pathway.  
13 **Behavioural Brain Research**. v. 384, n. 01 p. 09 – 21, 2018.
- 14
- 15 SPENCER, S., SCOFIELD, M., KALIVAS, P.W. The good and bad news about  
16 glutamate in drug addiction. **Journal of Psychopharmacology**. v. 30, n. 11, p.  
17 1095 – 1098, 2016.
- 18
- 19 STADDON, J.E., CERUTTI, D.T. Operant conditioning. **Annual Review of**  
20 **Psychology**. v. 54, n. 01, p. 115 – 144, 2003.
- 21
- 22 STAHL, S.M. **Essential Psychopharmacology**. 3<sup>a</sup> ed, New York, NY:  
23 Cambridge University Press. 2008.
- 24
- 25 STEIN, C. Opioid Receptors. **Annual Review of Medicine**. v. 67, n. 01, p. 433 –  
26 451, 2015.
- 27
- 28 STEWART, J., BADIANI, A. Tolerance and sensitization to the behavioral effects  
29 of drugs. **Behav Pharmacology**. v. 04, n. 04, p. 289 – 312, 1993.
- 30
- 31 TAMELINI, M.G., MARTINS, A.C.P., CAVALCANTI, E.F.A., MARTINS, H.S.  
32 Abuso e dependência de substâncias psicoativas, Clínica médica: dos sinais e

- 1 sintomas ao diagnóstico e tratamento. **Manole**. Ed. 01, n. 01, p. 1050 – 1064,  
2 2007.
- 3
- 4 TANG, Y., ZOU, H., STRONG, J.A., CUI, Y., XIE, Q., ZHAO, G., ... YU, L.  
5 Paradoxical effects of very low dose MK-801. **European Journal of**  
6 **Pharmacology**. v. 537, n. 01 – 03, p. 77 – 84, 2006.
- 7
- 8 THOMPSON, B.L., OSCAR-BERMAN, M., KAPLAN, G.B. Opioid-induced  
9 Structural and Functional Plasticity of Medium-Spiny Neurons in the Nucleus  
10 Accumbens. **Neuroscience & Biobehavioral Reviews**. v. 120, p. 417 – 430,  
11 2020.
- 12
- 13 TRECOT, A.M., DATTA, S., LEE, M., HANSEN, H. Opioid pharmacology. **Pain**  
14 **Physician**. v. 11, n. 02, p. 133 – 153, 2008.
- 15
- 16 TRUDEAU, L.E., HNASKO, T.S., WALLÉN-MACKENZIE, A., MORALES, M.,  
17 RAYPORTK, S., SULZERK, D. The multilingual nature of dopamine neurons.  
18 **Progress in Brain Research**. v. 211, p. 141 – 164, 2014.
- 19
- 20 UNODC, United Nations Office on Drugs and Crime, **World Drug Report -**  
21 **United Nations Publication**. Ch. 02, n. 16. XI. 07, 2016.
- 22
- 23 UNODC, United Nations Office on Drugs and Crime, Calculations based on  
24 monitoring reports of illicit crops and responses to questionnaires for annual  
25 reports. **World Drug Report - United Nations Publication**. Ch. 02, n. 18. XI. 07,  
26 2018.
- 27 UNODC, United Nations Office on Drugs and Crime, **World Drug Report -**  
28 **United Nations publication**, Ch. 02, n. 19. XI. 08, 2019.
- 29
- 30 UNODC - United Nations Office on Drugs and Crime, **World Drug Report -**  
31 **United Nations Publication**. Ch. 02, n. 16. XI. 07, 2022.
- 32

- 1 VANDERSCHUREN, L.J., TJON, G.H., NESTBY, P., MULDER, A.H.,  
2 SCHOFFELMEER, A.N., & VRIES, T.J. Morphine-induced long-term  
3 sensitization to the locomotor effects of morphine and amphetamine depends on  
4 the temporal pattern of the pretreatment regimen. **Psychopharmacology**. v.  
5 131, n. 02, p. 115 – 122, 1997.
- 6
- 7 VANDERSCHUREN, L.J., SCHMIDT, E.D., DE VRIES, T.J., VAN MOORSEL,  
8 C.A., TILDERS, F.J., SCHOFFELMEER, A.N. A single exposure to amphetamine  
9 is sufficient to induce long-term behavioral, neuroendocrine, and neurochemical  
10 sensitization in rats. **Journal of Neuroscience**. v. 19, n. 21, p. 9579 – 9586,  
11 1999.
- 12 VANDERSCHUREN, L.J., VRIES, T.J., WARDEH, G., HOGENBOOM, F.A.,  
13 SCHOFFELMEER, A. N. A single exposure to morphine induces long-lasting  
14 behavioural and neurochemical sensitization in rats. **European Journal of**  
15 **Neuroscience**. v. 14, n. 09, p. 1533 – 1538, 2001.
- 16
- 17 VARGA, E.V., NAVRATILOVA, E., STROPOVA, D., JAMBROSIC, J., ROESKE,  
18 W.R., YAMAMURA, H.I. Agonist-specific regulation of the  $\delta$ -opioid receptor. **Life**  
19 **Sciences**. v. 76, n. 06, p. 599 - 612. 2004.
- 20
- 21 VIGANÒ, D., RUBINO, T., DI CHIARA, G., ASCARI, I., MASSI, P. & PAROLARO,  
22 D.  $\mu$  Opioid eceptor signaling in morphine sensitization. **Neuroscience**. v. 117,  
23 n. 04, p. 921 – 929, 2003.
- 24
- 25 VOLLSTÄDT-KLEIN, S., LOEBER, S., KIRSCH, M., BACH, P., RICHTER, A.,  
26 BÜHLER, M., VON DER GOLTZ, C., HERMANN, D., MANN, K., KIEFER, F.  
27 Effects of Cue-Exposure Treatment on Neural Cue Reactivity in Alcohol  
28 Dependence: A Randomized Trial. **Biol Psychiatry**. v. 69, n. 11, p. 1060 - 1066,  
29 2011.
- 30
- 31 WAHLSTRÖM, A., PACIFICI, G.M., LINDSTRÖM, B., HAMMAR, L., RANE, A.  
32 Human liver morphine UDP-glucuronyl transferase enantioselectivity and  
33 inhibition by opioid congeners and oxazepam. **British Journal of**  
34 **Pharmacology**. v. 94, n. 03, p. 864 – 870, 1988.
- 35



- 1 WALTER, S., KUSCHINSKY, K. Conditioning of morphine-induced locomotor  
2 activity and stereotyped behaviour in rats. **Journal of Neural Transmission**  
3 **General Section**. v. 78, n. 03, p. 231 – 247, 1989.
- 4
- 5 WATANABE, M., MAEMURA, K., KANBARA, K., TAMAYAMA, T., HAYASAKI,  
6 H. GABA and GABA receptors in the central nervous system and other organs.  
7 **International Review of Cytology**. v. 213, n. 01, p. 01 – 47, 2002.
- 8
- 9 WELLS, M.J.; WELLS, J. Conditioning and sensitization in snails. **Animal**  
10 **Behaviour**. v.19, n. 02 p.305 – 312, 1971.
- 11
- 12 WICKENS, J.R. Synaptic plasticity in the basal ganglia. **Behavioural Brain**  
13 **Research**. v. 199, n. 01, p. 119 – 128, 2009.
- 14
- 15 WISE, R.A., ROMPRE, P.P. Brain Dopamine and Reward. **Annual Review of**  
16 **Psychology**. v. 40, n. 01, p. 191 – 225, 1989.
- 17
- 18 WISE, R.A. Addictive drugs and brain stimulation reward. **Annual Reviews**  
19 **Neuroscience**. v. 19, n. 01, p. 319 – 340, 1996.
- 20
- 21 WISE, R.A. Addiction becomes a brain disease. **Neuron**. v. 26, n. 01, p. 27 – 33,  
22 2000.
- 23
- 24 WOLF, M.E., KHANSA, M.R. Repeated administration of MK-801 produces  
25 sensitization to its own locomotor stimulant effects but blocks sensitization to  
26 amphetamine. **Behavioural Research**. v. 562, p. 164 - 168, 1991.
- 27
- 28 WYLLIE, D.J.A., LIVESEY, M.R., HARDINGHAM, G.E. Influence of GluN2  
29 subunit identity on NMDA receptor function. **Neuropharmacology**. v. 74, n. 01,  
30 p. 04 – 17, 2013.
- 31

1 YADAV, S.K., PRAKASH, J., CHOUHAN, S., WESTFALL, S., VERMA, M.,  
2 SINGH, T.D., SINGH, S.P. Comparison of the neuroprotective potential of  
3 *Mucuna pruriens* seed extract with estrogen in 1-methyl-4-phenyl-1,2,3,6-  
4 tetrahydropyridine (MPTP) - induced PD mice model. **Neurochemistry**  
5 **International**. v. 65, p. 01 – 13, 2014.

6

7 YAMAKAGE, M., NAMIKI, A. Calcium channels basic aspects of their structure,  
8 function and gene encoding, anesthetic action on the channels - A review.  
9 **Canadian Journal of Anesthesia**. v. 49, n. 02, p. 151 – 164, 2002.

10

11 YI, Y., SONG, Y., LU, Y. Parvalbumin Interneuron Activation-Dependent Adult  
12 Hippocampal Neurogenesis Is Required for Treadmill Running to Reverse  
13 Schizophrenia-Like Phenotypes. **Frontiers in Cell and Developmental Biology**,  
14 v. 8, n. 24, 2020.

15

16 ZALESKI, M., LARANJEIRA, R.R., MARQUES, A.C.P.R., RATTO, L., ROMANO,  
17 M., ALVES, H.N.P., SOARES, M.B.M., ABELARDINO, V., KESSLER, F.,  
18 BRASILIANO, S. NICASTRI, S., HOCHGRAF, P.B., GIGLIOTTI, A.deP.,  
19 LEMOS, T. Guidelines of the Brazilian Association of Studies on Alcohol and  
20 Other Drugs (ABEAD) for diagnosis and treatment of psychiatric comorbidity with  
21 alcohol and other substance and dependence. **International Review of**  
22 **Psychiatry**. v. 29, n. 03, p. 254 – 262, 2017.

23

24

25